



**Eddleston Water Restoration Project  
Macroinvertebrate responses 2012-2019**

**Tweed Forum**

**APEM Ref: P0002209**

**June 2020**



**Client:** Tweed Forum  
**Address:** South Court  
Drygrange Steading  
Melrose  
Roxburghshire  
TD6 9DJ  
**Project reference:** P00002209  
**Date of issue:** 27 June 2020

---

**Project Director:** Dr David Bradley  
**Project Manager:** Dr Andrew Davey  
**Laboratory Manager:** Sally Donaldson

---

APEM Ltd  
Riverview  
A17 Embankment Business Park  
Heaton Mersey  
Stockport  
SK4 3GN

Tel: 0161 442 8938  
Fax: 0161 432 6083

Registered in England No. 02530851

Report should be cited as:

“APEM (2020). Eddleston Water Restoration Project: Macroinvertebrate responses 2012-2019. APEM Scientific Report P00002009. Tweed Foundation, June 2020, v3.0 Final Report, 32 pp.”



## Revision and Amendment Register

Version Number	Date	Sections	Pages	Summary of Changes	Approved by
1.0	09/04/2020	1	25	Interim report with summary of main findings	D. Bradley
2.0	11/06/2020	4	28	Draft final report	D. Bradley
3.0	30/06/2020	4	32	Final report addressing comments from J. Dodd.	D. Bradley

## Contents

Executive Summary .....	1
1. Introduction and Methods.....	2
1.1 The Eddleston Water Project .....	2
1.2 Experimental design .....	2
1.3 Field sampling.....	3
1.4 Laboratory sample analysis .....	4
1.5 Data analysis .....	4
2. Results.....	6
2.1 How do the sites differ in mesohabitat composition, and how has this changed over time? 6	
2.2 How do the sites differ in macroinvertebrate community composition, and how has this changed over time? .....	7
2.3 How does macroinvertebrate community composition vary by habitat and site? .....	14
3. Discussion .....	18
3.1 Conclusions .....	18
3.2 Lessons learnt .....	19
3.3 Recommendations.....	20
4. References .....	21
<b>Appendix 1</b> Summary of sampling programme.....	22
<b>Appendix 2</b> Results of statistical analyses comparing reach-level biotic indices among sites and time periods .....	23
WHPT_ASPT.....	23
Family level LIFE .....	24
Species level LIFE .....	25
Family level PSI.....	26
Species level PSI.....	27
Taxon richness .....	28
Total abundance.....	29
CCI .....	30
% EPT .....	31

% Oligochaetes and chironomids.....	32
-------------------------------------	----

## List of Figures

Figure 1.1 Schematic diagram of the experimental design showing the treatment (red circles) and control (blue circles) sites where macroinvertebrate samples were collected .....	3
Figure 2.1 Changes in proportional mesohabitat composition 2012-2019, based on the allocation of kicks to habitat units. Note: habitat re-configuration was completed on 25/07/2013 at Cringletie and on 11/09/2013 at Lake Wood.....	6
Figure 2.2 Change in WHPT-ASPT score at the four sampling locations before, following and after channel reconfiguration.....	9
Figure 2.3 Change in family-level LIFE score at the four sampling locations before, following and after channel reconfiguration.....	9
Figure 2.4 Change in species-level LIFE score at the four sampling locations before, following and after channel reconfiguration.....	10
Figure 2.5 Change in family-level PSI score at the four sampling locations before, following and after channel reconfiguration.....	10
Figure 2.6 Change in species-level PSI score at the four sampling locations before, following and after channel reconfiguration.....	11
Figure 2.7 Change in taxon richness at the four sampling locations before, following and after channel reconfiguration.....	11
Figure 2.8 Change in total abundance at the four sampling locations before, following and after channel reconfiguration.....	12
Figure 2.9 Change in Community Conservation Index at the four sampling locations before, following and after channel reconfiguration.....	12
Figure 2.10 Change in % Ephemeroptera, Plecoptera and Trichoptera at the four sampling locations before, following and after channel reconfiguration .....	13
Figure 2.11 Change in % Oligochaeta and Chironomidae at the four sampling locations before, following and after channel reconfiguration .....	13
Figure 2.12 Boxplot comparison of WHPT-ASPT by mesohabitat type and site .....	14
Figure 2.13 Boxplot comparison of species-level LIFE by mesohabitat type and site .....	15
Figure 2.14 Boxplot comparison of species-level PSI by mesohabitat type and site.....	15
Figure 2.15 Boxplot comparison of taxon richness by mesohabitat type and site .....	16
Figure 2.16 Boxplot comparison of macroinvertebrate abundance by mesohabitat type and site. Note that abundance has been standardised by the number of kicks in each mesohabitat type to control for differences in sampling effort. ....	16

Figure 2.17 Boxplot comparison of Community Conservation Index by mesohabitat type and site ..... 17

Figure 2.18 Boxplot comparison of % Ephemeroptera, Plecoptera and Trichoptera by mesohabitat type and site ..... 17

Figure 2.19 Boxplot comparison of % Oligochaeta and Chironomidae by mesohabitat type and site ..... 18

## List of Tables

Table 1.1 Macroinvertebrate biotic indices analysed in this study .....4

## Executive Summary

The Eddleston Water Project is a major national research project that aims to generate robust evidence of the impact, cost and benefits of working with natural processes to deliver Natural Flood Management (NFM) at multiple scales from the location of the individual measures, through to the cumulative effect at the catchment scale.

This report evaluates the impact of channel reconfiguration on the benthic macroinvertebrate community in Eddleston Water using data from a bespoke Before-After-Control-Impact (BACI) monitoring framework. Monitoring was conducted before (2012 to summer 2013), immediately following (autumn 2013 to 2015) and after (2016, 2017 and 2019) channel reconfiguration at two 'impact' sites (Lake Wood and Cringletie), and also at two 'control' sites: one (Signal Cottage) located upstream and one located (Rosetta) downstream of the impact sites. Macroinvertebrate community composition was measured at the mesohabitat-scale and reach-scale using a suite of ten biotic metrics (WHPT-ASPT; family- and species-level LIFE; family- and species-level PSI; taxon richness; total abundance; CCI; % of total abundance of Ephemeroptera, Plecoptera and Trichoptera (%EPT); and % of total abundance of oligochaetes and chironomids (%OligoChiro)). Changes in response to the channel reconfiguration work were interpreted in terms of changes in mesohabitat composition at the four sites.

The key results are as summarised below.

1. Prior to channel reconfiguration, the two impact sites – Lake Wood and Cringletie – had much less riffle/run and more glide habitat than the two control sites and had lower values than the two control sites for seven out of the eight biotic metrics (exception was %EPT).
2. Channel reconfiguration in 2013 initially increased the proportion of riffle and run habitat and increased overall habitat diversity, but subsequent geomorphological adjustment appears to have partially reversed these changes.
3. Against a background of rapidly increasing taxon richness at all sites, channel reconfiguration caused an abrupt shift in macroinvertebrate community composition at the impact sites from one dominated numerically by mayflies, stoneflies and caddisflies to one dominated by oligochaetes and chironomids.
4. Following the initial disturbance caused by the channel reconfiguration work, the impact and control sites have partially converged in macroinvertebrate composition but only total abundance and the %OligoChiro have increased significantly as a result of the intervention.
5. Six years after the channel reconfiguration work, five of the biotic indices (WHPT-ASPT, LIFE-species, PSI-family, PSI-species, and %EPT) remain significantly lower at the impact sites compared with the control sites.

In conclusion, the benthic macroinvertebrate community in Eddleston Water appears to be strongly influenced by mesohabitat composition. Channel reconfiguration has led to a partial improvement in macroinvertebrate community status (as measured by a variety of standard biotic indices) but full recovery from historical channel straightening is thought to have been constrained to date by the limited geomorphological changes at Lake Wood and Cringletie.



## 1. Introduction and Methods

### 1.1 The Eddleston Water Project

The Eddleston Water Project is a major national research project that aims to generate robust evidence of the impact, cost and benefits of working with natural processes to deliver Natural Flood Management (NFM) at multiple scales from the location of the individual measures, through to the cumulative effect at the catchment scale. Delivering NFM and environmental benefits from the restoration of natural habitats will expand the current knowledge base and aim to demonstrate the multiple benefits possible from NFM.

The three main aims of the Eddleston Water Project are:

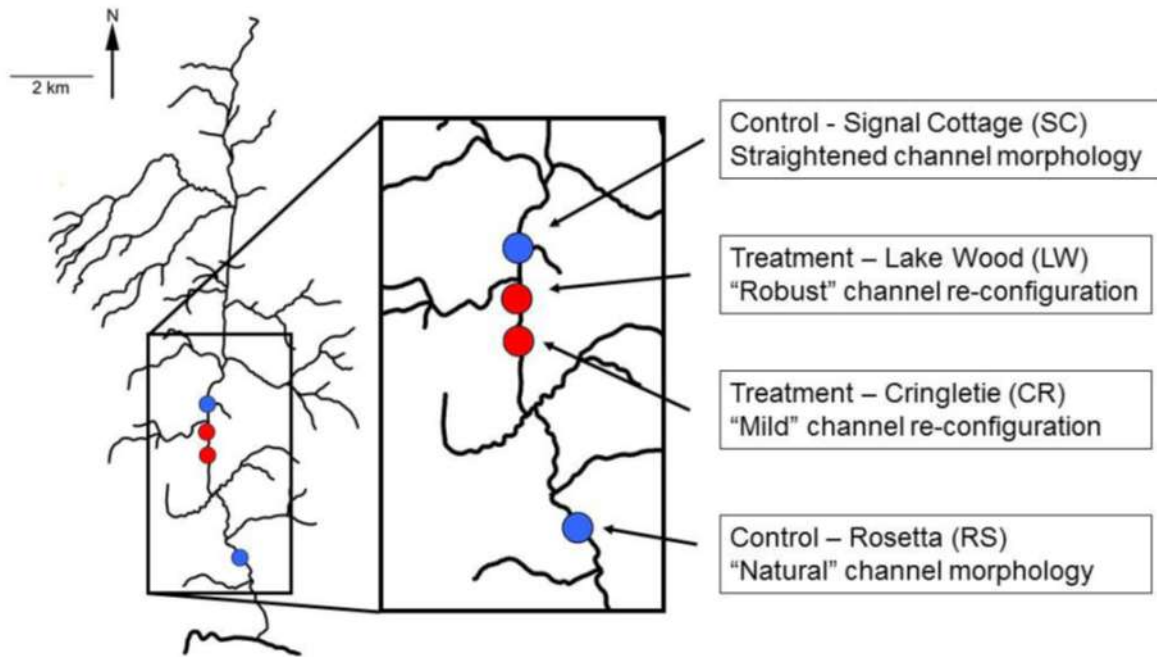
1. to investigate the possibility of reducing the risk of flooding to the communities of Eddleston and Peebles by restoring some of the original natural features of the river, its flood plain and surrounding hill slopes;
2. to examine the potential for added benefits for wildlife and fisheries through improvements to river habitats; and
3. to work with landowners and communities in the Eddleston valley to maximise the benefits they would gain from such work, whilst maintaining the profitability of local farms.

In order to address the second aim, a monitoring strategy has been established to assess the effects of NFM measures on macroinvertebrates, macrophytes, fishes and geomorphology. Building on earlier studies (Veritas Ecology 2017; APEM 2018), this report evaluates the impact of channel reconfiguration on the benthic macroinvertebrate community in Eddleston Water using data from a bespoke Before-After-Control-Impact (BACI) monitoring framework (Feld et al. 2011). Mesohabitat- and reach-level macroinvertebrate sample data from control and impact sites were analysed to:

- describe the spatial and temporal patterns in macroinvertebrate composition;
- explore how these patterns are related to mesohabitat composition; and
- evaluate the local impact on macroinvertebrates of channel reconfiguration.

