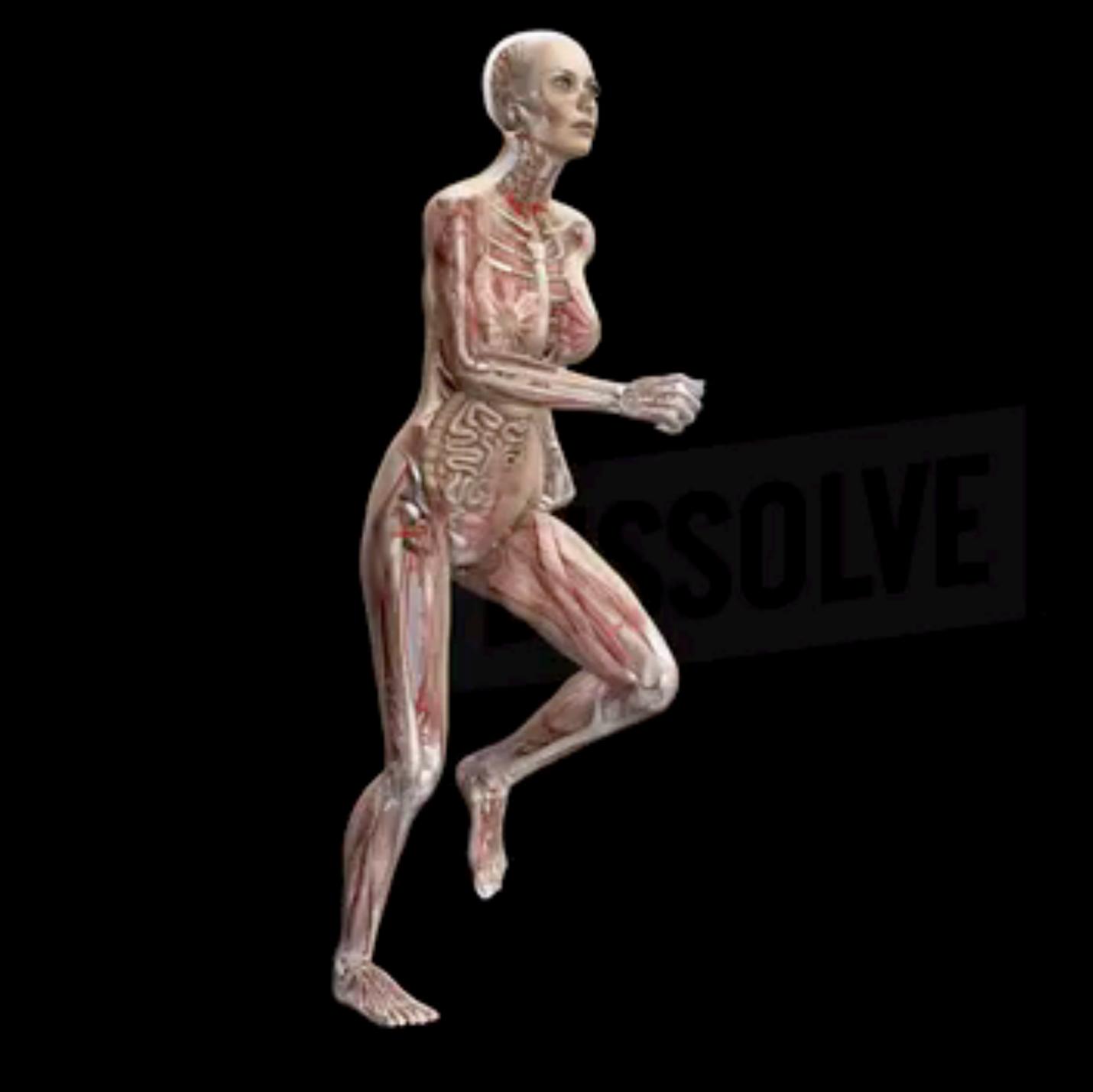


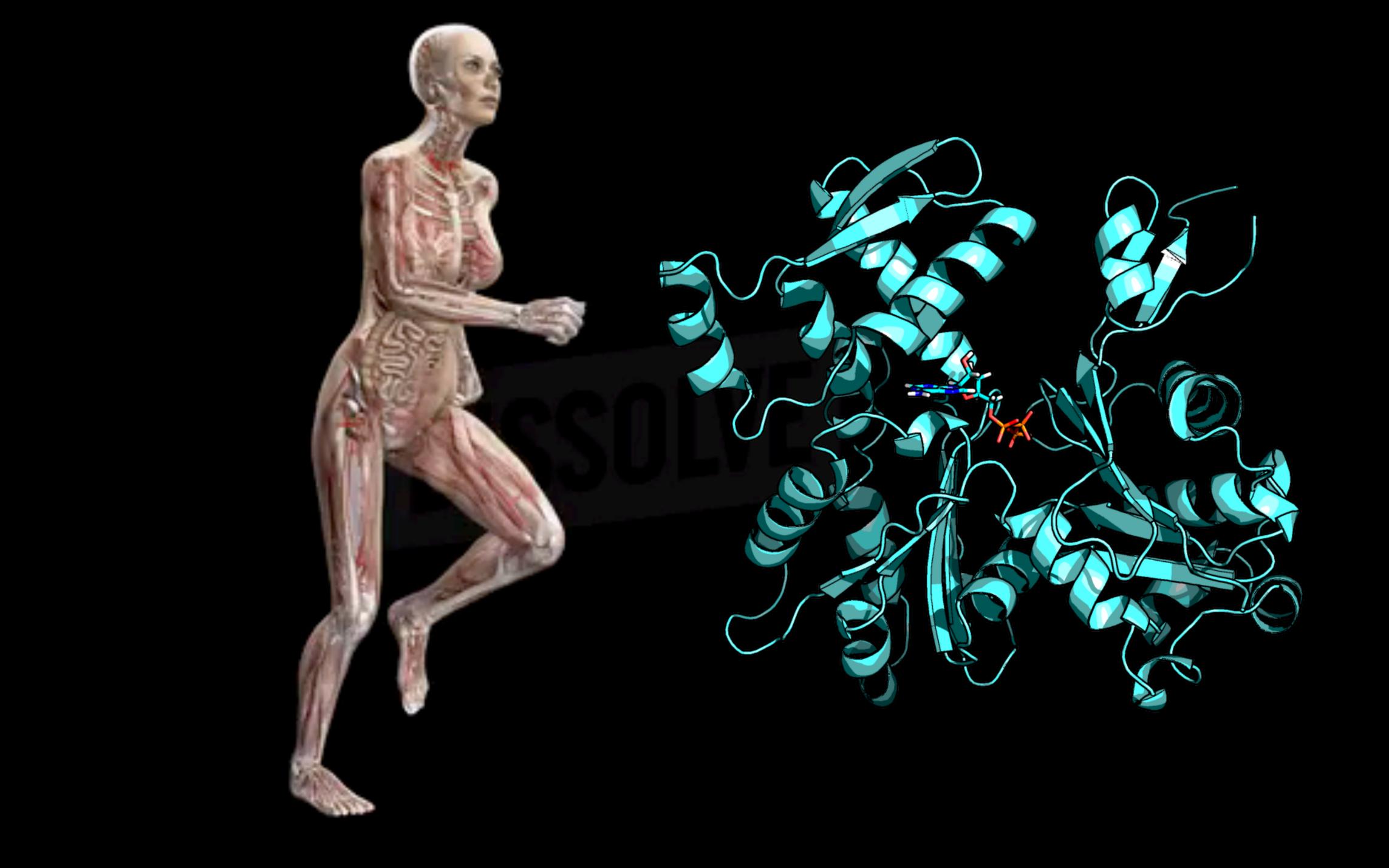
# USING PHOTONS AND ELECTRONS TO UNDERSTAND MALARIA PARASITE MOTILITY

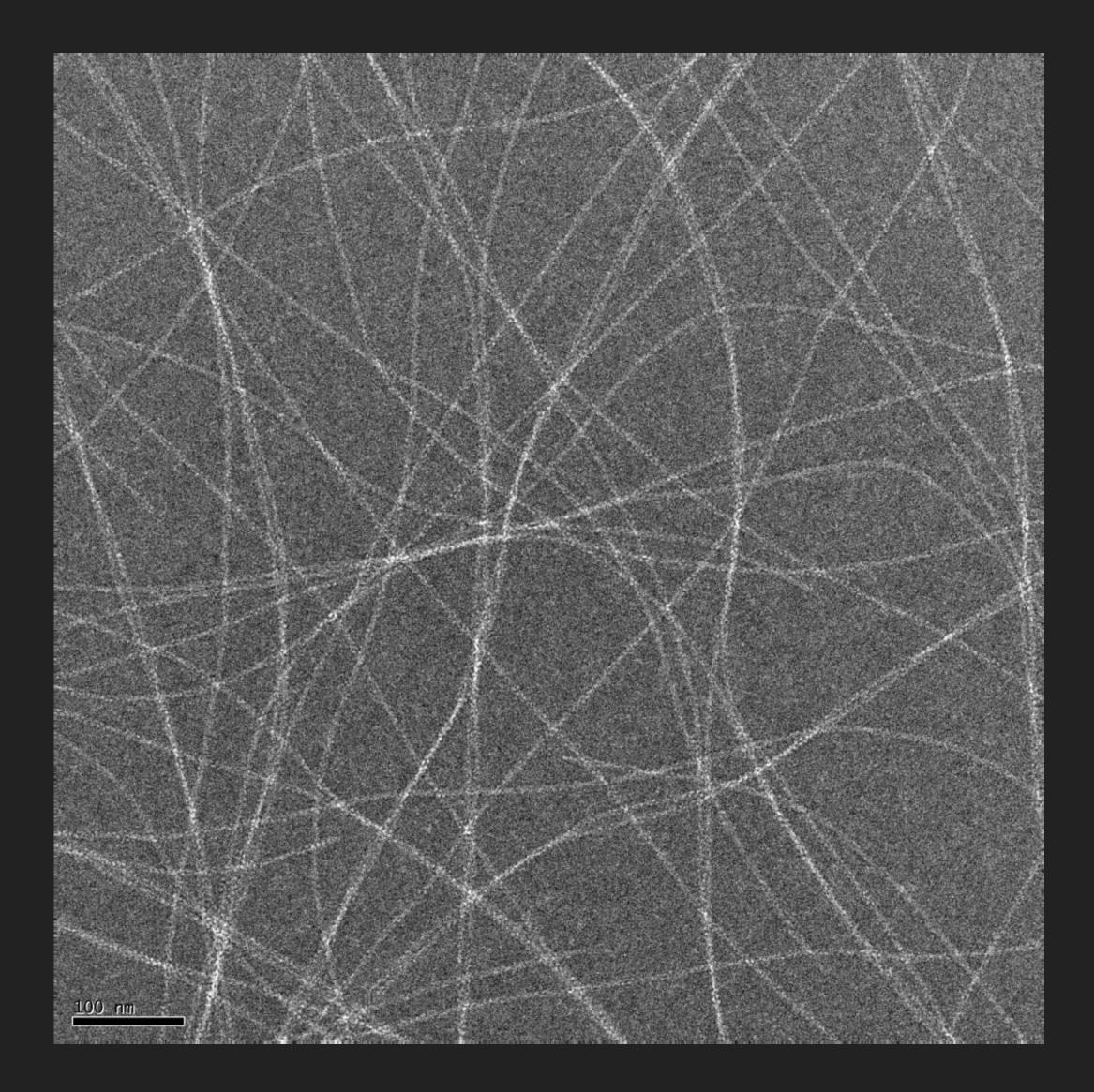
