


Willkommen
Welcome
Bienvenue



In vitro studies for antibacterial concepts
in the field of bone implant materials

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Bone Innovation Summit 2019
Feb 12-14, 2019

Biointerfaces

MATERIALS

MEET

LIFE



Materials

biomolecules at surfaces

surface functionalisation to steer biological response

bacteria at surfaces

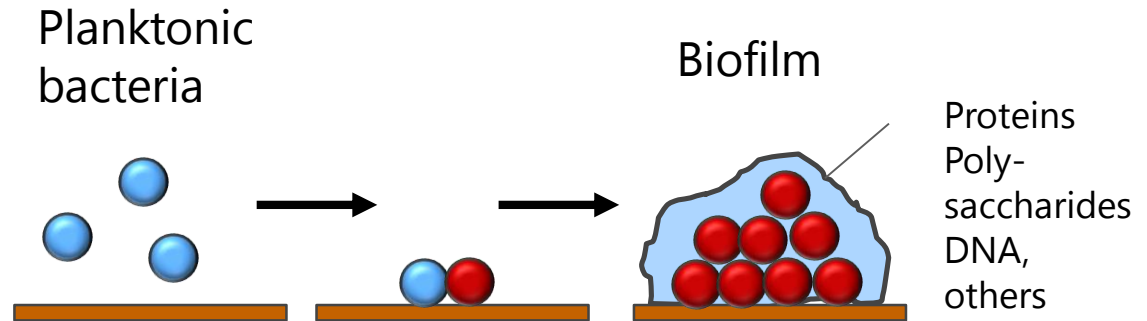
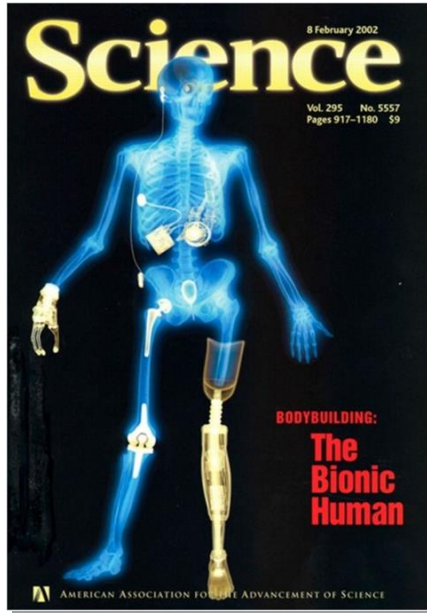
bacteria-free surfaces/ specific bacteria population