### 1.2 Experimental design

Eddleston Water is a tributary of the River Tweed. Monitoring was conducted before (2012 to summer 2013), immediately following (autumn 2013 to 2015) and after (2016, 2017 and 2019) channel reconfiguration at two 'impact' sites (Lake Wood and Cringletie), and also at two 'control' sites: one (Signal Cottage) located upstream and one located (Rosetta) downstream of the impact sites. Figure 1.1 shows the macroinvertebrate sampling locations.



**Figure 1.1 Schematic diagram of the experimental design showing the treatment (red circles) and control (blue circles) sites where macroinvertebrate samples were collected**

At the impact sites, a new meandering channel was excavated, and the old channel was filled in. The details of the channel reconfiguration work differed slightly at the two sites.

1. The new channel at Lake Wood was much more sinuous than the new channel in Cringletie.
2. No substrate material was transferred from the old channel to the new at Lake Wood, as there was ample diversity of substrate encountered where the new channel was excavated. In contrast, at Cringletie, the material that was excavated to create the new channel was very soft and homogenous, and so material was taken from the old bed and laid in the new channel to create some harder patches of riffles.

Despite these differences, the two locations treated as replicate impact sites for evaluation purposes. The works were completed on 25/07/2013 at Cringletie and on 11/09/2013 at Lake Wood. Similarly, Signal Cottage and Rosetta were treated as replicate control sites despite having contrasting straightened and natural morphologies.

### 1.3 Field sampling

Macroinvertebrate samples were collected by SEPA in spring (all years), summer (2012-2014 only) autumn (all years except 2012). All samples were collected by the same operator for the duration of the project to minimise operator variability. Samples were collected using a modified version of the kick/sweep sampling method used by UK government agencies for monitoring under the EU Water Framework Directive (Environment Agency, 2017). A total of 20 kick samples were taken at each site, split proportionately between five mesohabitat types (riffle, run, glide, pool and slack). Three replicates were collected from each mesohabitat type on each sampling date. The exception was 2012, when a single three-

minute kick/sweep sample was taken from each site on each sampling date. An overview of the sampling programme is provided in Appendix 1.

#### 1.4 Laboratory sample analysis

Aquatic macroinvertebrate samples were analysed in accordance with the requirements outlined in SEPA's for mixed taxon level analysis (ES-Ecol-p-021). Samples were initially washed inside a fume cupboard using a 500µm sieve to remove fine silt and preservative. Samples were subsequently sorted to Mixed Taxon Level 5 (TL5) in accordance with SEPA Procedure ES-Ecol-G-007. Actual abundances were recorded rather than logarithmic abundances for counts up to 100. If abundances were greater than 100 then estimates of abundance will be calculated by multiplying the abundance within one quadrat by the number of quadrats within one sorting tray.

#### 1.5 Data analysis

The 20 individual kick samples taken on each sampling occasion were aggregated to produce a single sample for each mesohabitat type ('habitat-scale' samples). These were then aggregated again to produce a single 'reach-scale' sample.

Macroinvertebrate community composition was measured at the mesohabitat-scale and reach-scale using a suite of eight biotic indices, detailed in Table 1.1.

**Table 1.1 Macroinvertebrate biotic indices analysed in this study**

Index	Description
WHPT_ASPT	Whalley-Hawkes-Paisley-Trigg Average Score Per Taxon – an index of overall biological quality using macroinvertebrate families (Paisley, Whalley & Trigg, 2013).
LIFE_F	Lotic-invertebrate Index for Flow Evaluation – indexes the effect of flow variations on macroinvertebrate communities (Extence, Balbi & Chadd, 1999). Calculated at species level.
LIFE_S	Lotic-invertebrate Index for Flow Evaluation – indexes the effect of flow variations on macroinvertebrate communities (Extence, Balbi & Chadd, 1999). Calculated at species level.
PSI_F	Proportion of Sediment-sensitive Invertebrates – describes the degree to which river sites are impacted by fine sediment (Extence et al. 2011). Calculated at family level.
PSI_S	Proportion of Sediment-sensitive Invertebrates – describes the degree to which river sites are impacted by fine sediment (Extence et al. 2011). Calculated at species level.
Taxon richness	Count of the number of distinct taxa identified in the sample.
Total abundance	Estimated total number of macroinvertebrates in the sample.
CCI	Community Conservation Index – provides a standardised measure of the conservation value of macroinvertebrate communities at a site which can be compared across sites throughout Great Britain. CCI reflects both the rarity of the species found within each sample and the overall diversity of the sample. (Chadd & Extence, 2004).
%EPT	Ephemeroptera, Plecoptera and Trichoptera as a % of total abundance –

	indicates the relative abundance of three environmentally sensitive macroinvertebrate groups.
%OligoChiro	Oligochaeta and Chironomidae as a % of total abundance – indicates the relative abundance of two environmentally insensitive macroinvertebrate groups.

The effect of the channel reconfiguration work on each biotic index at a reach scale was evaluated using a mixed-effects regression model to test for a statistically significant ( $\alpha=0.05$ ) interaction between treatment (Control or Impact) and time period (Before, Following and After channel reconfiguration). Time (year and month of sampling) and Site were included as crossed random effect to measure and account for the year-to-year and site-to-site variability. The structure of the model reflected the BACI sampling design and was designed to test the null hypothesis that the temporal patterns at the control and impact sites were the same; consequently, no model simplification was carried out. Diagnostic plots were examined to verify that the assumptions of homogeneous variance and independent, normally distributed errors.

The results were interpreted in terms of changes in mesohabitat composition at the four sites, and by comparing the habitat-level indices among sites.

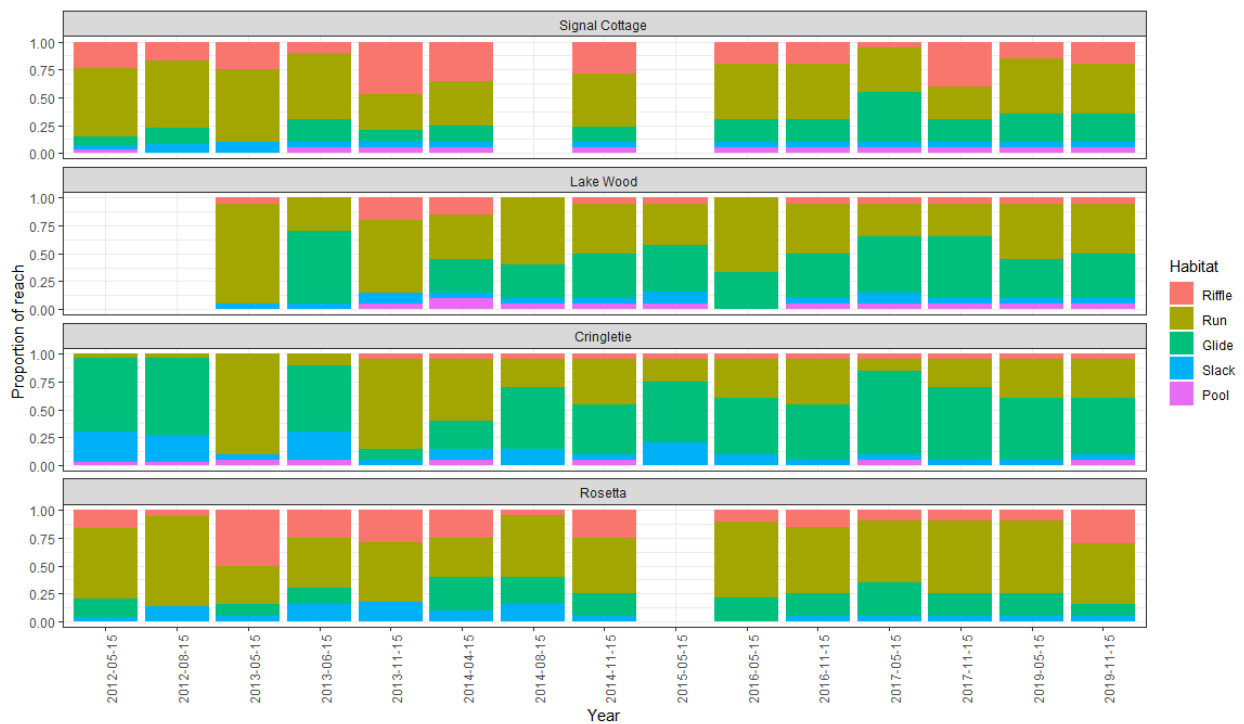
All statistical analyses were performed using R v3.6.1 (R Core development Team 2020).

## 2. Results

### 2.1 How do the sites differ in mesohabitat composition, and how has this changed over time?

Figure 2.1 plots the proportion of riffle, run, glide, slack and pool at each of the four sites over the 2012-2019 study period. The two control sites – Signal Cottage and Rosetta – both had predominantly riffle and run habitat, despite having contrasting straightened and natural morphologies. Mesohabitat composition fluctuated slightly over time, with a slight trend towards more glide habitat at Signal Cottage.

By contrast, the two impact sites – Lake Wood and Cringletie – had a majority of glide and slack habitat prior to restoration. Immediately following channel reconfiguration, the proportion of riffle and run habitat at Lake Wood increased from 30% to 85% but then reduced to ca. 50% over the next two years. Similarly, channel reconfiguration initially increased the proportion of run habitat at Cringletie from 10% to 85% and created some new riffles (5%), but over the next two years the proportion of run and riffle reduced back to ca. 40%.



**Figure 2.1 Changes in proportional mesohabitat composition 2012-2019, based on the allocation of kicks to habitat units. Note: habitat re-configuration was completed on 25/07/2013 at Cringletie and on 11/09/2013 at Lake Wood.**

## 2.2 How do the sites differ in macroinvertebrate community composition, and how has this changed over time?

Figure 2.2 to Figure 2.11 illustrate the changes in macroinvertebrate community composition at the four sampling locations before, following and after channel reconfiguration. Detailed statistical comparison of sites and time periods is presented in Appendix 2.

With the exception of WHPT\_ASPT, which was significantly higher in autumn and lower in summer, none of the metrics displayed clear seasonal differences.

In the Before period, prior to channel reconfiguration, nine of the ten biotic indices (the exception was %EPT) were lower at the Lake Wood and Cringletie impact sites than at the two control sites. However, the limited amount of baseline (pre-intervention) data limited statistical power to detect any impact of historical channel straightening, and only LIFE\_F, LIFE\_S and PSI\_F showed a statistically significant difference between impact and control sites.

Over the entire (2012-2019) monitoring period, there was a strong and highly significant ( $p < 0.001$ ) increase in **taxon richness** at all sites, from a mean of ca. 25 per site in 2012 to a mean of over 60 per site by 2019 (Figure 2.7). Mirroring this, there was also a less pronounced but statistically significant ( $p = 0.001$ ) increase in **WHPT-ASPT** (Figure 2.2) and **CCI** (Figure 2.9). The reason for this underlying trend is not known; Eddleston Water has been consistently classed as High status for water quality and Good for macroinvertebrates under the Water Framework Directive (WFD), so there is no evidence that the river is recovering from historical water pollution. Nonetheless, there were no statistically significant differences between control and impact locations and all four sites exhibited similar trends, so there was no evidence that channel reconfiguration affected these three indices.

Prior to the channel reconfiguration work, mayflies, stoneflies and caddisflies (**%EPT**) accounted for ca. 50% of total abundance across all sites and there was no significant difference between the control and impact locations. Immediately following channel reconfiguration, %EPT at Lake Wood and Cringletie fell to 5% ( $p < 0.001$ ) but by autumn 2014 %EPT has increased again and was similar to pre-work levels (Figure 2.10). Between 2015 and 2019 there was a small (non-significant) increase in %EPT at the two control sites (relative to the before period) but this was not mirrored at the two impact sites. As a result, %EPT at the impact sites decreased relative to the control sites during the After period ( $p < 0.01$ ; Figure 2.10). Exactly the opposite pattern was observed for **%OligoChiro** (Figure 2.11). Thus, channel reconfiguration caused an abrupt shift in macroinvertebrate community composition from one dominated numerically by mayflies, stoneflies and caddisflies to one dominated by oligochaetes and chironomids, which are rapid colonisers of newly created or freshly disturbed substrates. As a consequence, **total abundance** at the two impact sites increased ( $p < 0.001$ ) by 153% between the Before and Following periods (relative to the controls) and remained 50% higher in the After period (Figure 2.8).