#### **INARI KURSULA**

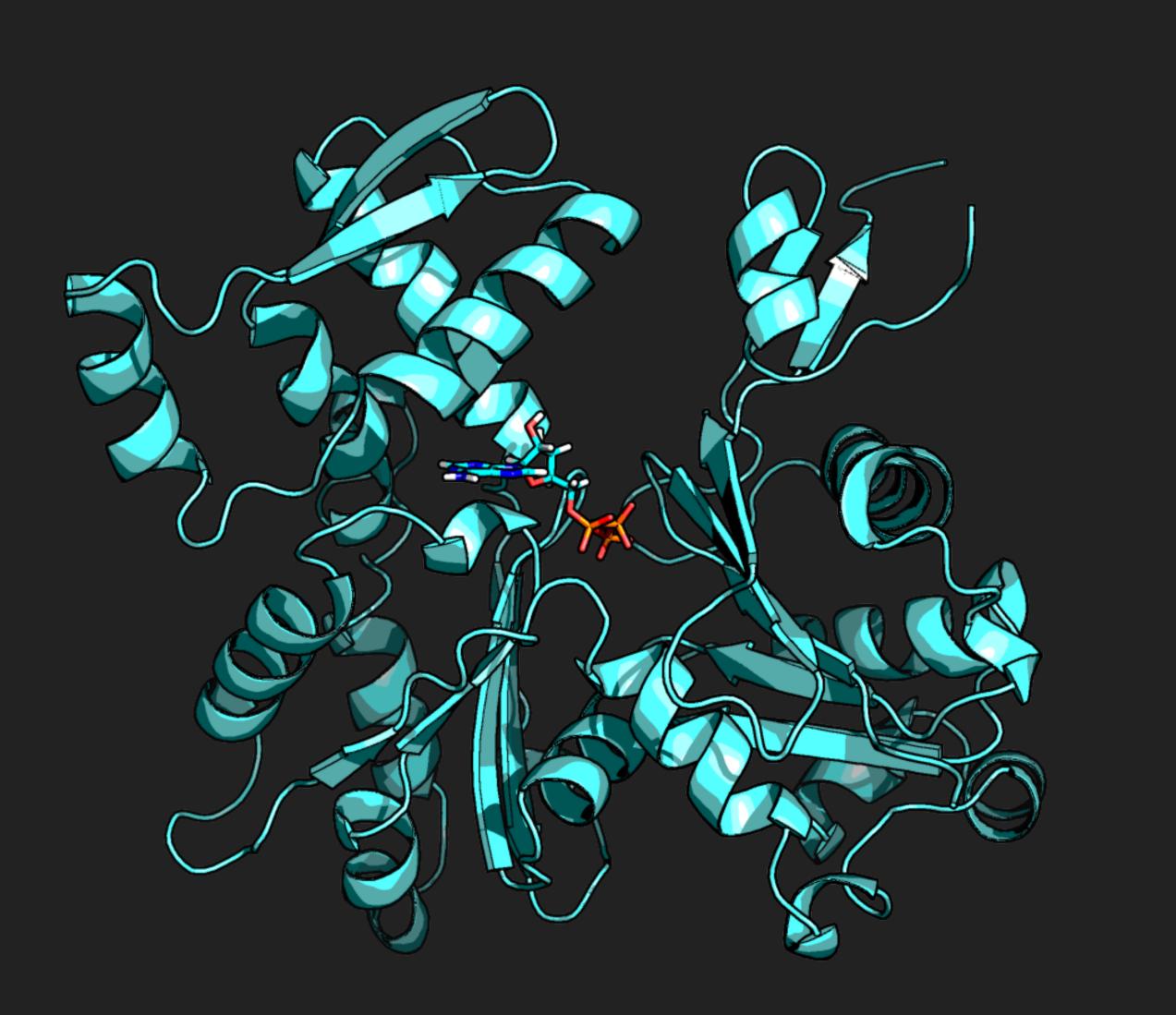
DEPARTMENT OF BIOMEDICINE, UNIVERSITY OF BERGEN, NORWAY FACULTY OF BIOCHEMISTRY AND MOLECULAR MEDICINE, UNIVERSITY OF OULU, FINLAND