cells at surfaces

functional biocompatibility



Biomaterials solve & *generate* problems

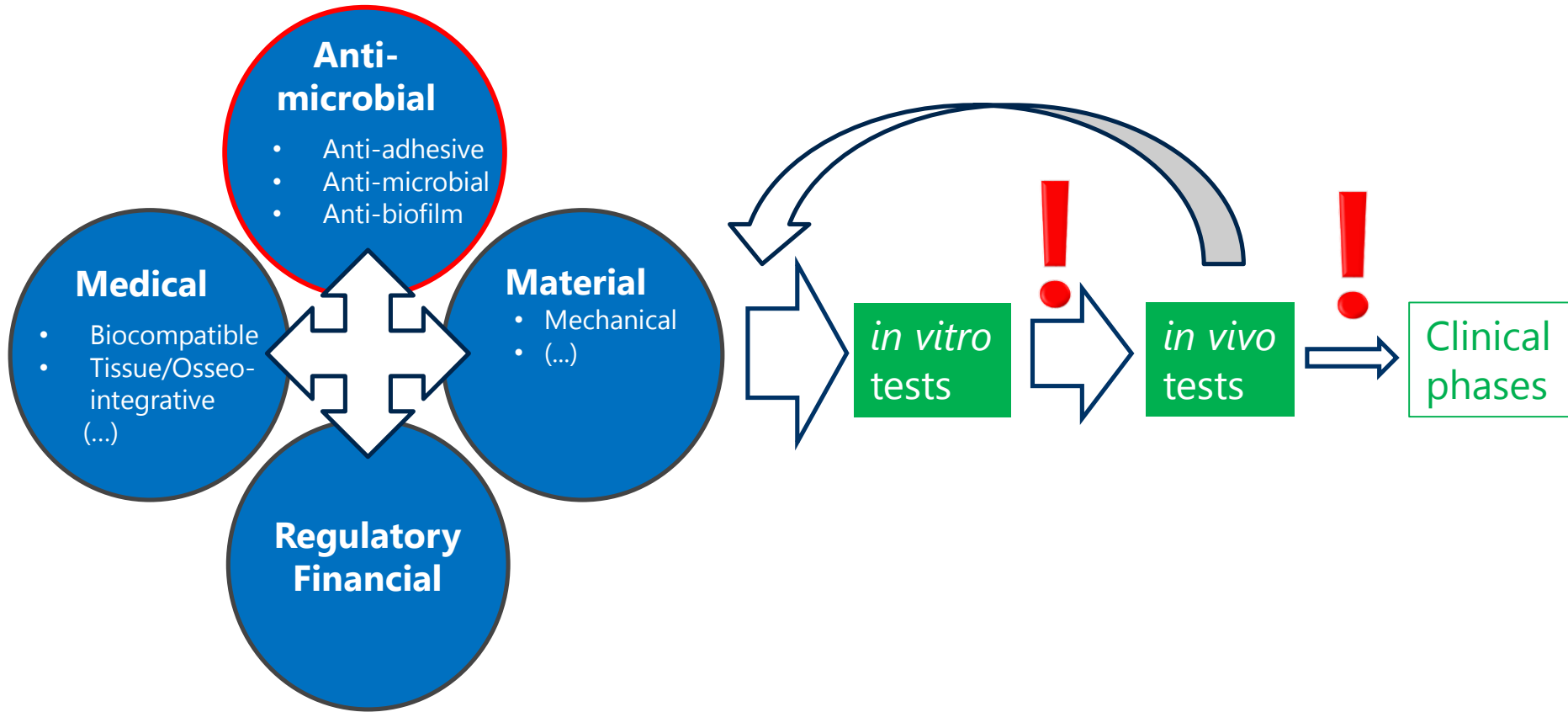


Biofilms: Communities of microorganisms embedded in a matrix of extracellular polymeric substances

Bacteria in biofilms tolerate the ~1000-fold antibiotics concentration, compared to planktonic populations

➔ Antimicrobial materials with anti-biofilm properties are highly demanded

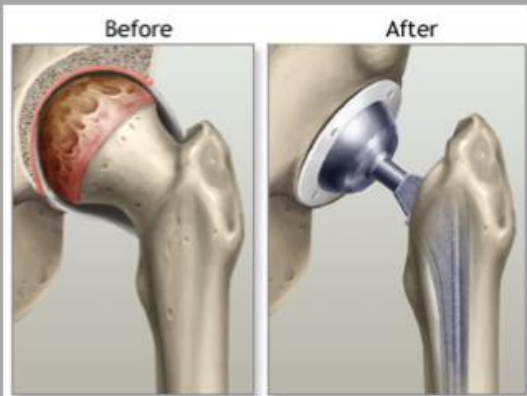
Many promising antimicrobial biomaterials show decent *in vitro* activity but only poor *in vivo* efficacy



➔ Partially due to the lack of predictive laboratory biofilm *in vitro* models

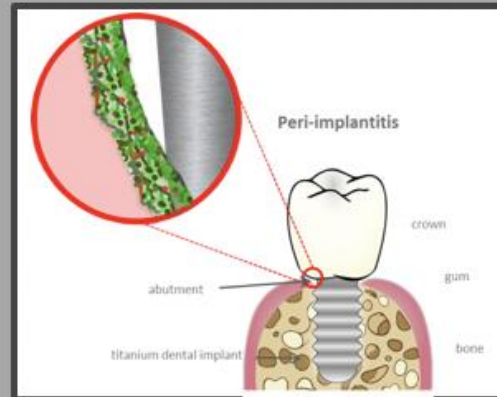
The site of action defines the antimicrobial strategy – and the *in vitro* bioassay

Hip joint implants



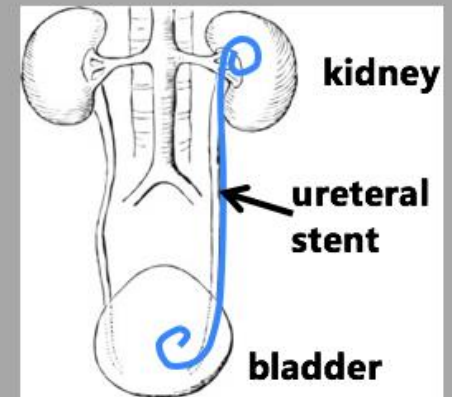
Low bacterial load
(infection during surgery
or late infections)