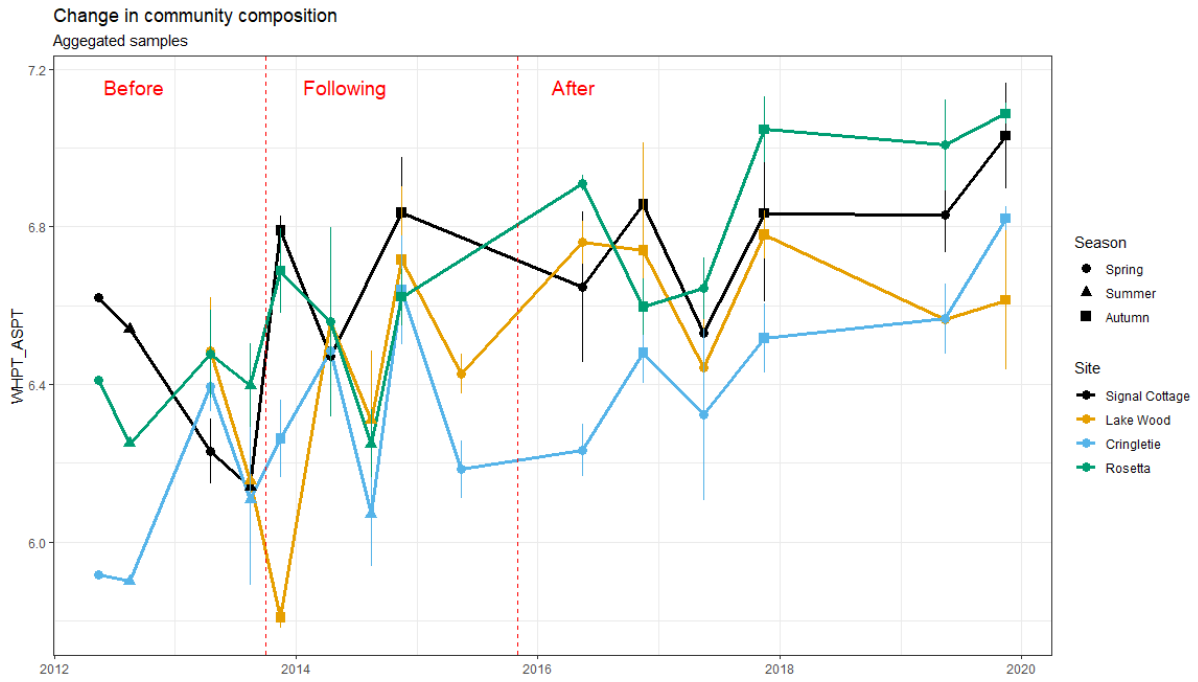
Prior to channel reconfiguration, **LIFE\_S** was on average 0.19 lower at the impact sites than at the control sites ( $p = 0.004$ ), which is consistent with the lower proportion of riffle and run habitat at Lake Wood and Cringletie (Figure 2.3). Following channel reconfiguration, mean LIFE\_S decreased sharply at the two control sites ( $P < 0.001$ ). LIFE\_S also decreased at the two impact sites, but by a smaller ( $p = 0.04$ ) amount, suggesting that the intervention had the effect of holding up LIFE\_S scores at Lake Wood and Cringletie. In the After period, LIFE\_S at the two control sites recovered and in the most recent year of sampling (2019), LIFE\_S

was again 0.23 lower at the impact sites than at the control sites (Figure 2.3). A similar pattern was observed for **LIFE\_F**. However, the reduction in LIFE\_F was more prolonged at the control sites and minimal at the impact sites, and so the results had a higher level of statistical significance.

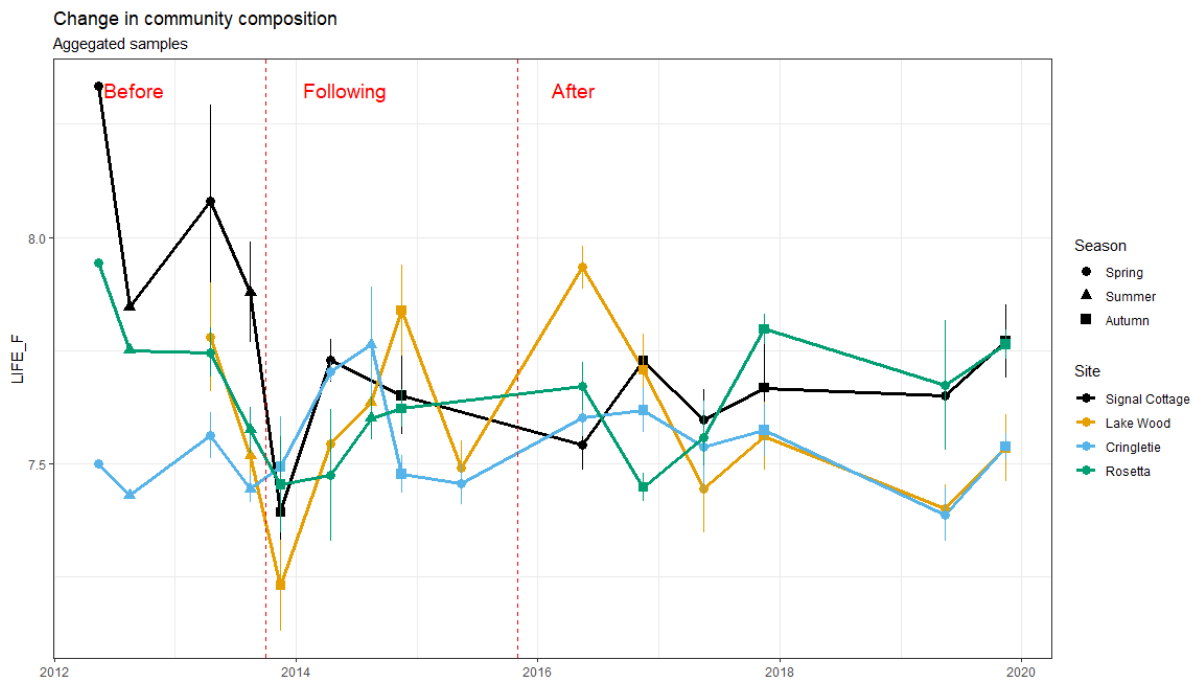
Finally, **PSI\_S** was marginally lower (3 percentage points) at the impact sites than at the control sites during the Before period ( $p = 0.07$ ). In the After period, PSI\_S increased by 5.7 percentage points at the control sites ( $p = 0.008$ ) but only by 2.0 percentage points at the impacts sites (Figure 2.5); the different responses at the control and impacts sites were marginally non-significant ( $p = 0.06$ ). **PSI\_F** was also lower (4 percentage points) at the impact sites than at the control sites during the Before period ( $p = 0.01$ ), but there was no evidence that channel reconfiguration altered PSI\_F at the impact sites.

In the After period (2016-2019), there was no significant difference between impact and control sites in taxon richness, total abundance, CCI or %OligoChiro, but WHPT-ASPT, LIFE\_S, PSI\_S and %EPT were all significantly lower at the impact sites than at the control sites.

As well as testing for an effect of channel reconfiguration, the models were also able to partition the unexplained variation in the data into component sources of error (see the 'Random effects' in the model outputs in Appendix 2). Without exception, the single largest source of unexplained variation in every model was **residual variation**; this comprises both measurement error (i.e. the random variability observed among replicate samples) *and* site-specific trends (i.e. temporal changes in mean metrics scores at individual sites that cannot be explained by season, time period, or year/month of sampling). The variance attributable to **time** (the variability in mean metric scores from sampling visit to another) was at least an order of magnitude lower than the residual variance for WHPT-ASPT, LIFE, PSI and CCI. For the other metrics, the time variance was 1.5 to 2 times smaller than the residual variance. Finally, the variance attributable to **site** (i.e. the variability in mean metric scores among replicate control and impact sites) was universally very small, and sometimes estimated to be 0. Thus, after accounting for the fixed effects in the model, there appears to be relatively little temporal variability that is consistent across all sites and little spatial variability that is consistent over time. Instead, each site exhibits its own idiosyncratic temporal behaviour, and measurement error is appreciable despite the same ecologist collecting all the samples (i.e. no inter-operator variability) and using a well-established sampling protocol.

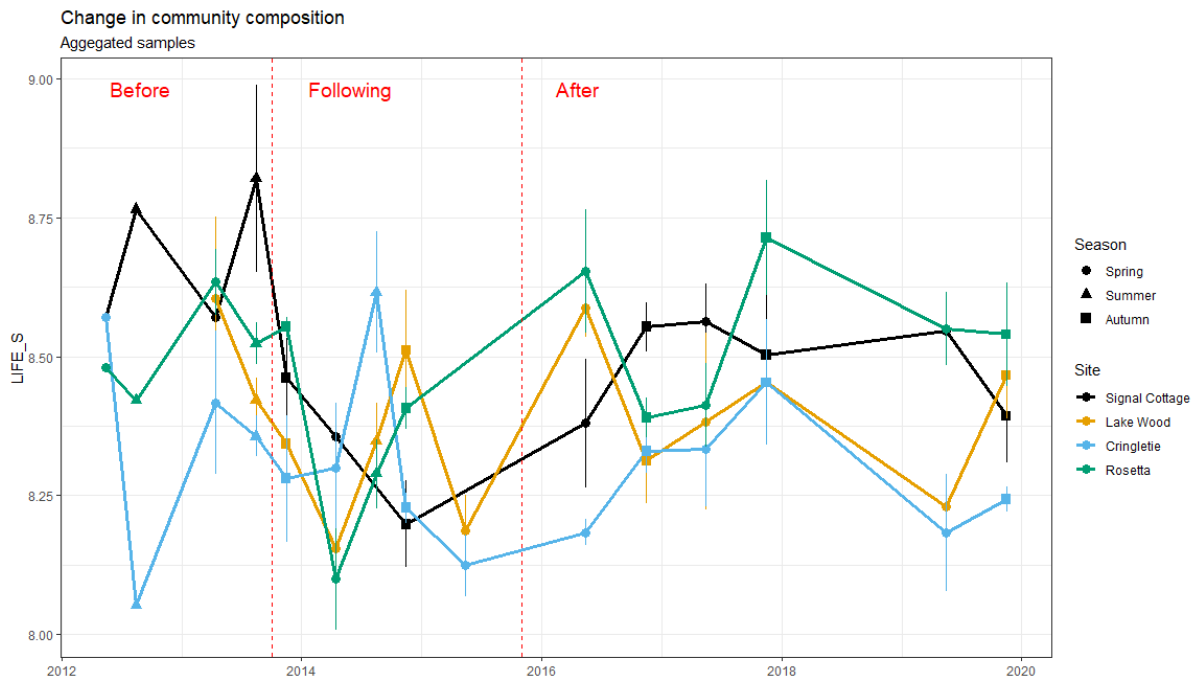


**Figure 2.2 Change in WHPT-ASPT score at the four sampling locations before, following and after channel reconfiguration**