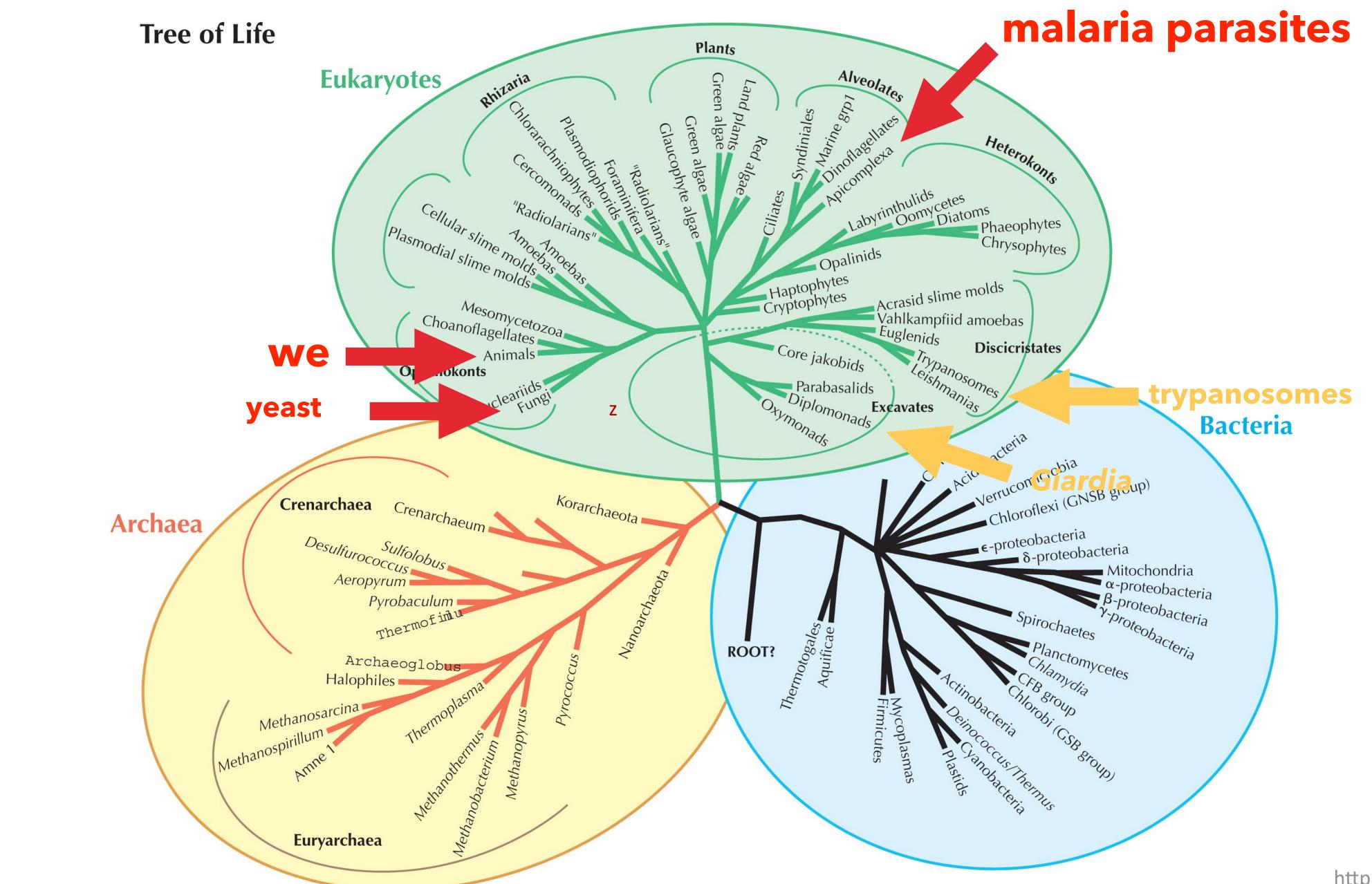








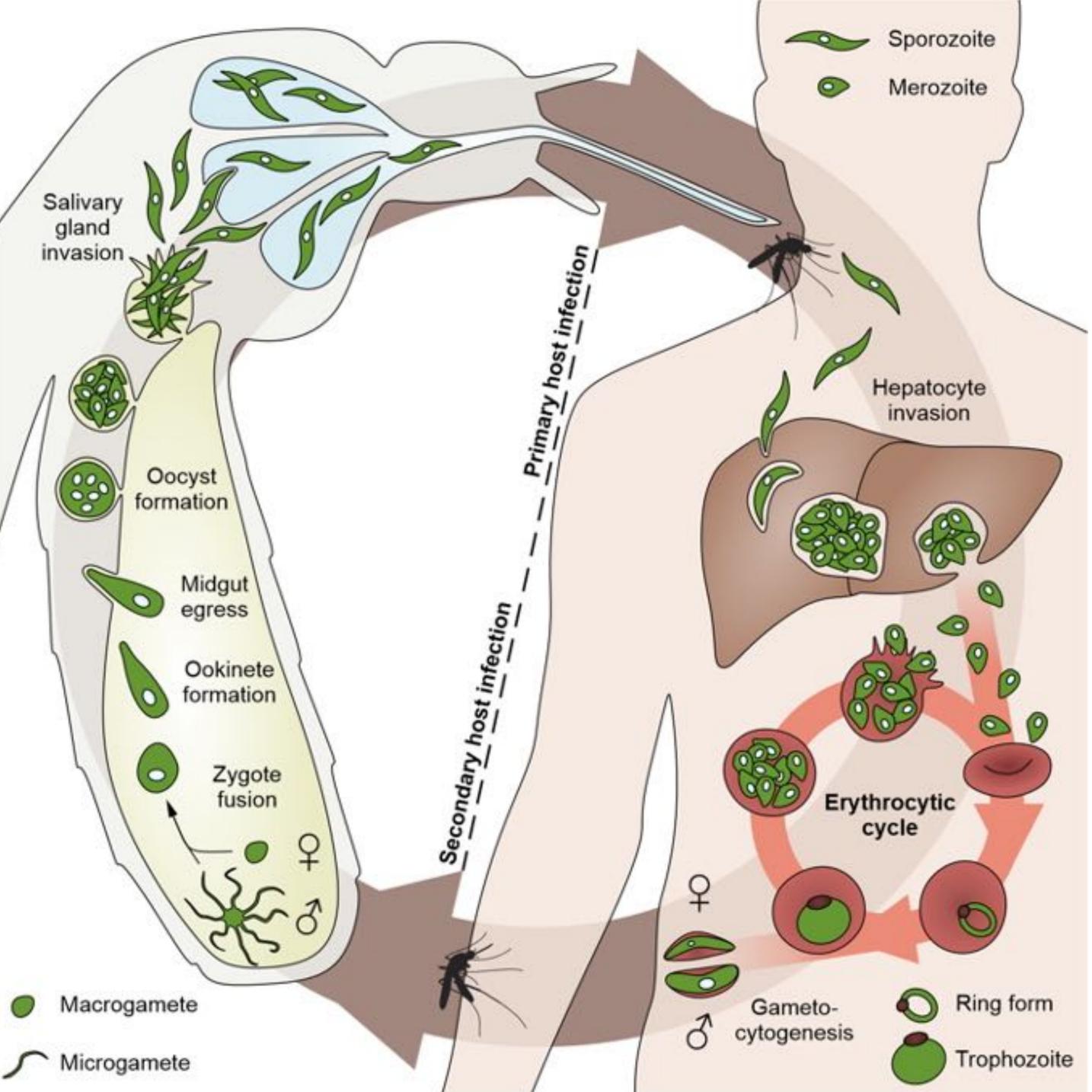
### CHOICE OF MODEL SYSTEM

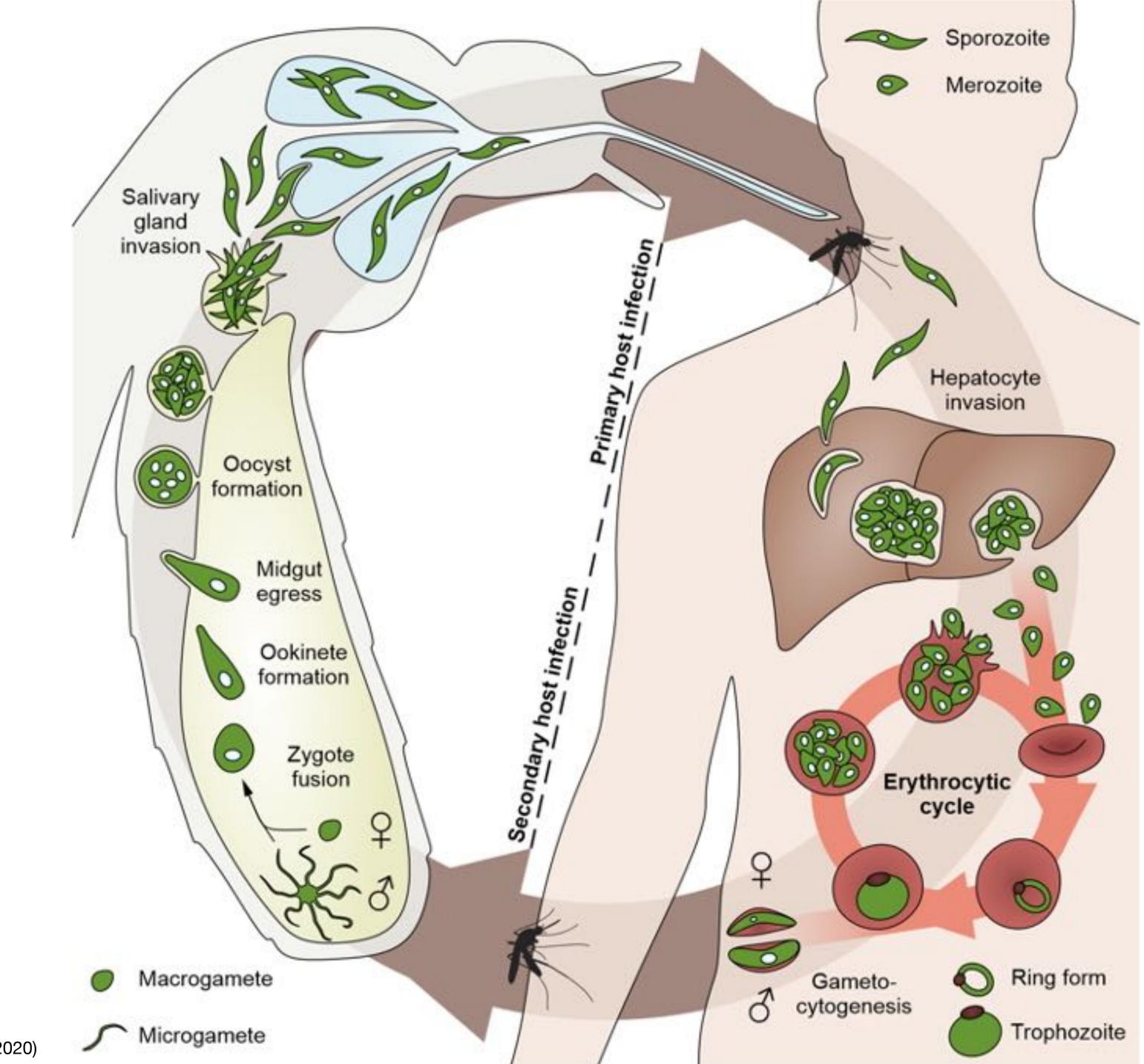