Dental implants



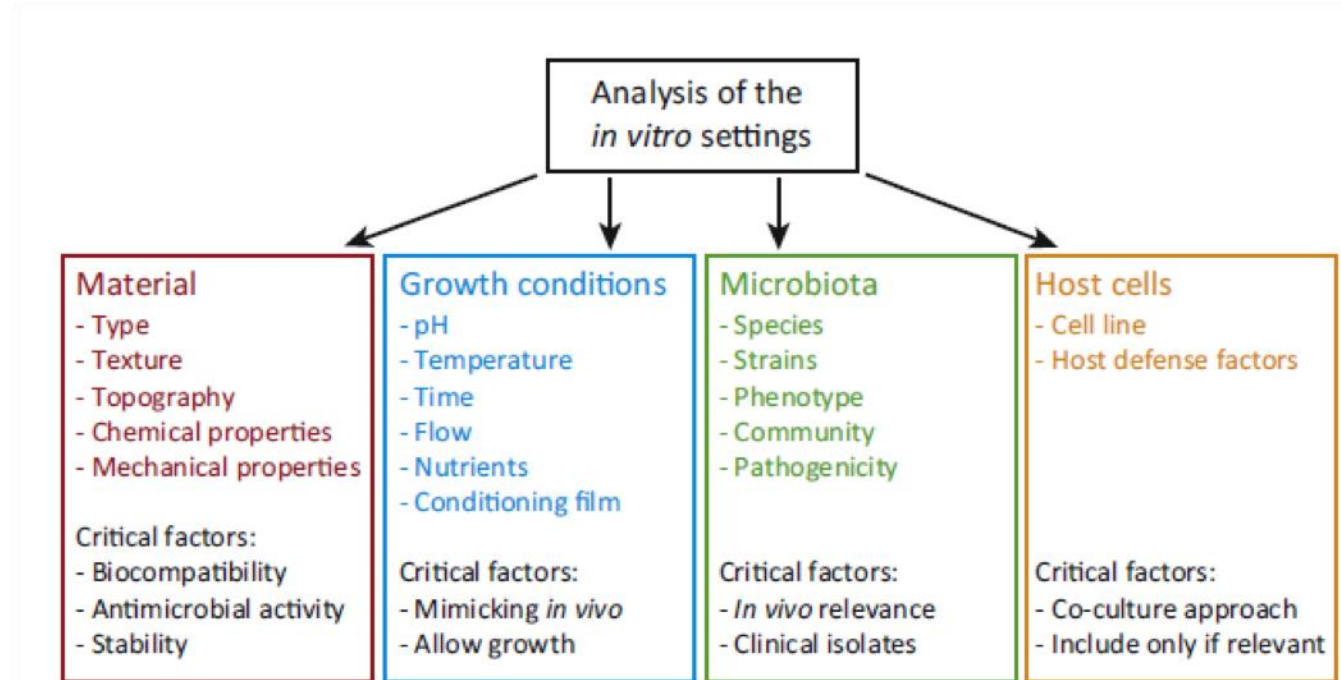
High bacterial load
multiple species involved

Medical devices: Ureteral stents



Low to high
bacterial load ?