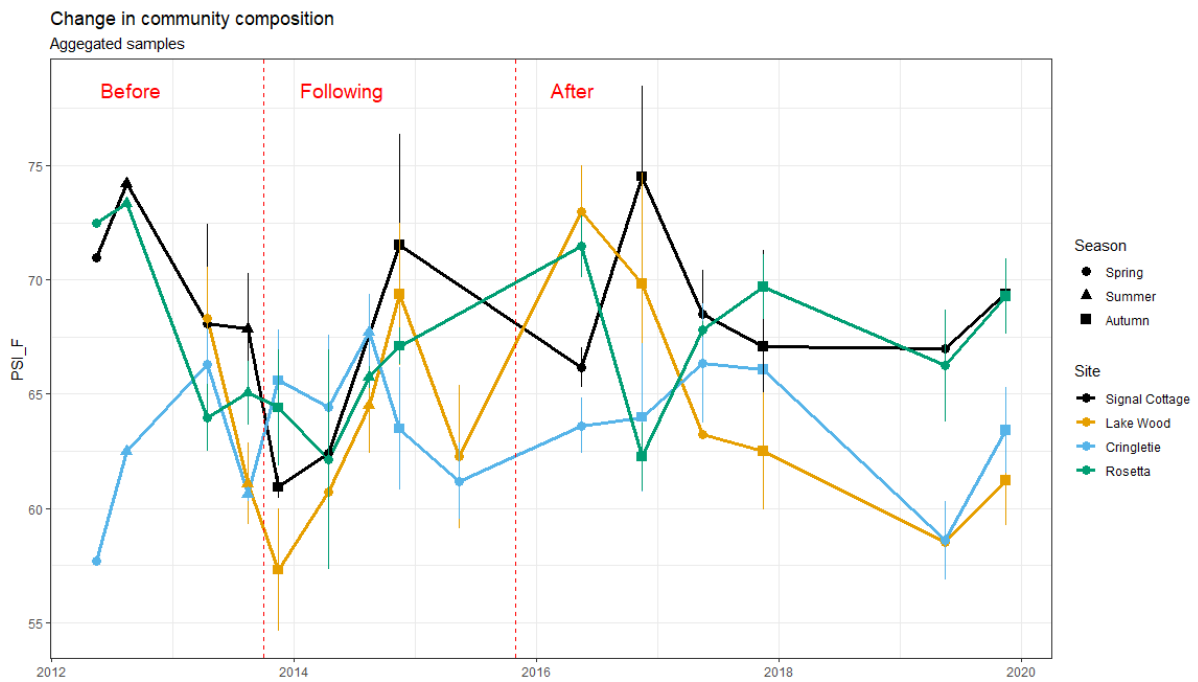


**Figure 2.3 Change in family-level LIFE score at the four sampling locations before, following and after channel reconfiguration**

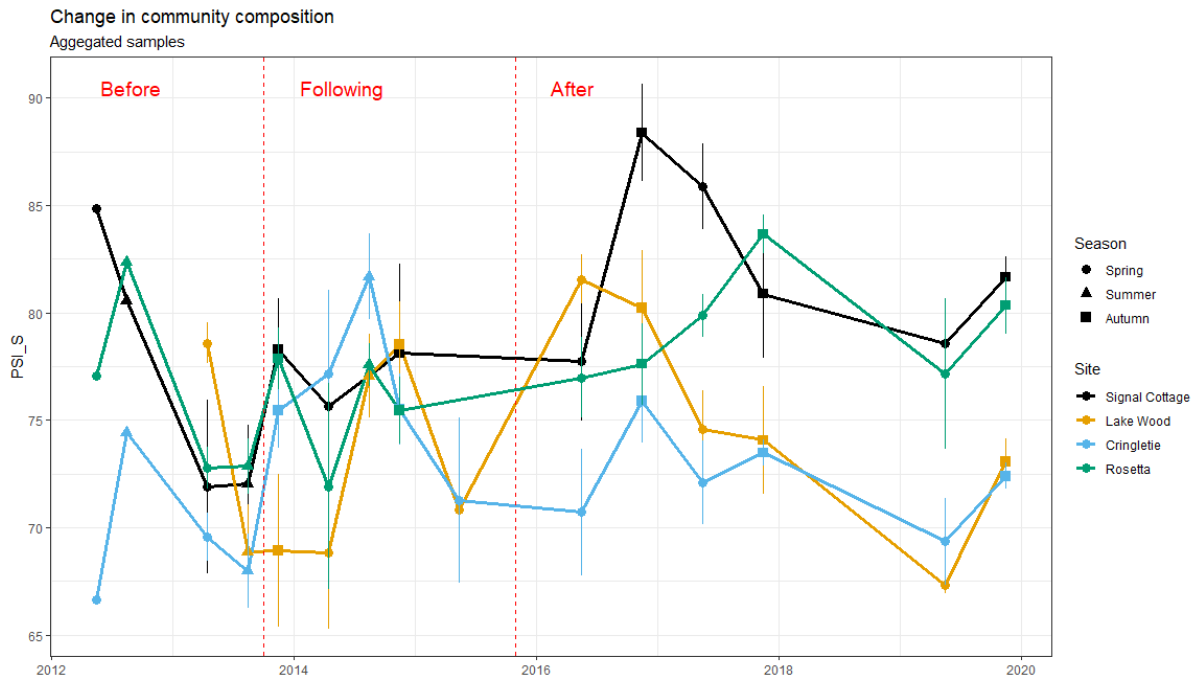




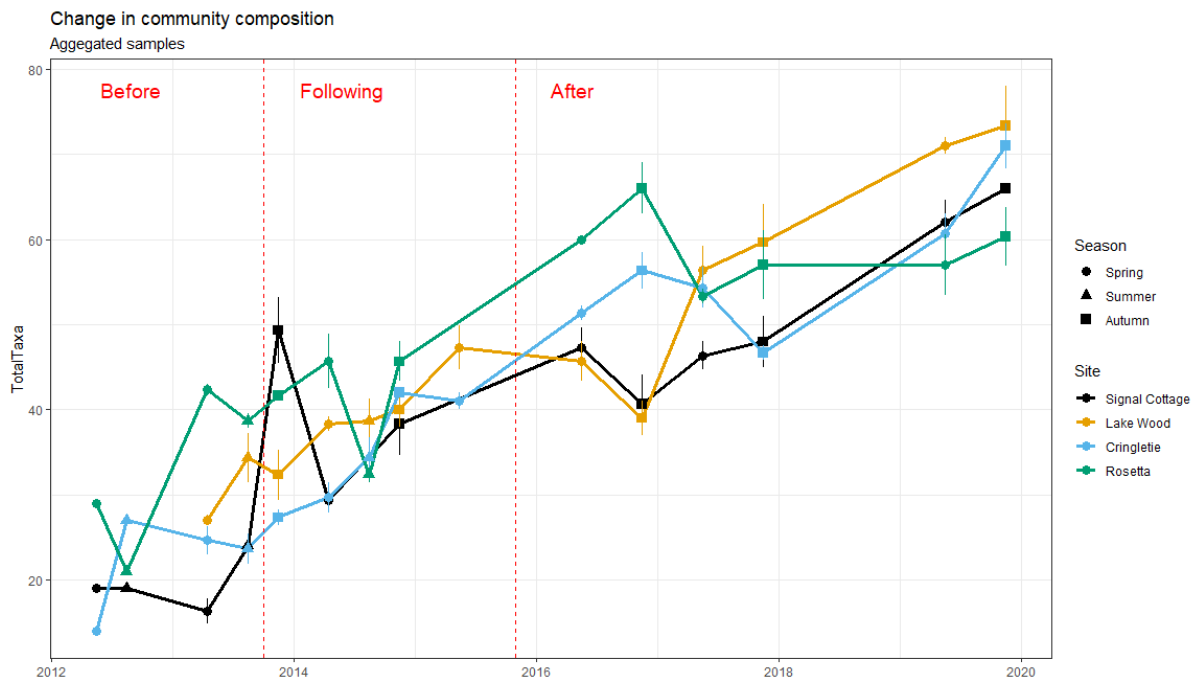
**Figure 2.4 Change in species-level LIFE score at the four sampling locations before, following and after channel reconfiguration**



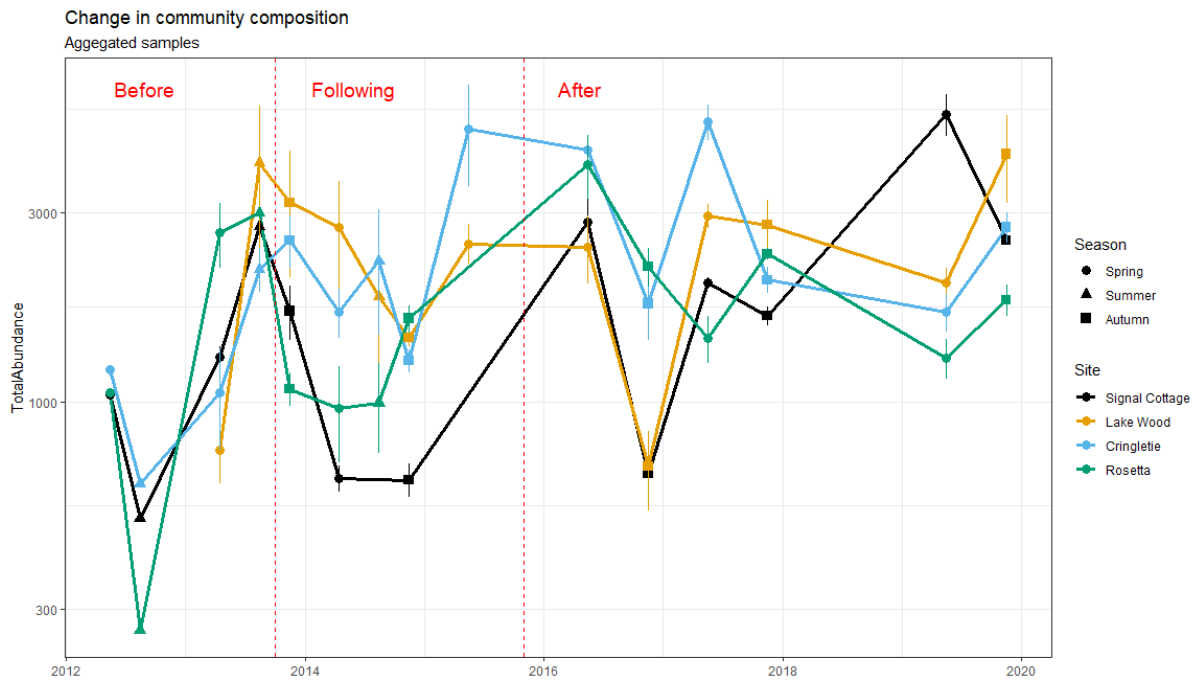
**Figure 2.5 Change in family-level PSI score at the four sampling locations before, following and after channel reconfiguration**



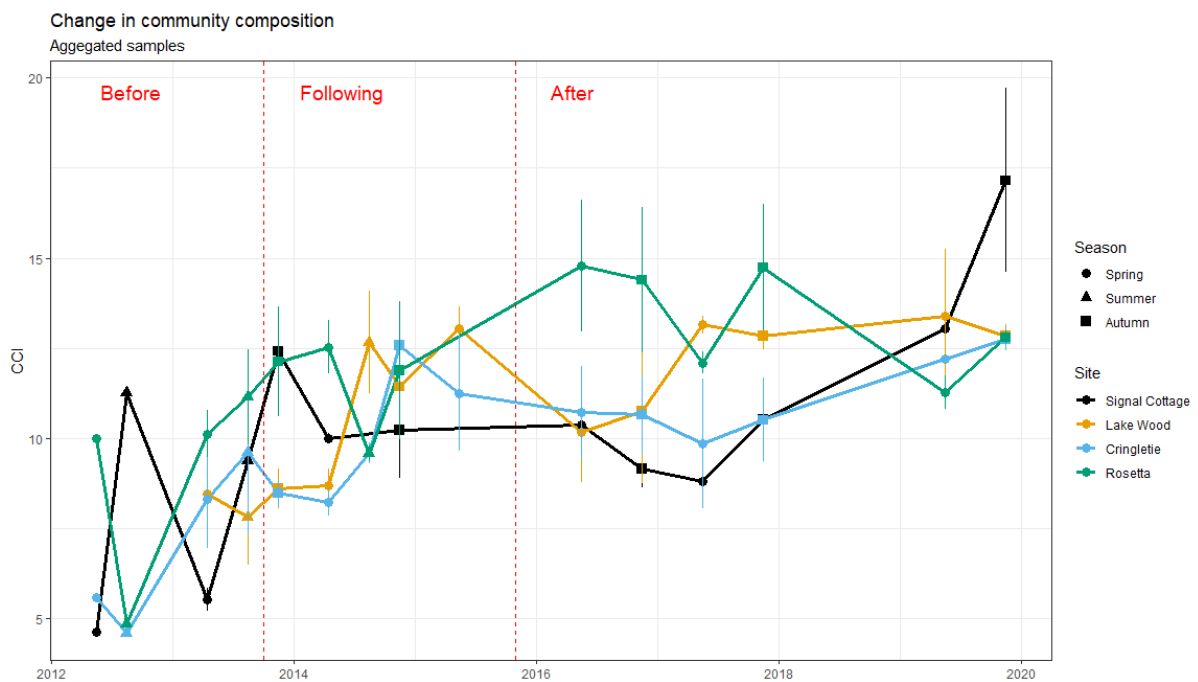
**Figure 2.6 Change in species-level PSI score at the four sampling locations before, following and after channel reconfiguration**