#### http://evolution-textbook.org



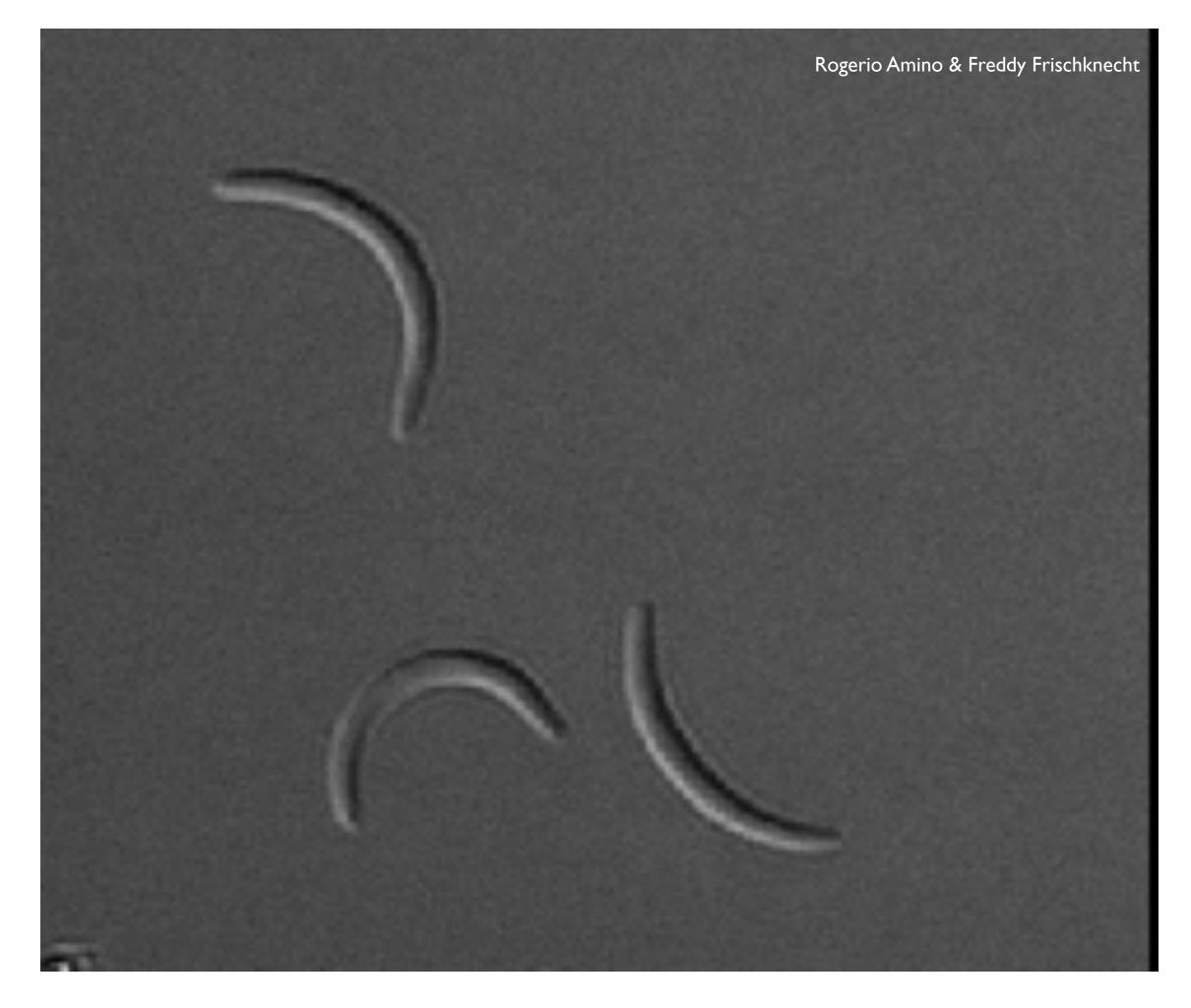
### THE PARASITE LIFE CYCLE



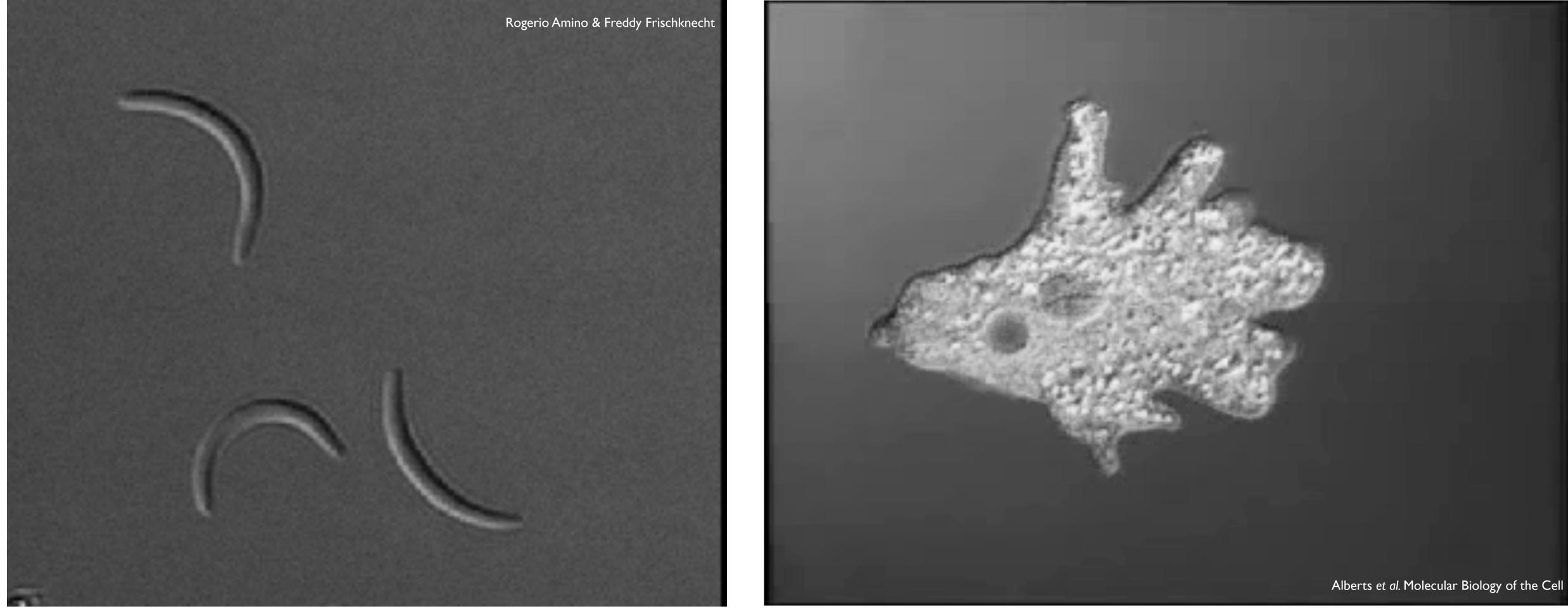


Ábris Bendes (2020)