Important factors for *in vitro* biofilm assessment

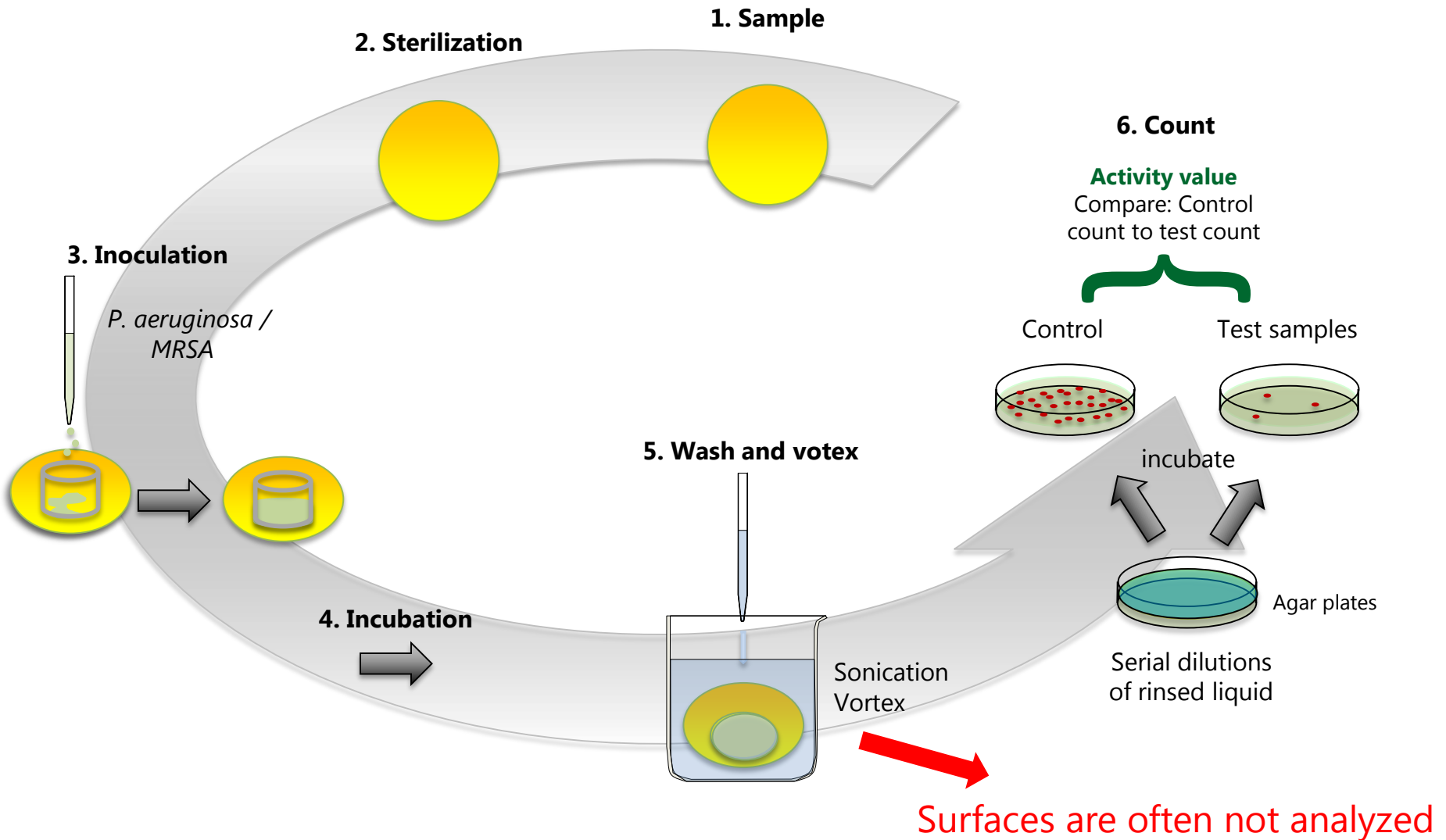


Objective:

Better, predictive biofilm *in vitro* models for antimicrobial materials testing

How are materials tested?

Example: standard antibacterial assays



Stainless steel implants: one example

External fixation



High rate of
pin track
infection



Inefficient treatment

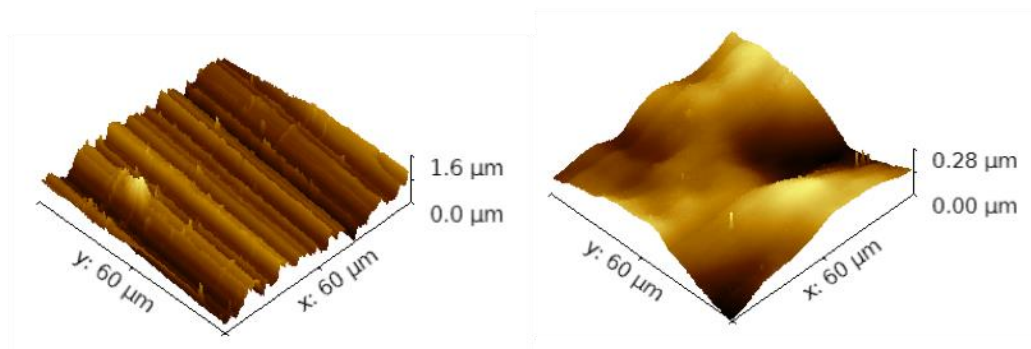


Origin of Pin-Tract Infection

Bacterium	%
<i>Staphylococcus aureus</i>	47.1
<i>Staphylococcus epidermidis</i>	11.8
<i>Escherichia coli</i>	9.4
<i>Pseudomonas aeruginosa</i>	9.4
<i>Streptococcus spp.</i>	3.5
<i>Enterococcus faecalis</i>	2.4
<i>Serratia marcescens</i>	2.4
<i>Vibrio vulnificus</i>	2.4
Mixed flora	3.5
Other	8.1