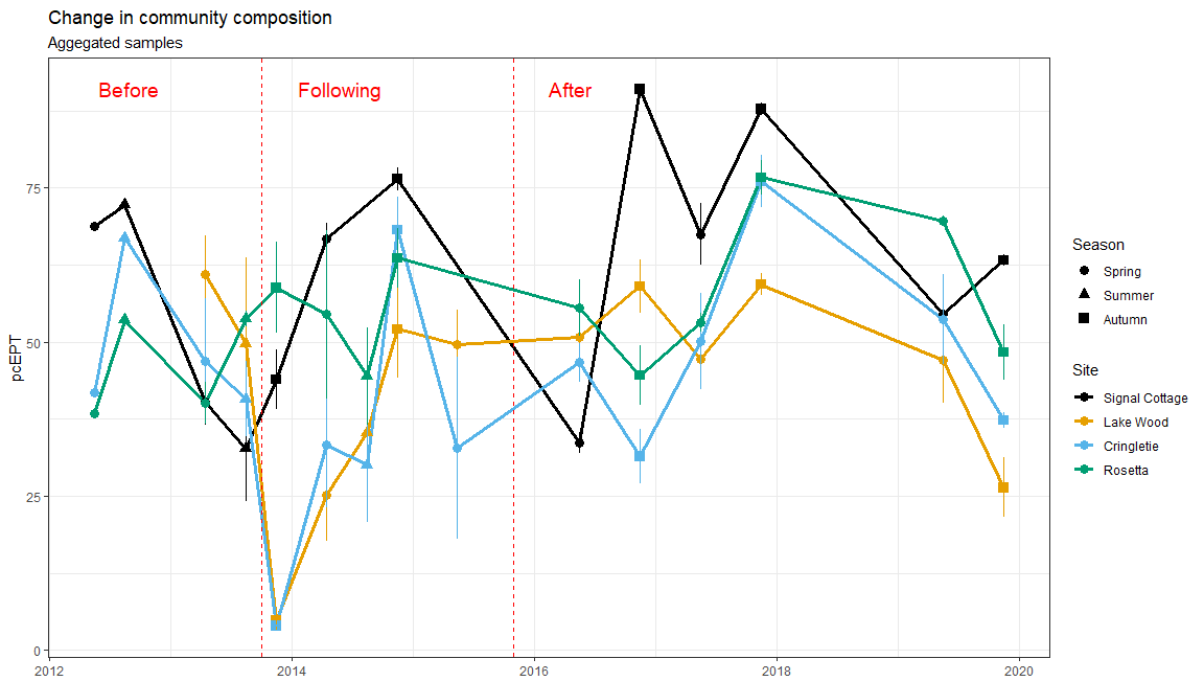
**Figure 2.7 Change in taxon richness at the four sampling locations before, following and after channel reconfiguration**



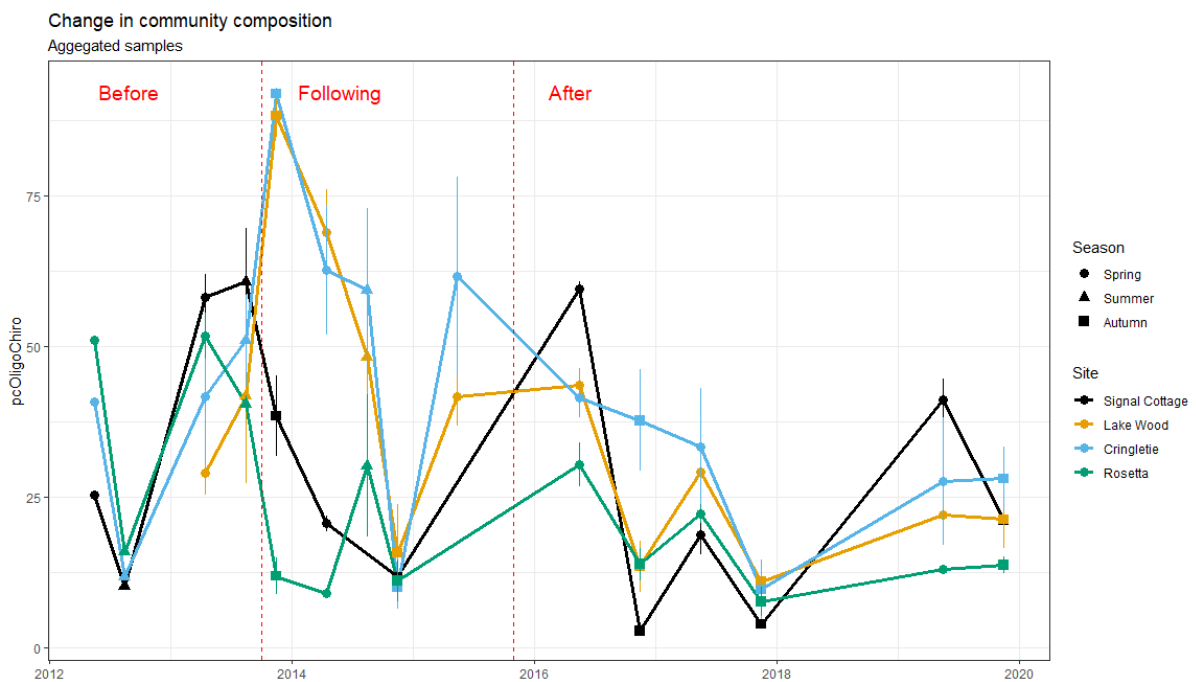
**Figure 2.8 Change in total abundance at the four sampling locations before, following and after channel reconfiguration**



**Figure 2.9 Change in Community Conservation Index at the four sampling locations before, following and after channel reconfiguration**



**Figure 2.10 Change in % Ephemeroptera, Plecoptera and Trichoptera at the four sampling locations before, following and after channel reconfiguration**



**Figure 2.11 Change in % Oligochaeta and Chironomidae at the four sampling locations before, following and after channel reconfiguration**

### 2.3 How does macroinvertebrate community composition vary by habitat and site?

Figure 2.12 to Figure 2.19 compare macroinvertebrate community composition at the four sites at the mesohabitat level. Samples collected before, following and after channel reconfiguration are plotted together.

All eight biotic indices showed large differences between habitat types. WHPT-ASPT, LIFE\_S, PSI\_S and %EPT were all highest in riffles and runs and lowest in slacks; the opposite pattern was true for %OligoChiro (i.e. lowest in riffles and runs and highest in slacks). Taxon richness and CCI were highest in runs and glides and lowest in pools and slacks. Finally, abundance was similar across all five habitat types after controlling for differences in sampling effort (i.e. macroinvertebrate density was similar).

Differences between the mesohabitat types were reasonably consistent across the four sites, but with a few exceptions. Notably, Signal Cottage had consistently higher WHPT-ASPT, PSI\_S and %EPT and consistently lower %OligoChiro in slacks and pools than at the other three sites. Rosetta had consistently higher taxon richness and CCI in riffles and runs than at the other sites, but the lowest CCI in glides. It is likely that these differences reflect persistent local differences in morphology between the sites.

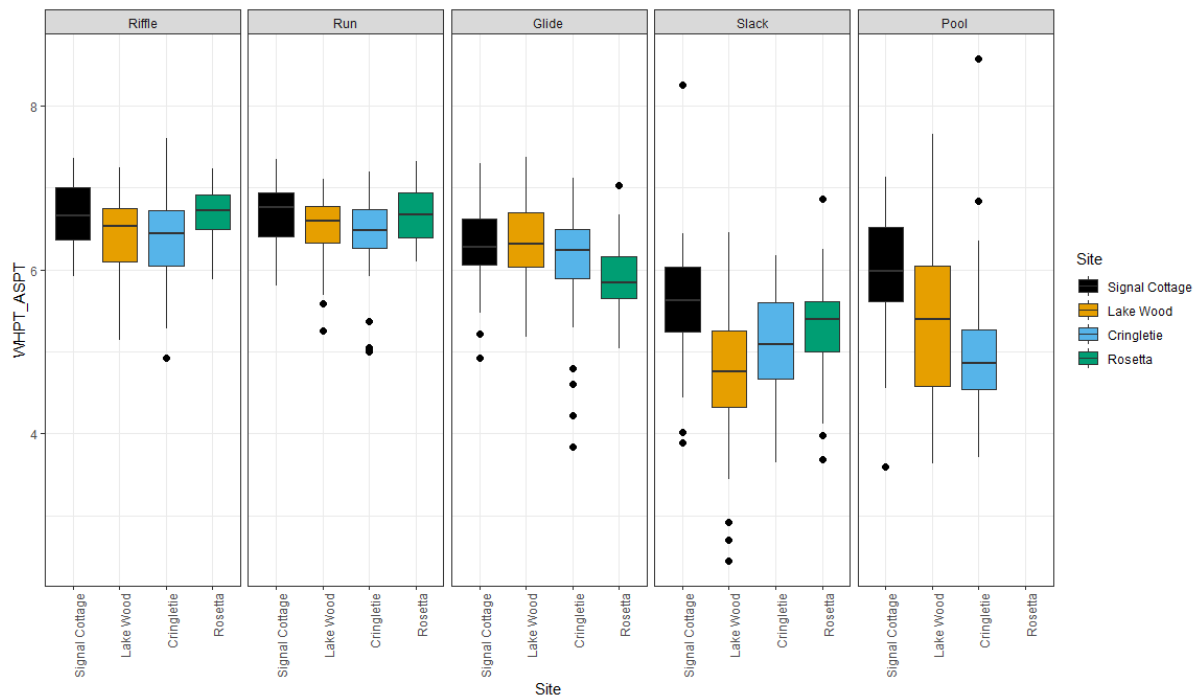


Figure 2.12 Boxplot comparison of WHPT-ASPT by mesohabitat type and site

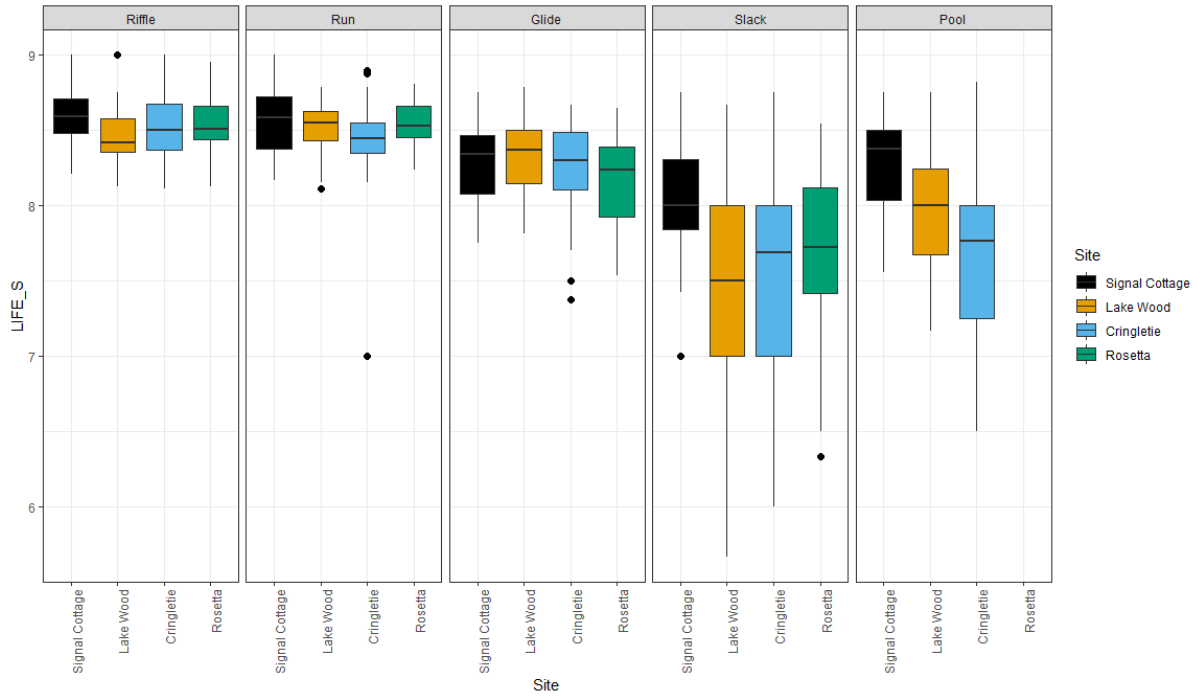


Figure 2.13 Boxplot comparison of species-level LIFE by mesohabitat type and site