### APICOMPLEXAN GLIDING MOTILITY

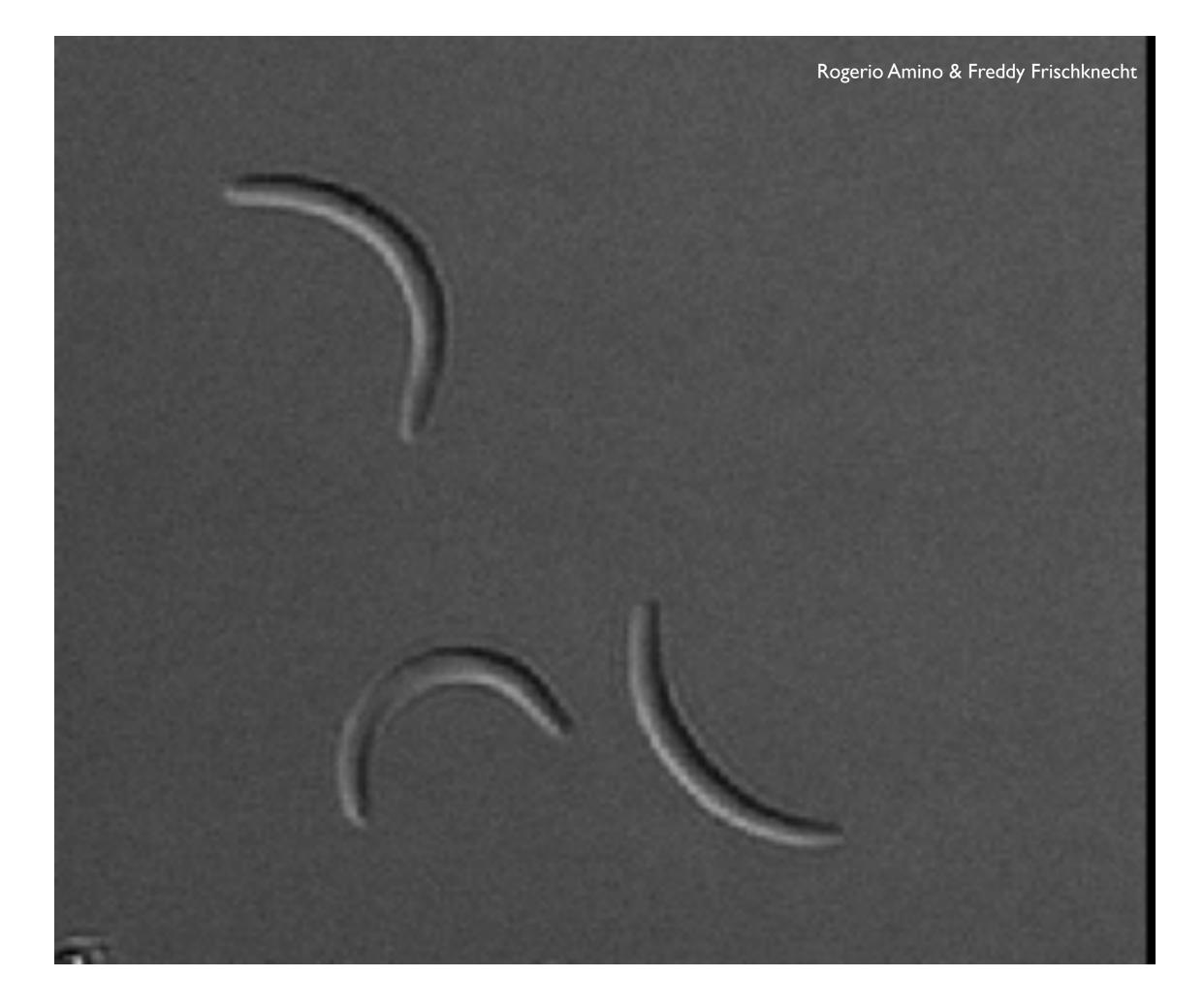


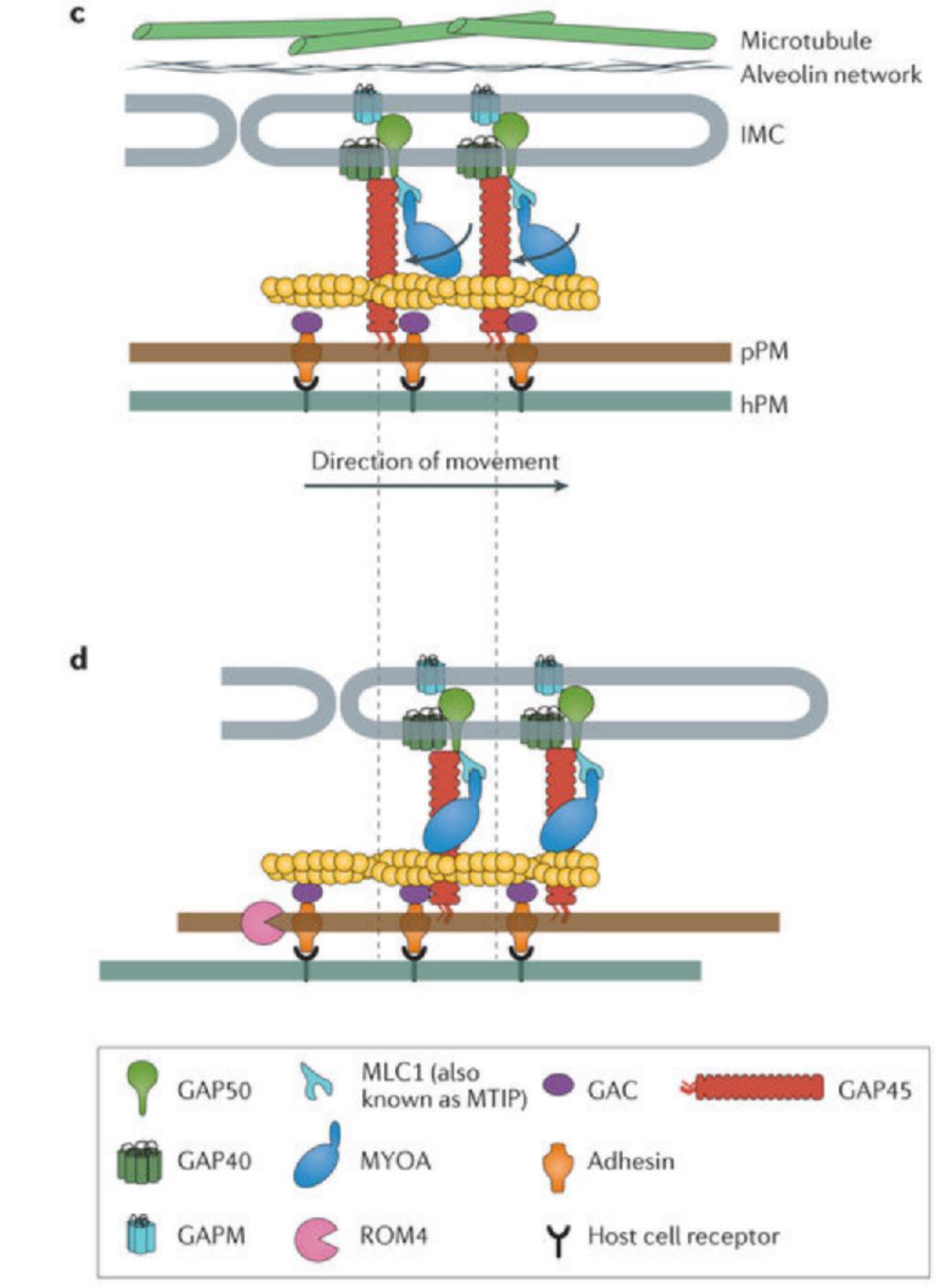
### **APICOMPLEXAN GLIDING MOTILITY**





### APICOMPLEXAN GLIDING MOTILITY





Frénal et al. (2017) Nat Rev Microbiol

# **AIMS OF OUR WORK**

- to understand the mechanism of actin polymerization in the malaria parasite and how actin and its regulatory components have evolved in eukaryotes in general
- to draw a complete molecular picture of the entire glideosome
- > to understand how force is generated and transmitted in gliding motility
- to evaluate the glideosome and actin regulatory components as drug and vaccine targets for fighting malaria





# WHAT IS SO SPECIAL ABOUT PLASMODIUM ACTINS?

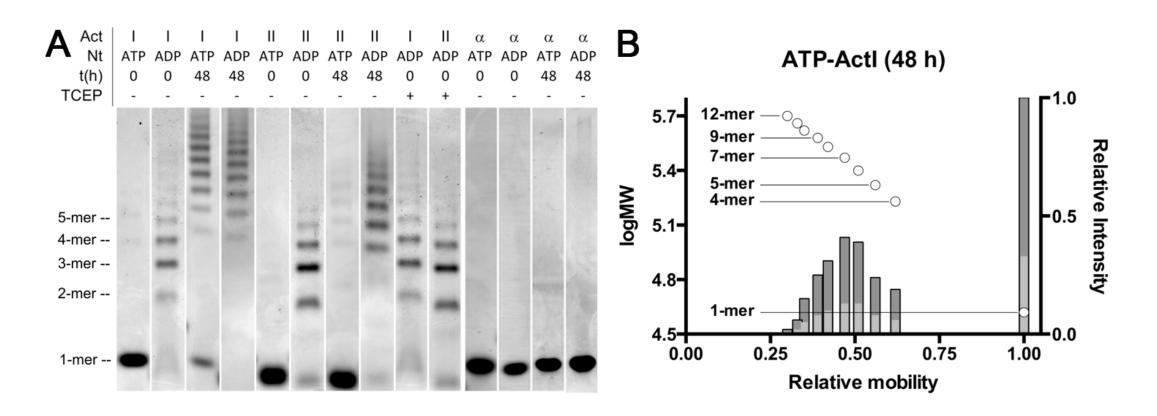
- Iow sequence identity between them and canonical actins and also between the two isoforms (<80%)
- form mainly very short filaments (~100 nm)
- ATP hydrolysis/phosphate release pathway and its link to polymerization seems different
- polymerization not governed by ionic strength in the same way as canonical actins
- stable dimers are a significant species