Antoci et al. *Am J Orthop.* 2008;37(9):E150-E154

Characterization of surface properties of stainless steels



	Roughness (nm) R_a, R_q	Contact angle (°) $\theta_{w\parallel}, \theta_{w\perp}$	Contact angle (°) $\theta_{mi\parallel}, \theta_{mi\perp}$	Zeta potential (mV)
Untreated	172.5 217.9	77.1 101.7	42.3 85.8	- 40.0
P240s	45.2 56.6	80.4 78.8	47.4 47.4	- 46.8

Influence of surface roughness on bacterial adhesion

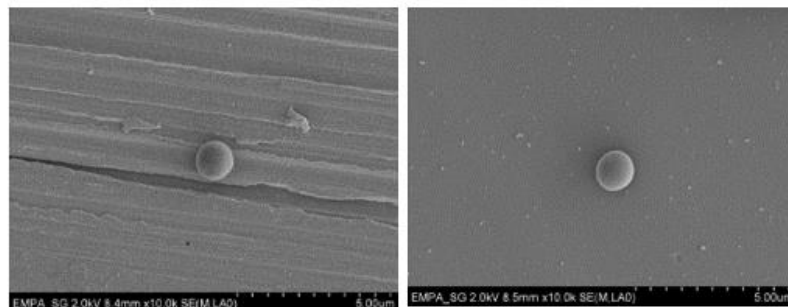
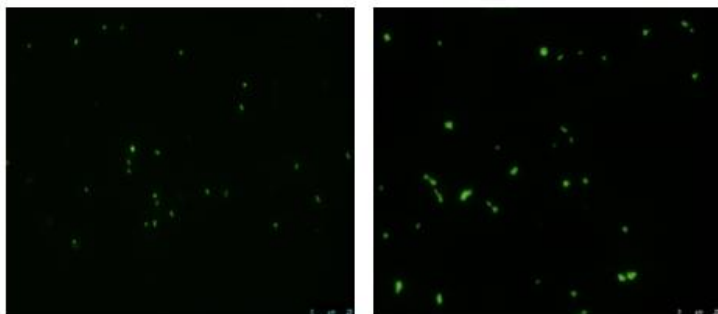
Untreated

P240s

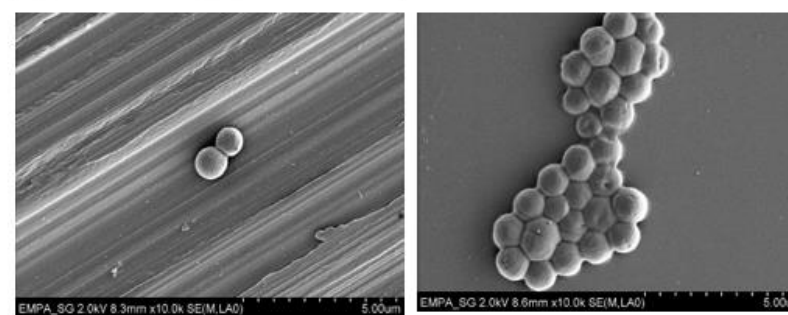
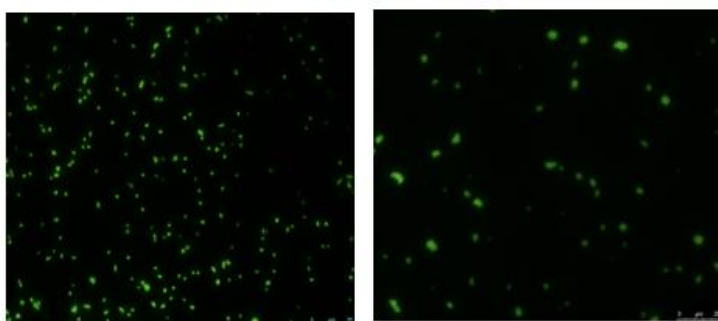
Untreated