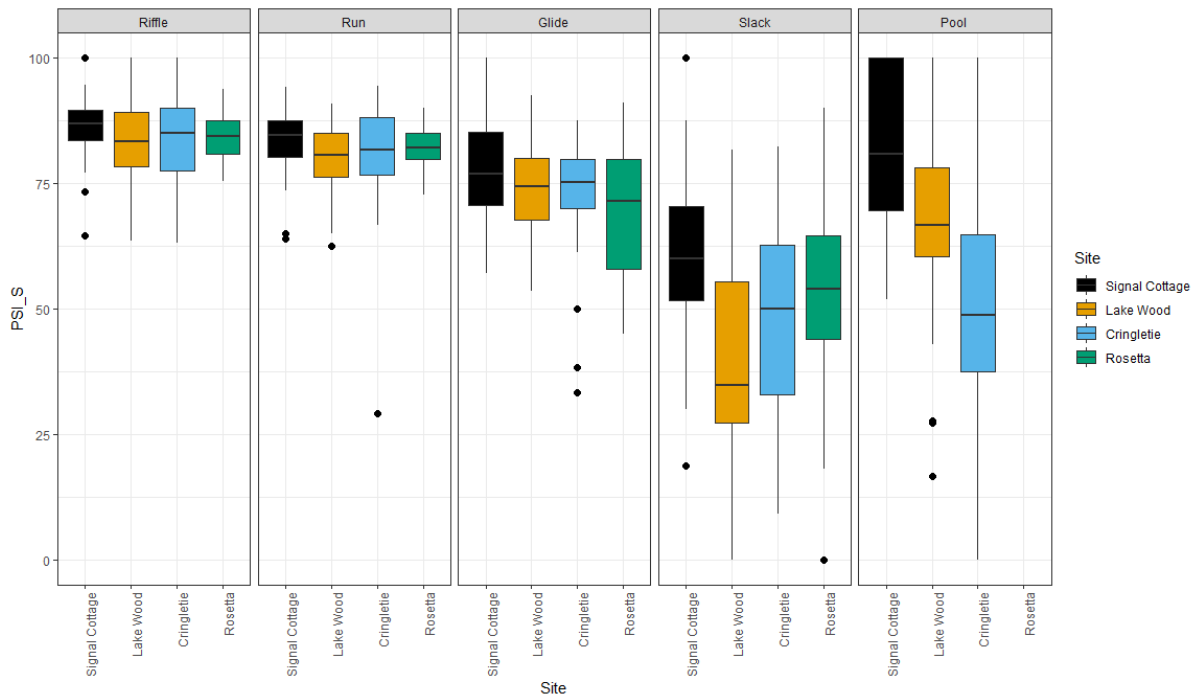
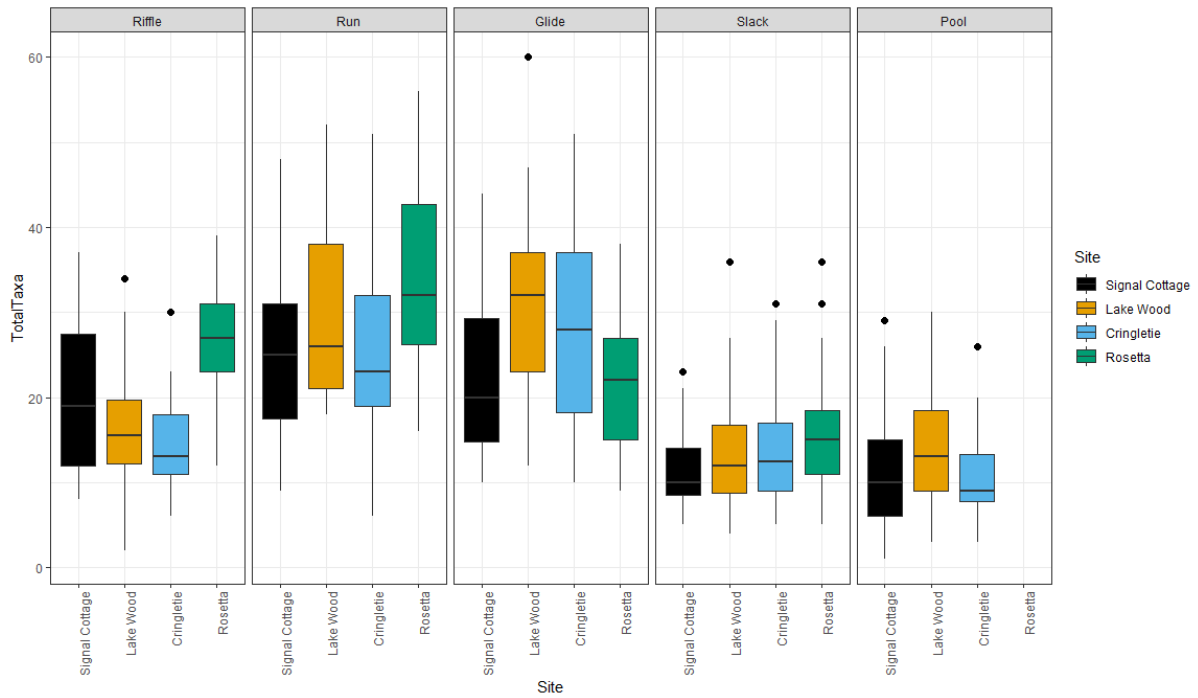
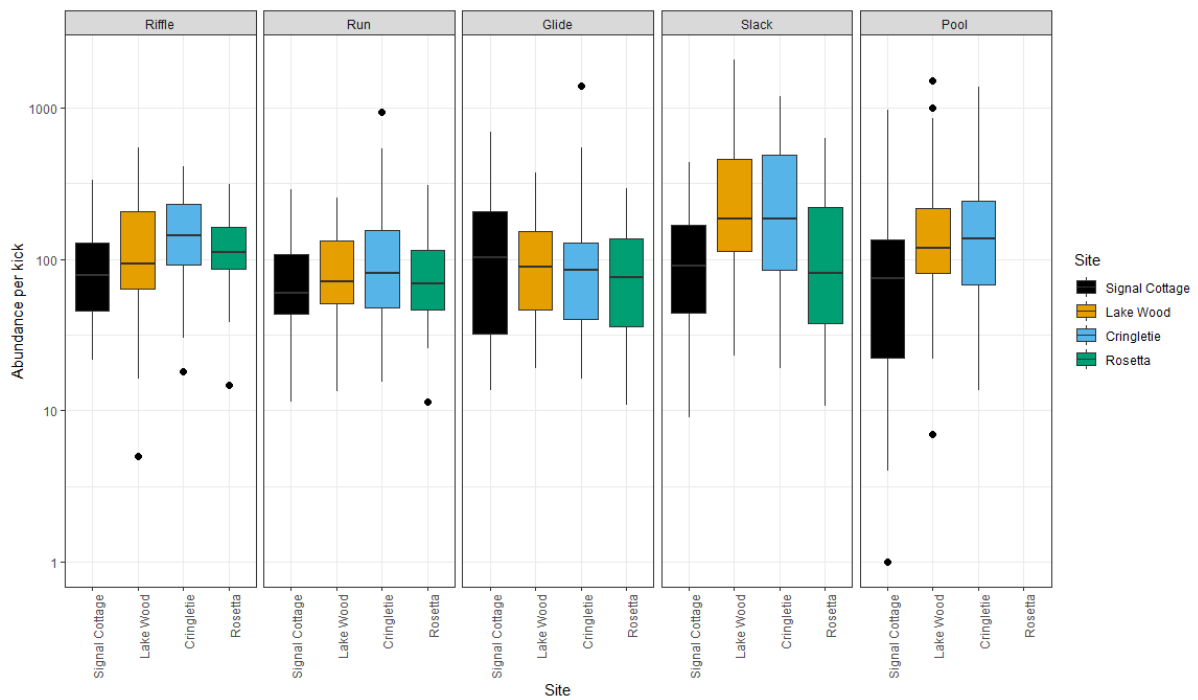


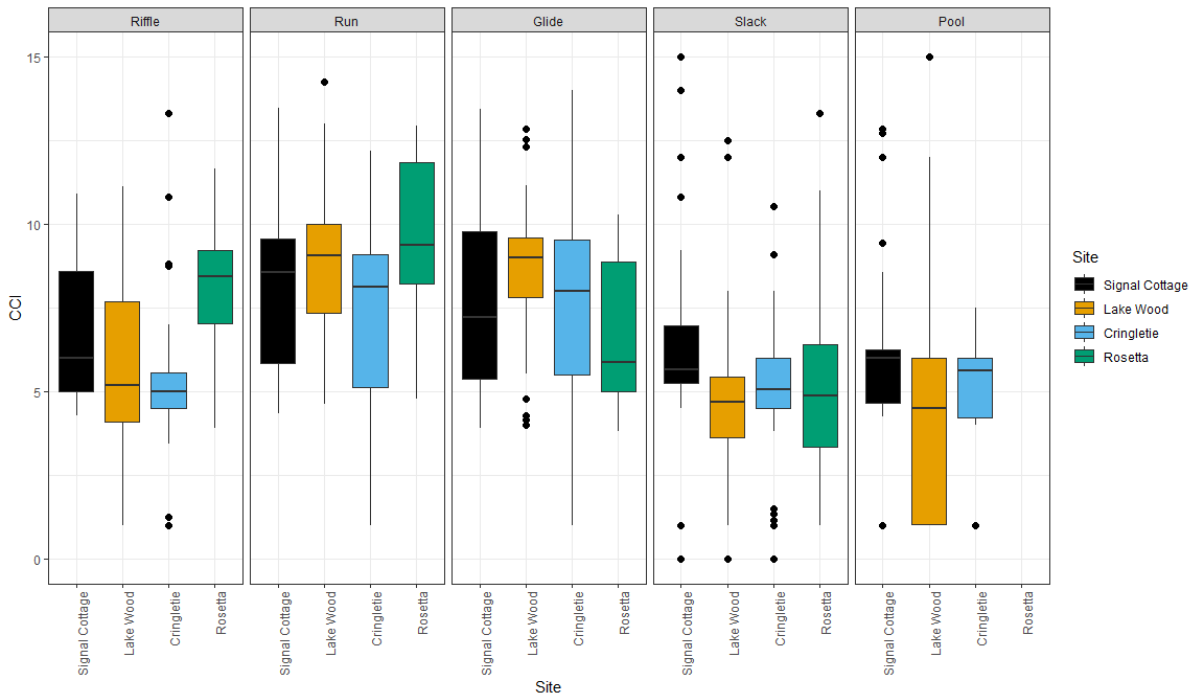
Figure 2.14 Boxplot comparison of species-level PSI by mesohabitat type and site