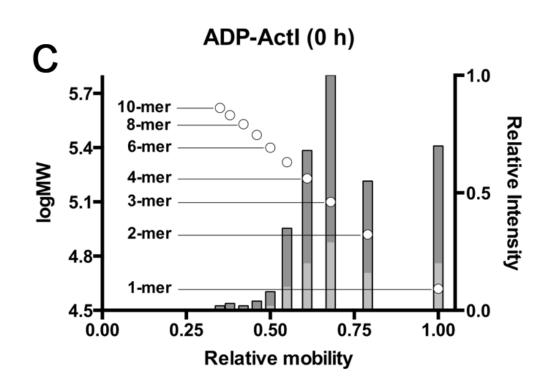


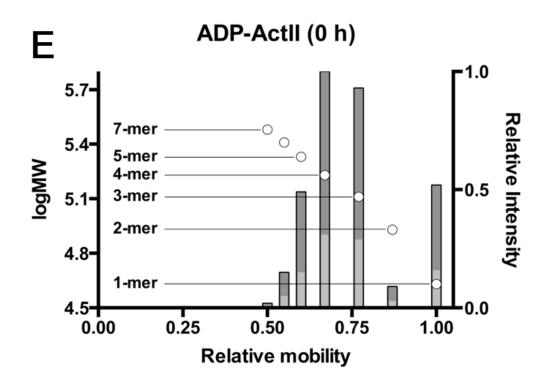
## **MECHANISM OF ACTIN POLYMERIZATION – IS IT DIFFERENT?**

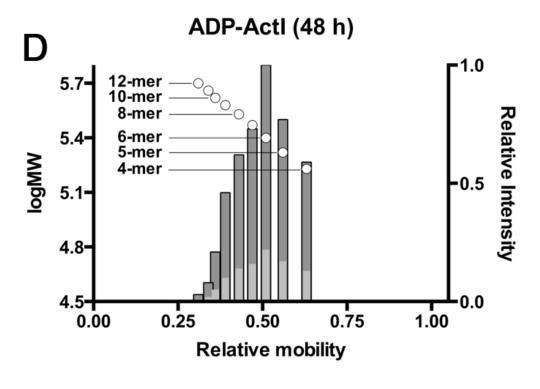


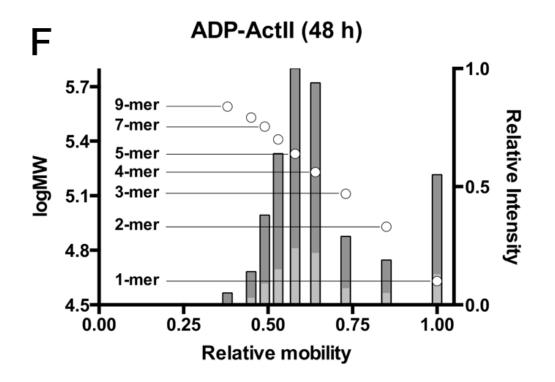
## ATP HYDROLYSIS IN PLASMODIUM ACTIN



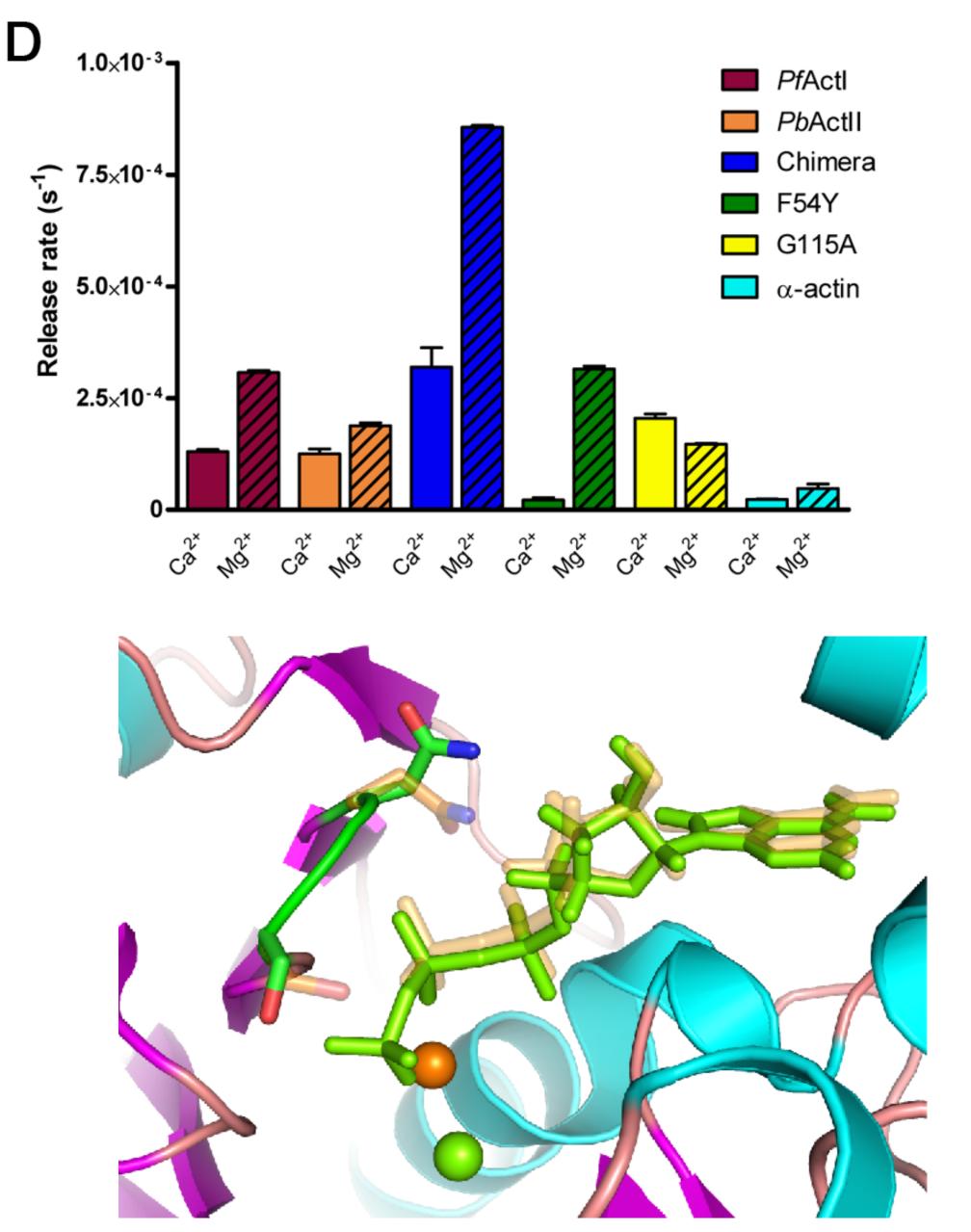