P240s

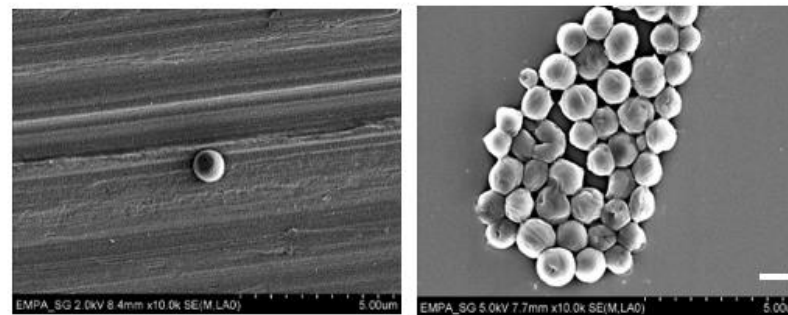
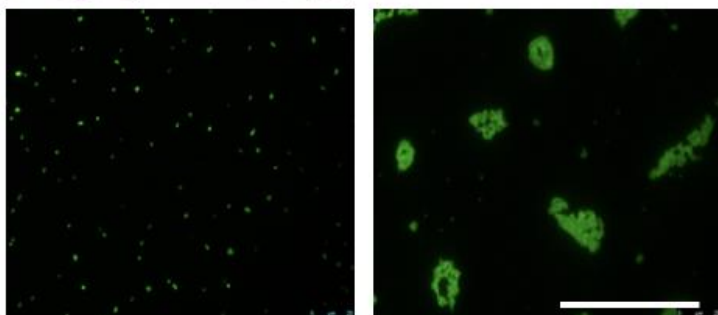
0h