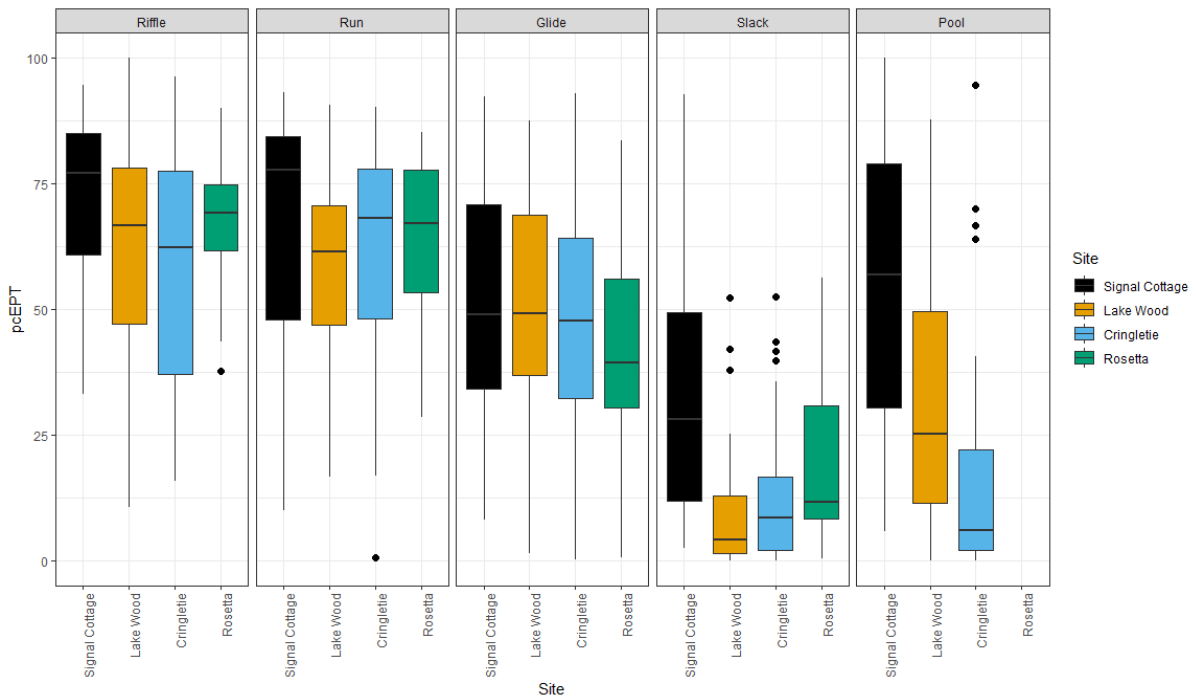
**Figure 2.15** Boxplot comparison of taxon richness by mesohabitat type and site



**Figure 2.16** Boxplot comparison of macroinvertebrate abundance by mesohabitat type and site. Note that abundance has been standardised by the number of kicks in each mesohabitat type to control for differences in sampling effort.

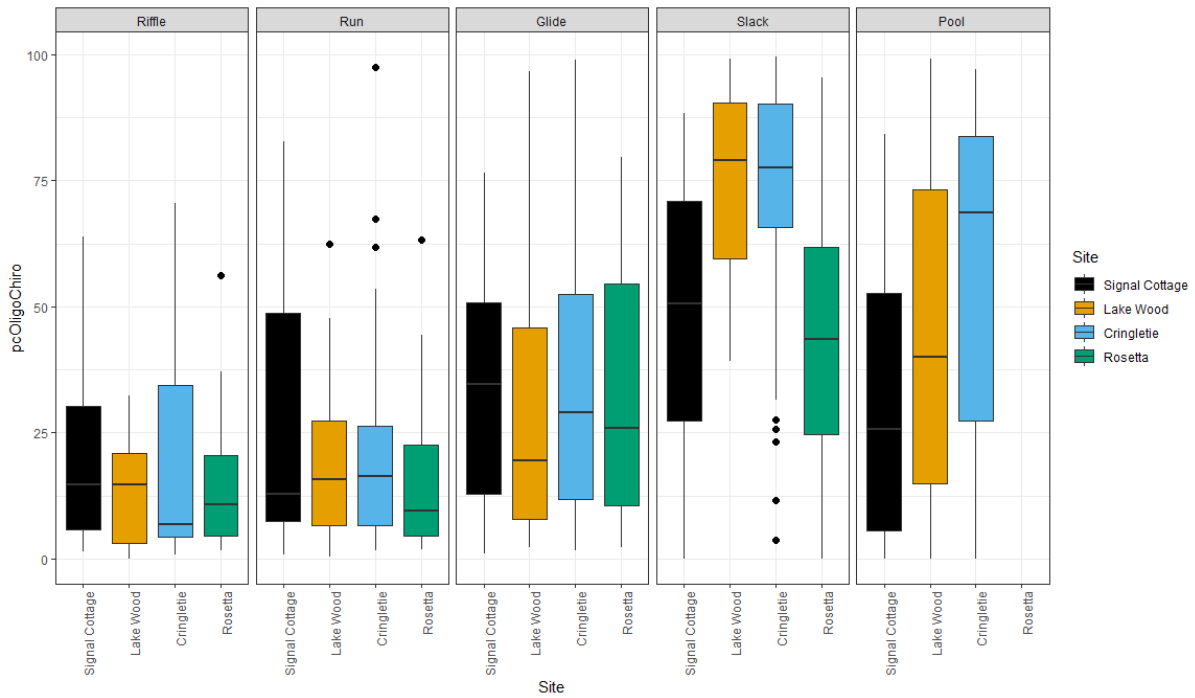


**Figure 2.17** Boxplot comparison of Community Conservation Index by mesohabitat type and site



**Figure 2.18** Boxplot comparison of % Ephemeroptera, Plecoptera and Trichoptera by mesohabitat type and site





**Figure 2.19** Boxplot comparison of % Oligochaeta and Chironomidae by mesohabitat type and site

### 3. Discussion

#### 3.1 Conclusions

In summary, the application of bespoke Before-After-Control-Impact (BACI) monitoring framework has allowed the impact of channel reconfiguration on benthic macroinvertebrates to be evaluated at both reach and mesohabitat scales. The key results are as summarised below.

1. Prior to channel reconfiguration, the two impact sites – Lake Wood and Cringletie – had much less riffle/run and more glide habitat than the two control sites and, with the exception of %EPT, had lower scores for all biotic indices than the two control sites.
2. Channel reconfiguration in 2013 initially increased the proportion of riffle and run habitat and increased overall habitat diversity, but subsequent geomorphological adjustment appears to have partially reversed these changes.
3. Against a background of rapidly increasing taxon richness at all sites, channel reconfiguration caused an abrupt shift in macroinvertebrate community composition at the impact sites from one dominated numerically by mayflies, stoneflies and caddisflies to one dominated by oligochaetes and chironomids.
4. Following the initial disturbance caused by the channel reconfiguration work, the impact and control sites have partially converged in macroinvertebrate composition but only total abundance and %OligoChiro have increased significantly as a result of the intervention.

5. Six years after the channel reconfiguration work, WHPT-ASPT, LIFE\_S, PSI\_F, PSI\_S and %EPT remain significantly lower at the impact sites compared with the control sites.

In conclusion, the benthic macroinvertebrate community in Eddleston Water appears to be strongly influenced by mesohabitat composition. Channel reconfiguration has led to a partial improvement in macroinvertebrate community status (as measured by a variety of standard biotic metrics) but full recovery from historical channel straightening is thought to have been constrained to date by the limited geomorphological changes at Lake Wood and Cringletie.

### 3.2 Lessons learnt

With the benefit of hindsight, it is possible to identify a number learning points that may be helpful when designing evaluation programmes for other river restoration projects in the future.

- Control sites are essential. Over the study period, there was a very strong and consistent increase in taxon richness at all four sites, which drove changes in some of the other biotic indices. If control sites had not been established, then the effect of channel reconfiguration would have been confounded by these background changes in macroinvertebrate community composition, and erroneous conclusions would have been drawn.
- In an ideal world, a longer period of baseline (pre-intervention) monitoring would have provided a more robust assessment of the impact of channel straightening at Lake Wood and Cringletie. With less than two complete years of sampling data available prior to the channel reconfiguration work, statistical power to detect differences was low.
- Geomorphological and biological responses to channel reconfiguration can take place over many years, and the extended period of post-intervention monitoring conducted at Eddleston Water has been valuable in revealing both short-term and longer-term effects of NFM measures.
- The use of upstream and downstream control sites helped to prevent confounding due to any longitudinal gradients in macroinvertebrate composition along the river, but there is a risk that the downstream site (Rosetta in this case) could be affected indirectly by the channel reconfiguration work (e.g. by mobilisation of fine sediment). Control sites should ideally be located on independent water courses that have similar characteristics, except for the effects of the intervention. In this case, however, there were no suitable independent controls available.
- The use of a dis-aggregated sampling method to gather data from individual mesohabitat types provided valuable insight into the importance of physical habitat in structuring the benthic macroinvertebrate community, yet still allowed responses to be assessed at a reach level.
- Macroinvertebrate metrics such as LIFE and PSI can show different patterns when calculated using family-level and species-level data. One reason might be that some families are more speciose than others. However, more granular species-level data does not necessarily reduce the level of residual (measurement) error in the data or produce clearer results when examining spatial differences and temporal trends.
- The use of a semi-quantitative (timed-effort) sampling method to collect macroinvertebrate samples resulted in an appreciable amount of measurement error (i.e. high variability among replicate samples), which limited the statistical power of the analysis to detect patterns in the data. Future studies may wish to consider the

benefits of collecting a greater number of replicate samples, or whether a fully-quantitative sampling approach might offer a more cost-effective solution.

### 3.3 Recommendations

This study has demonstrated a clear response of macroinvertebrates to channel reconfiguration. Habitat composition appears to have played a strong role in mediating these changes, but the rich combination of physical and biological monitoring data collected at Eddleston Water holds further potential to elucidate the mechanisms linking physical changes to biological responses. Specifically, we suggest that future analyses could:

- extend the BACI regression models to understand the influence of habitat composition, fine sediment, submergent vegetation and habitat diversity on macroinvertebrate composition and diversity;
- use multivariate ordination techniques to visualise spatial and temporal patterns in macroinvertebrate composition and identify sensitive taxonomic groups or species traits that could act as indicators of change;
- analyse changes in absolute abundance of key taxonomic groups; and
- examine mesohabitat-scale responses to channel reconfiguration, and how these influence responses at larger (reach) scales.

## 4. References

APEM (2019) Eddleston Water Project – Aquatic Macroinvertebrate Identification and Analyses – 2018 Annual Report. APEM Report to the Tweed Forum, February 2019.

Chadd, R.P and Extence, C.A. (2004) The conservation of freshwater macroinvertebrate populations: a community-based classification scheme. *Aquatic Conservation: Marine and Freshwater Ecosystems* 14: 597–624.

Environment Agency (2017) Freshwater macro-invertebrate sampling in rivers. Operational Instruction 018\_08, issued 01 March 2017.

Extence, C.A., Balbi, D.M. and Chadd, R.P. (1999) River flow indexing using British benthic macroinvertebrates: a framework for setting hydro-ecological objectives. *Regulated Rivers: Research and Management* 15: 543–574.

Extence, C. A., Chadd, R. P., England, J., Dunbar, M. J., and Taylor, E. D. (2011). The assessment of fine sediment accumulation in rivers using macroinvertebrate community response. *River Research and Applications* 29: 17–55.

Feld, C.K., Birk, S., Hering, D., Marzin, A., Melcher, A., Nemitz, D., Pont, D., Bradley, D.C., Pedersen, M. and Friberg, N. (2011) From natural to degraded rivers and back again: a test of restoration ecology and theory. *Advances in Ecological Research* 44: 119-209.