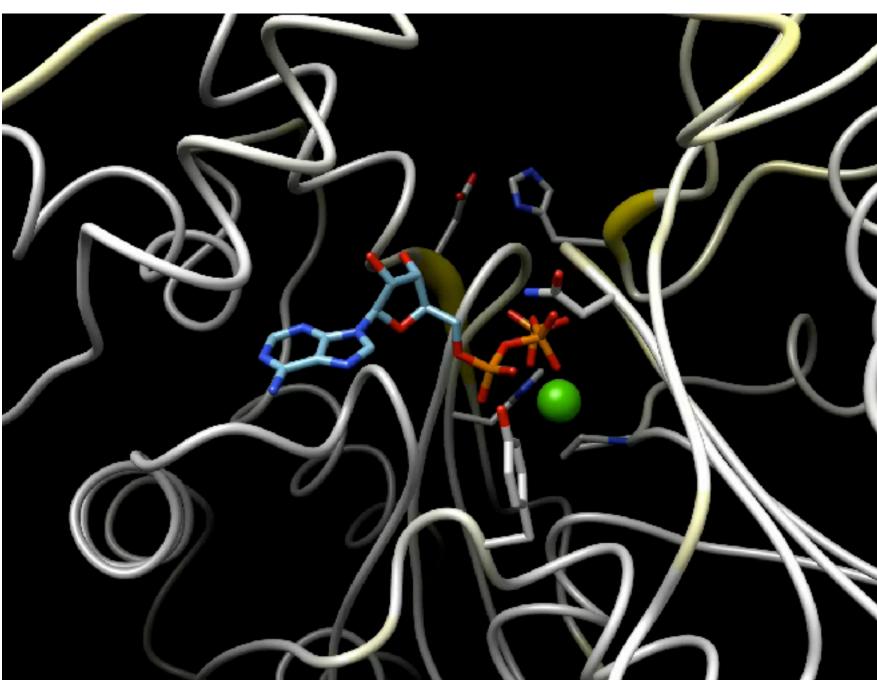


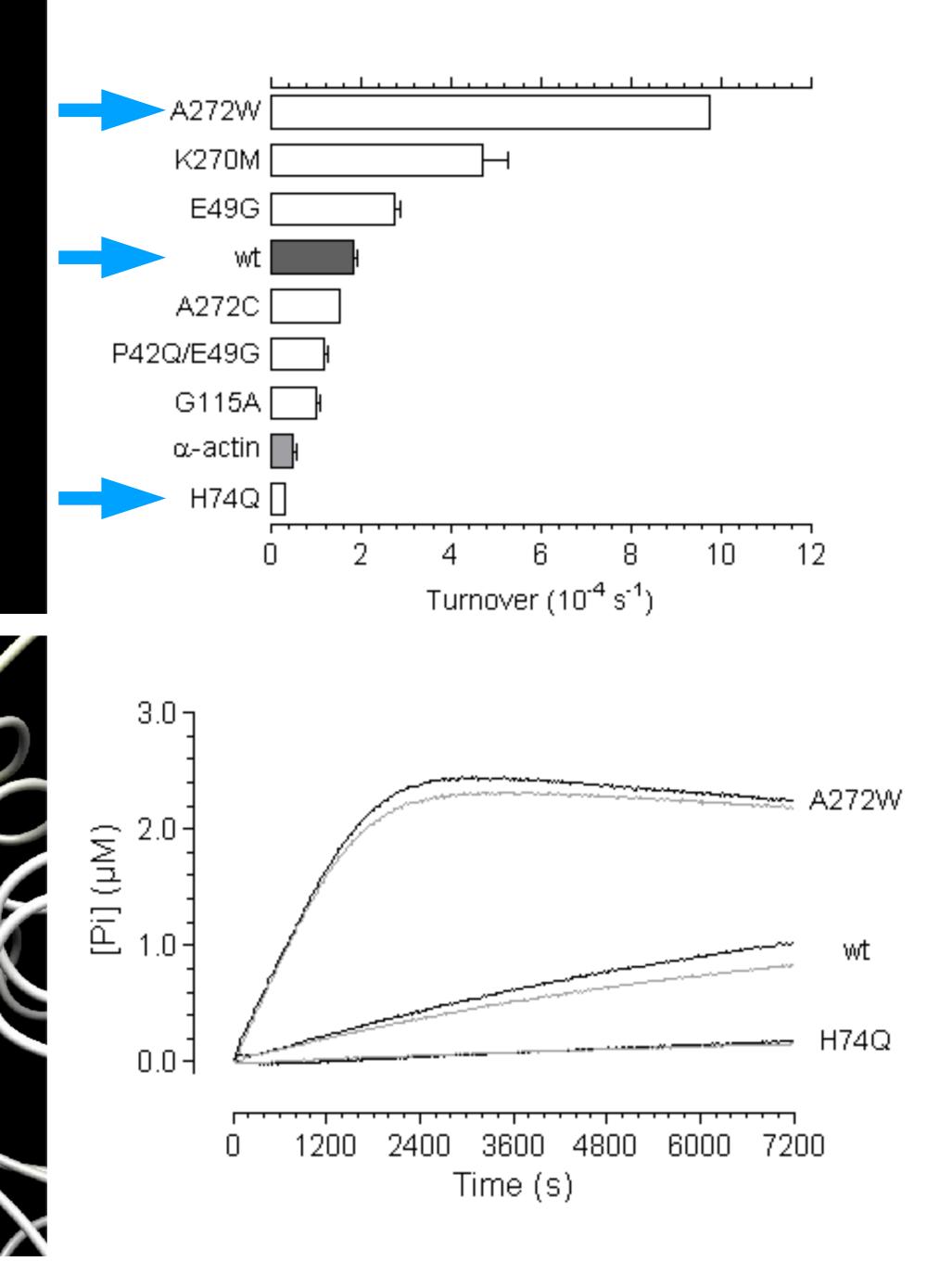


Vahokoski et al. (2014) PLOS Pathog

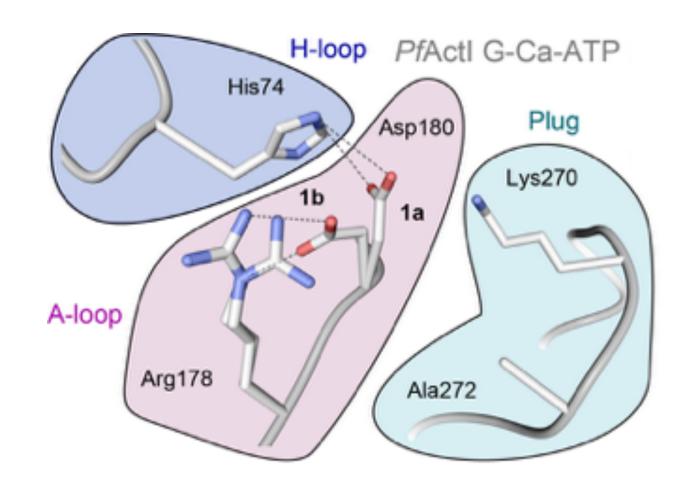


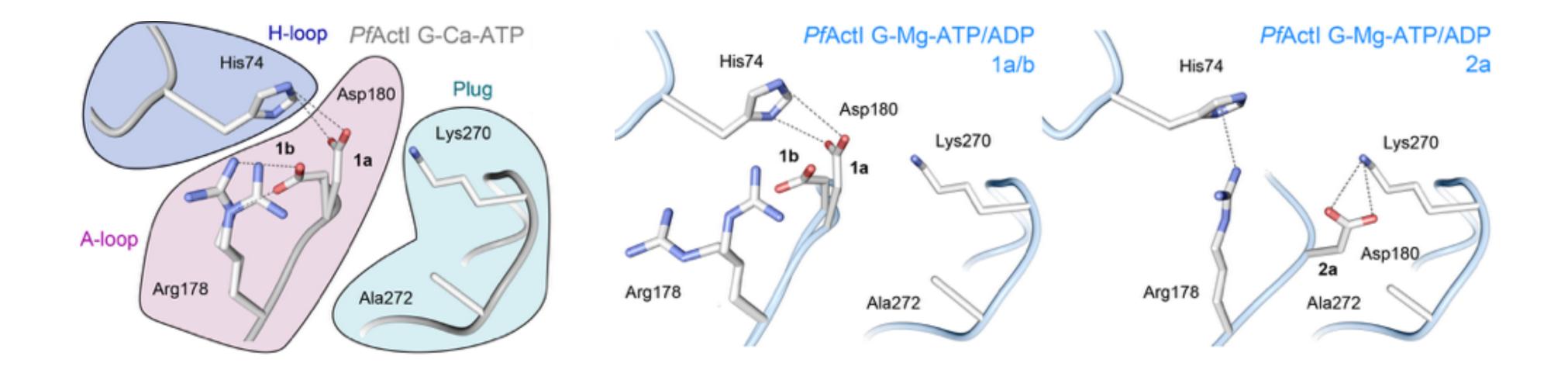


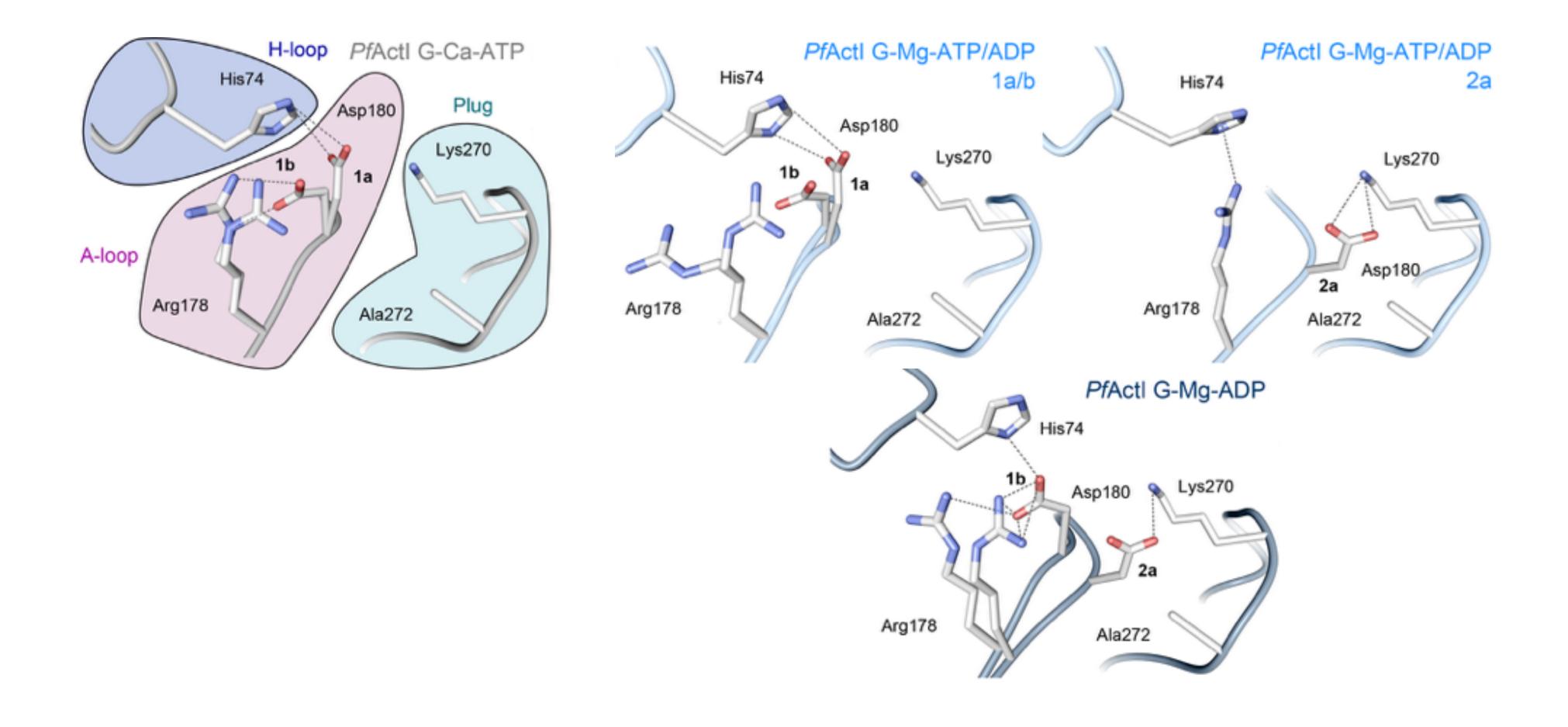


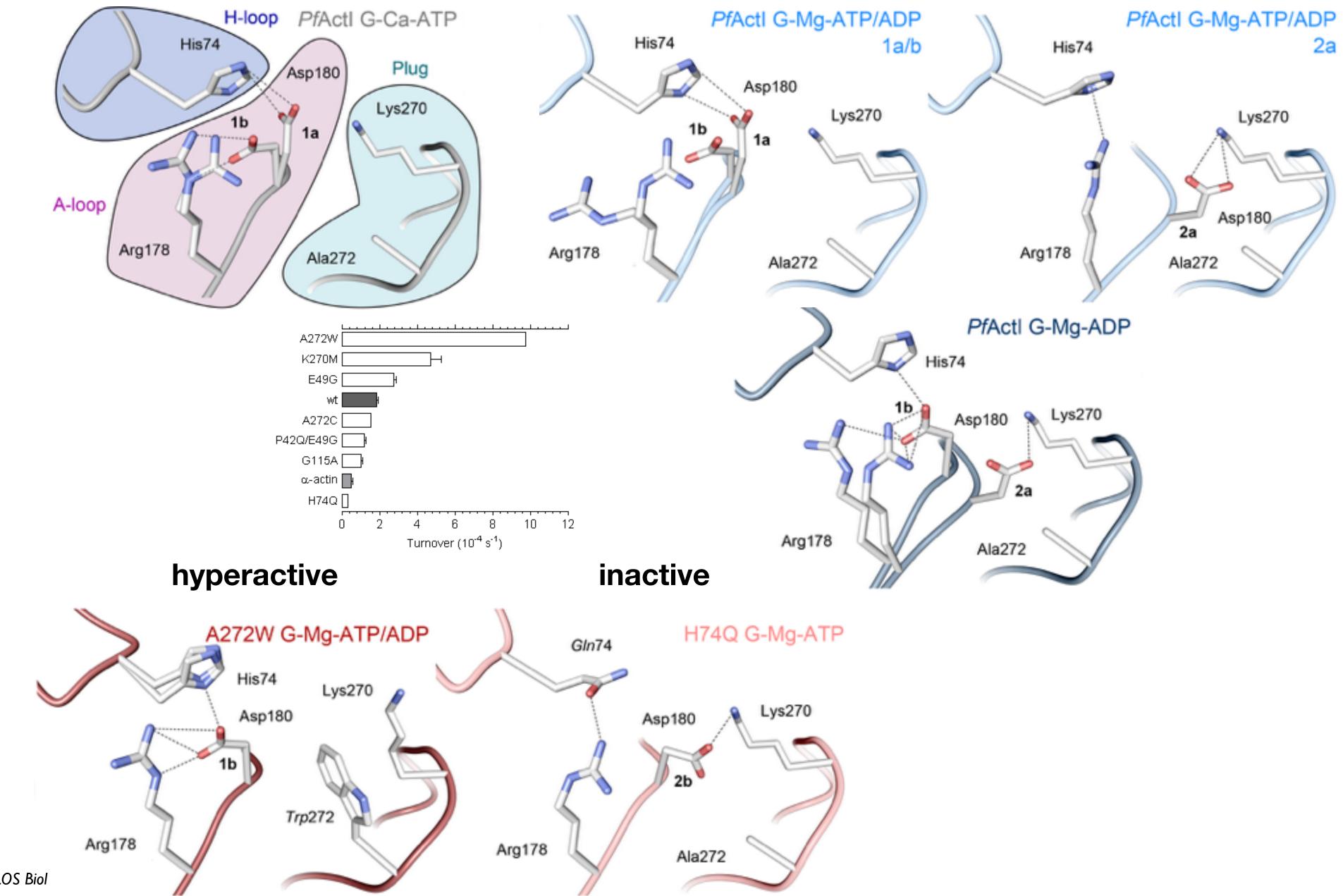