4h



24h



Surface topography influences greatly bacterial colonization

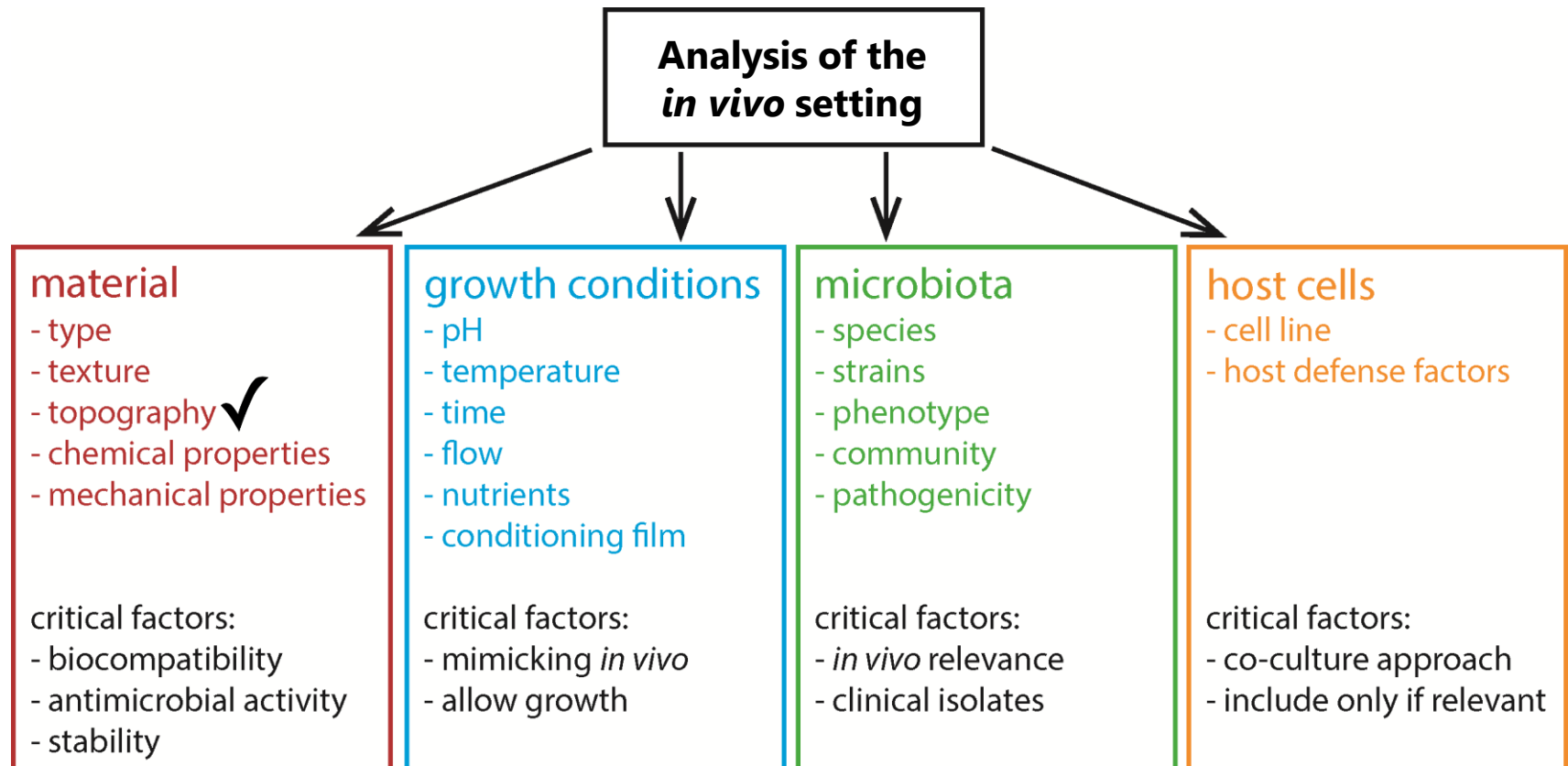
Bacterial adhesion on stainless steels

	Roughness (nm) R_a, R_q	Contact angle (°) $\theta_{w\parallel}, \theta_{w\perp}$	Contact angle (°) $\theta_{mi\parallel}, \theta_{mi\perp}$	Zeta potential (mV)	S.a. Viable cells (CFU/mL)*
Untreated	172.5 217.9	77.1 101.7	42.3 85.8	- 40.0	3.1×10^2
P240s	45.2 56.6	80.4 78.8	47.4 47.4	- 46.8	1.9×10^4