Paisley, M.F., Walley, W.J. and Trigg, D.J. (2013) Revision of the Biological Monitoring Working Party (BMWP) score system: Derivation of present-only and abundance related scores from field data. *River Research and Applications* 30: 887–904.

R Core Team. (2020) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. <http://www.R-project.org>

Veritas Ecology (2017) Early Change in the Physical Habitat and the Macroinvertebrate Community in the Eddleston Water in Response to Channel Re-Configuration 2012-2015. Veritas Ecology, Loch Lomond, Scotland.

## Appendix 1 Summary of sampling programme

Period	Year	Season	Month	Cringletie	Lake Wood	Signal Cottage	Rosetta
Before	2012	Spring	May	1 x3 min sample		1 x3 min sample	1 x3 min sample
		Summer	Aug	1 x3 min sample		1 x3 min sample	1 x3 min sample
		Autumn					
Following	2013	Spring	May				
		Summer	Jun				
		Autumn	Nov				
	2014	Spring	Apr				
		Spring	May*	Family level only	Family level only	Only 1 pool rep	
		Summer	Aug			**	
After	2015	Spring	May				
		Summer					
		Autumn					
	2016	Spring	May				
		Summer					
		Autumn	Nov				
	2017	Spring	May				
		Summer					
		Autumn	Nov				
	2018	Spring					
		Summer					
		Autumn					
	2019	Spring	May				
		Summer					
		Autumn	Nov				

20 kicks x 3 reps at each site in each season, unless otherwise stated. Grey = no sampling.

\* Excluded from analysis in preference to April 2014 samples, to match species-level identification used in other years.

\*\* No sampling because site impacted by channel reconfiguration immediately upstream.

## Appendix 2 Results of statistical analyses comparing reach-level biotic indices among sites and time periods

### WHPT\_ASPT

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: WHPT\_ASPT ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
26.1	59.5	-2.1	4.1	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.63084	-0.59040	0.07547	0.64521	2.04260

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	0.0042932	0.06552
Site	(Intercept)	0.0006289	0.02508
Residual		0.0564574	0.23761

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )
(Intercept)	6.43567	0.08250	30.31523	78.008	< 2e-16 ***
SeasonSummer	-0.18293	0.08227	19.22392	-2.223	0.03836 *
SeasonAutumn	0.12894	0.06012	14.72932	2.145	0.04908 *
TreatmentImpact	-0.11327	0.09073	48.59312	-1.248	0.21785
Period2Following	0.11481	0.09944	41.91978	1.155	0.25482
Period2After	0.33458	0.09589	32.20434	3.489	0.00143 **
TreatmentImpact:Period2Following	-0.10646	0.11139	142.78593	-0.956	0.34085
TreatmentImpact:Period2After	-0.15145	0.10363	138.16610	-1.461	0.14615

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

## Family level LIFE

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: LIFE\_F ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
-77.2	-43.8	49.6	-99.2	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.4447	-0.6355	-0.1171	0.5482	2.6214

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	0.0053241	0.07297
Site	(Intercept)	0.0006412	0.02532
Residual		0.0271893	0.16489

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	7.884039	0.069597	23.689154	113.282	< 2e-16	***
SeasonSummer	-0.040541	0.072647	16.554200	-0.558	0.584277	
SeasonAutumn	-0.003769	0.054778	13.250180	-0.069	0.946172	
TreatmentImpact	-0.292861	0.065717	32.296326	-4.456	9.41e-05	***
Period2Following	-0.316113	0.081748	29.168831	-3.867	0.000569	***
Period2After	-0.227432	0.080808	22.953097	-2.814	0.009849	**
TreatmentImpact:Period2Following	0.296934	0.077778	139.519036	3.818	0.000202	***
TreatmentImpact:Period2After	0.207028	0.072028	135.564049	2.874	0.004703	**

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

## Species level LIFE

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]  
 Formula: LIFE\_S ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)  
 Data: agdata

AIC	BIC	logLik	deviance	df.resid
-84.6	-51.3	53.3	-106.6	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.2980	-0.7028	-0.1065	0.5715	2.9798

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	0.0005686	0.02384
Site	(Intercept)	0.0001030	0.01015
Residual		0.0285487	0.16896

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	8.58857	0.05104	27.44489	168.261	< 2e-16	***
SeasonSummer	0.05397	0.04878	14.94924	1.106	0.28609	
SeasonAutumn	0.06421	0.03467	10.94528	1.852	0.09113	.
TreatmentImpact	-0.18741	0.06272	63.97427	-2.988	0.00398	**
Period2Following	-0.29679	0.06315	41.82885	-4.699	2.83e-05	***
Period2After	-0.10400	0.05981	31.81767	-1.739	0.09171	.
TreatmentImpact:Period2Following	0.16848	0.07876	141.14350	2.139	0.03414	*
TreatmentImpact:Period2After	0.01741	0.07360	134.22498	0.237	0.81332	

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1





## Family level PSI

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: PSI\_F ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
927.4	960.8	-452.7	905.4	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.21202	-0.58163	-0.09944	0.57253	3.09646

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	2.204	1.485
Site	(Intercept)	0.000	0.000
Residual		20.259	4.501

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )
(Intercept)	67.64876	1.61844	31.04305	41.799	<2e-16 ***
SeasonSummer	0.87987	1.68665	18.06248	0.522	0.6082
SeasonAutumn	1.10968	1.24669	14.05590	0.890	0.3884
TreatmentImpact	-4.25808	1.65225	141.32068	-2.577	0.0110 *
Period2Following	-3.54767	1.98575	36.67029	-1.787	0.0823 .
Period2After	0.07783	1.93180	28.26389	0.040	0.9681
TreatmentImpact:Period2Following	3.19042	2.11357	144.94813	1.509	0.1333
TreatmentImpact:Period2After	0.16931	1.96352	140.75755	0.086	0.9314

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1



### Species level PSI

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: PSI\_S ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
933.1	966.4	-455.5	911.1	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.5179	-0.6313	0.0135	0.5631	2.2510

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	2.3331	1.5275
Site	(Intercept)	0.1506	0.3881
Residual		20.8745	4.5689

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	73.8856	1.6738	23.0889	44.141	< 2e-16	***
SeasonSummer	2.3576	1.7239	15.2872	1.368	0.19121	
SeasonAutumn	2.2436	1.2750	11.8383	1.760	0.10426	
TreatmentImpact	-3.2292	1.7220	50.3009	-1.875	0.06657	.
Period2Following	0.8961	2.0257	31.6217	0.442	0.66123	
Period2After	5.7233	1.9714	24.1293	2.903	0.00778	**
TreatmentImpact:Period2Following	1.6069	2.1465	139.1003	0.749	0.45535	
TreatmentImpact:Period2After	-3.7684	1.9936	133.6621	-1.890	0.06089	.

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1



## Taxon richness

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: TotalTaxa ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
1077.0	1110.4	-527.5	1055.0	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.73796	-0.58596	-0.08071	0.71565	2.75104

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	22.637	4.758
Site	(Intercept)	9.238	3.039
Residual		46.565	6.824

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	27.560	4.230	19.690	6.516	2.56e-06	***
SeasonSummer	-1.168	4.132	16.173	-0.283	0.78107	
SeasonAutumn	1.176	3.215	13.836	0.366	0.72003	
TreatmentImpact	-2.172	3.946	8.392	-0.550	0.59642	
Period2Following	12.960	4.364	22.470	2.970	0.00696	**
Period2After	27.185	4.434	19.050	6.131	6.73e-06	***
TreatmentImpact:Period2Following	-1.486	3.238	138.276	-0.459	0.64706	
TreatmentImpact:Period2After	3.949	2.986	136.109	1.323	0.18821	

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

## Total abundance

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]  
 Formula: log10(TotalAbundance) ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)  
 Data: agdata

AIC	BIC	logLik	deviance	df.resid
-16.3	17.1	19.1	-38.3	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.1232	-0.6039	-0.1712	0.5063	2.6112

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	0.02631	0.1622
Site	(Intercept)	0.00000	0.0000
Residual		0.03732	0.1932

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	3.15495	0.11790	17.81627	26.760	7.77e-16	***
SeasonSummer	-0.05007	0.13580	14.74019	-0.369	0.7176	
SeasonAutumn	-0.09841	0.10685	12.98017	-0.921	0.3738	
TreatmentImpact	-0.10804	0.07119	137.77875	-1.518	0.1314	
Period2Following	-0.07062	0.14045	18.95690	-0.503	0.6209	
Period2After	0.20168	0.14426	16.62306	1.398	0.1805	
TreatmentImpact:Period2Following	0.40065	0.09166	139.26346	4.371	2.40e-05	***
TreatmentImpact:Period2After	0.17682	0.08451	137.46408	2.092	0.0382	*

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1



CCI

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: CCI ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
725.3	758.6	-351.7	703.3	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-1.8537	-0.8007	-0.0011	0.5817	3.8248

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	0.2585	0.5084
Site	(Intercept)	0.2650	0.5148
Residual		5.4471	2.3339

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	8.3487	0.8302	21.1111	10.056	1.66e-09	***
SeasonSummer	0.5242	0.7442	14.1405	0.704	0.492596	
SeasonAutumn	0.6046	0.5367	10.5309	1.126	0.284976	
TreatmentImpact	-0.6487	0.9990	21.9400	-0.649	0.522869	
Period2Following	2.5173	0.9269	35.3573	2.716	0.010159	*
Period2After	3.7762	0.8855	26.5470	4.264	0.000226	***
TreatmentImpact:Period2Following	-0.1103	1.0921	140.5996	-0.101	0.919675	
TreatmentImpact:Period2After	-0.1291	1.0176	134.3415	-0.127	0.899220	

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1



## % EPT

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: pcEPT ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
1286.0	1319.4	-632.0	1264.0	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.18096	-0.69983	-0.03744	0.67837	2.10007

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	8.836e+01	9.400e+00
Site	(Intercept)	2.358e-08	1.536e-04
Residual		1.922e+02	1.386e+01

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	48.0423	7.2596	21.6955	6.618	1.27e-06	***
SeasonSummer	-0.2232	8.2124	16.8138	-0.027	0.9786	
SeasonAutumn	3.3761	6.3804	14.3479	0.529	0.6048	
TreatmentImpact	5.3601	5.1044	139.7895	1.050	0.2955	
Period2Following	9.8426	8.6964	23.6083	1.132	0.2691	
Period2After	12.4047	8.8294	19.9768	1.405	0.1754	
TreatmentImpact:Period2Following	-30.9949	6.5651	141.6222	-4.721	5.58e-06	***
TreatmentImpact:Period2After	-18.7393	6.0605	139.4230	-3.092	0.0024	**

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1



### % Oligochaetes and chironomids

Linear mixed model fit by maximum likelihood . t-tests use Satterthwaite's method [lmerModLmerTest]

Formula: pcOligoChiro ~ Season + Treatment \* Period2 + (1 | YearMonth) + (1 | Site)

Data: agdata

AIC	BIC	logLik	deviance	df.resid
1297.8	1331.2	-637.9	1275.8	142

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.11065	-0.62559	-0.04632	0.73558	2.36515

Random effects:

Groups	Name	Variance	Std.Dev.
YearMonth	(Intercept)	127.570	11.295
Site	(Intercept)	9.898	3.146
Residual		197.502	14.054

Number of obs: 153, groups: YearMonth, 15; Site, 4

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t )	
(Intercept)	44.368	8.595	20.190	5.162	4.61e-05	***
SeasonSummer	-5.529	9.531	15.683	-0.580	0.5701	
SeasonAutumn	-11.669	7.480	13.716	-1.560	0.1415	
TreatmentImpact	-9.403	6.063	19.512	-1.551	0.1370	
Period2Following	-19.190	9.905	20.512	-1.937	0.0666	.
Period2After	-17.886	10.146	17.839	-1.763	0.0950	.
TreatmentImpact:Period2Following	44.861	6.674	137.201	6.722	4.42e-10	***
TreatmentImpact:Period2After	15.282	6.151	135.235	2.484	0.0142	*

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