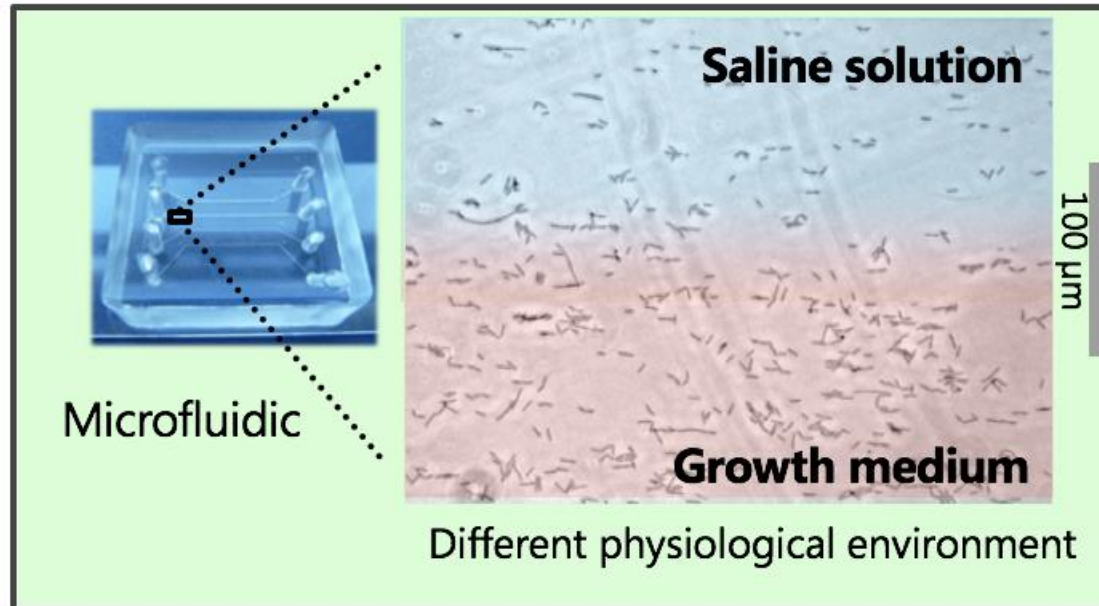
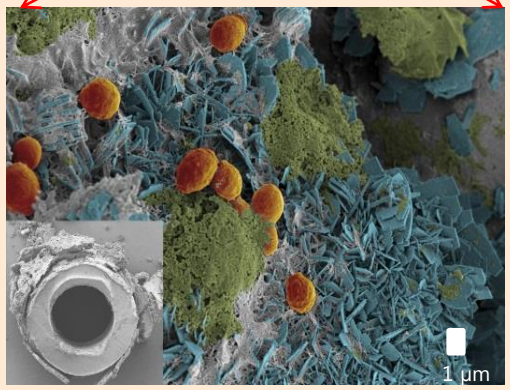
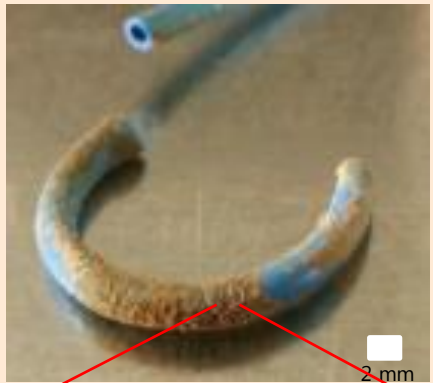
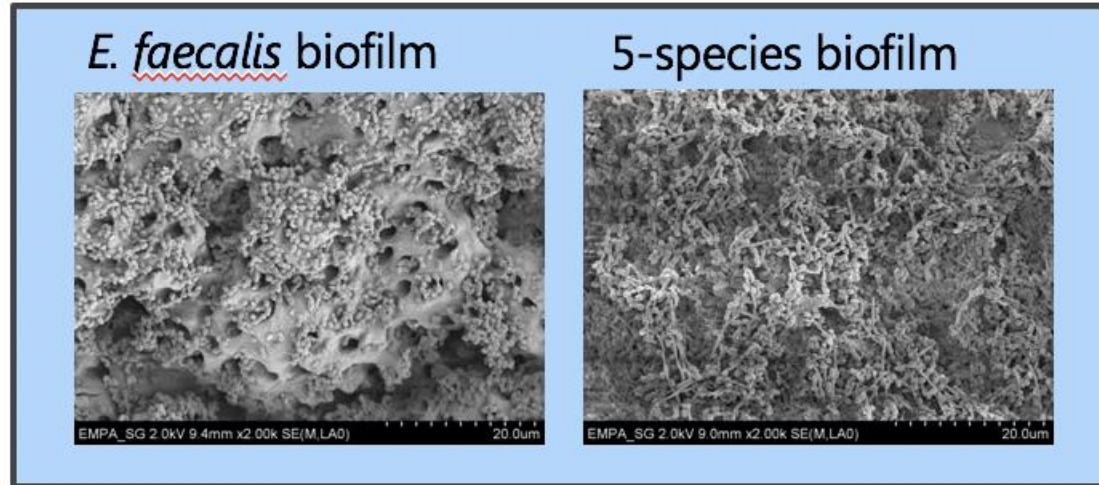
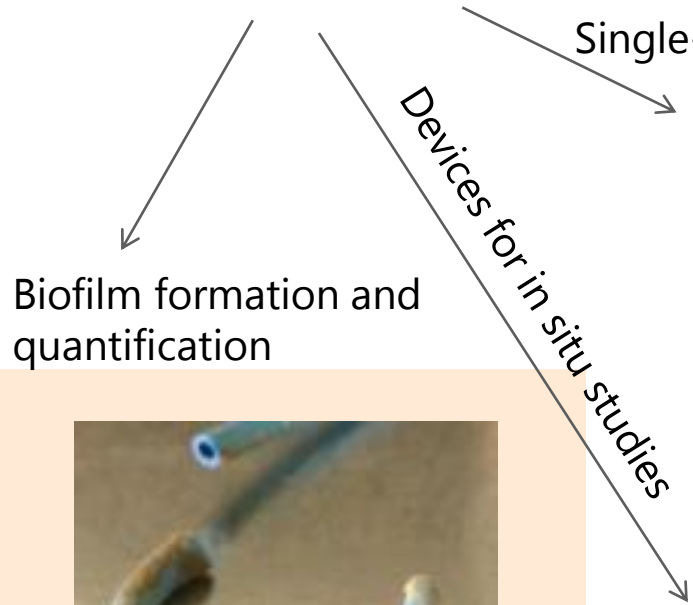
*: adhered viable cells after incubation of 4 h; S.a.: *S. aureus*

Surface topography influences greatly bacterial colonization

Take home messages

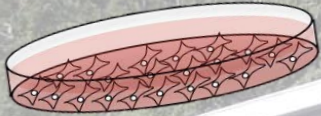


Examples: model systems



Major need

Understanding the limitations of in vitro studies -> developing predictive assays





Thank you