### THE A-LOOP IS LINKED TO PHOSPHATE RELEASE

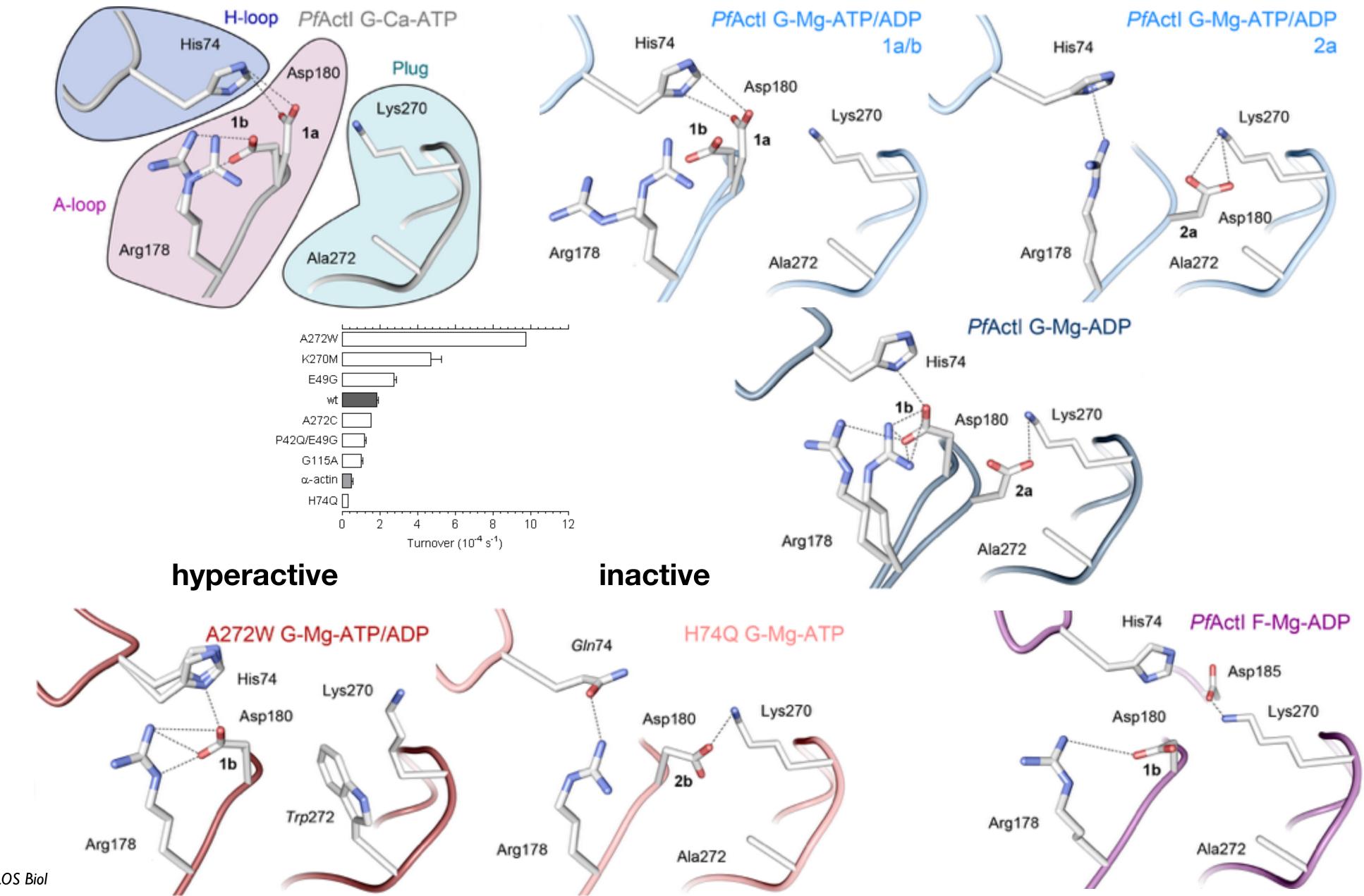








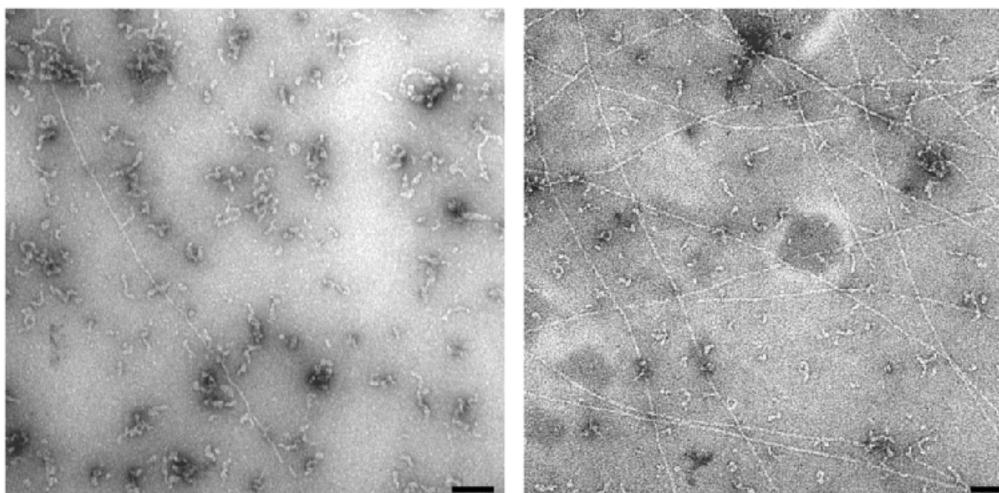
Kumpula et al. (2019) PLOS Biol

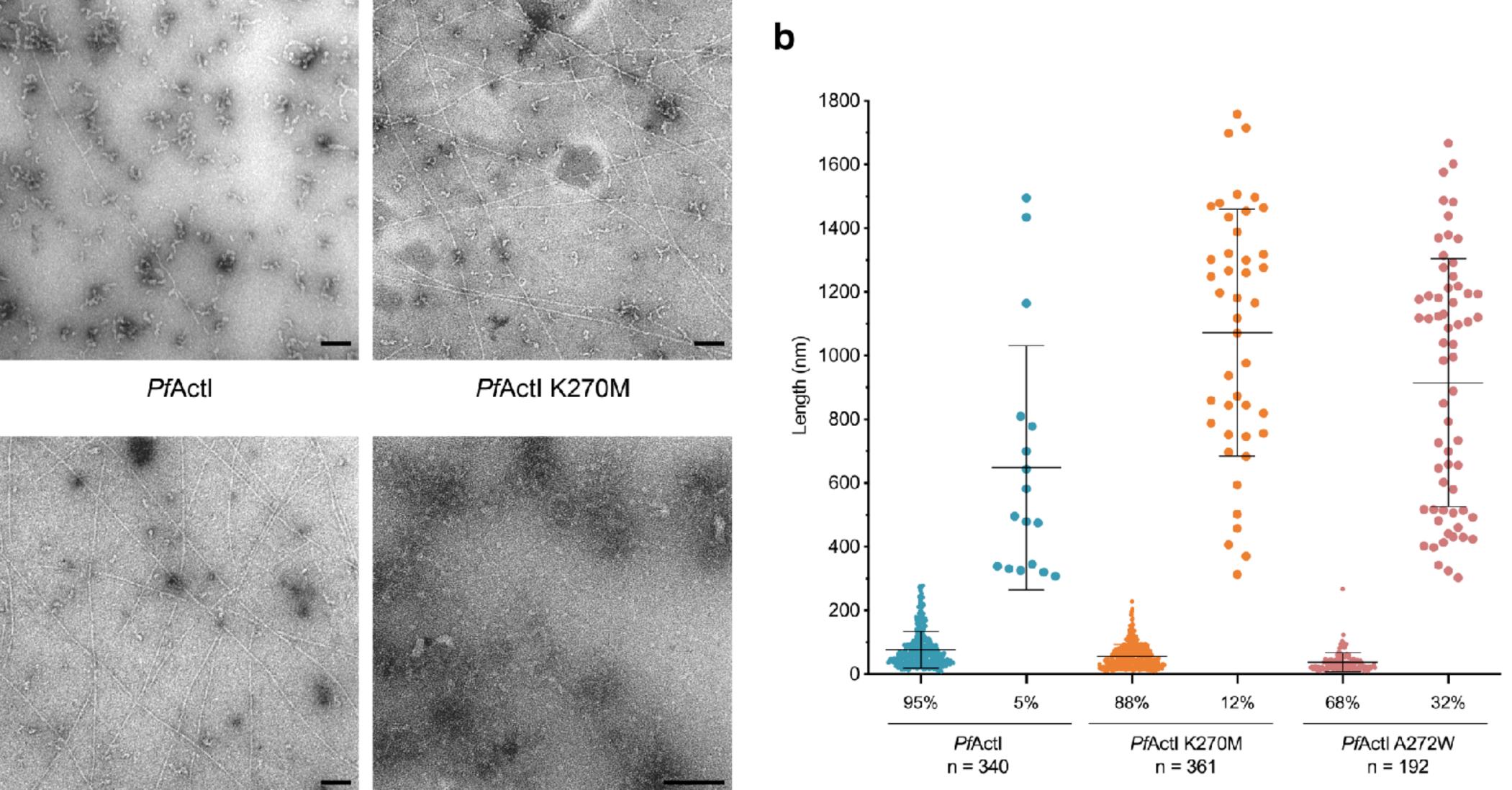


Kumpula et al. (2019) PLOS Biol

## THE A-LOOP GOVERNS ALSO FILAMENT LENGTH

а





PfActl A272W

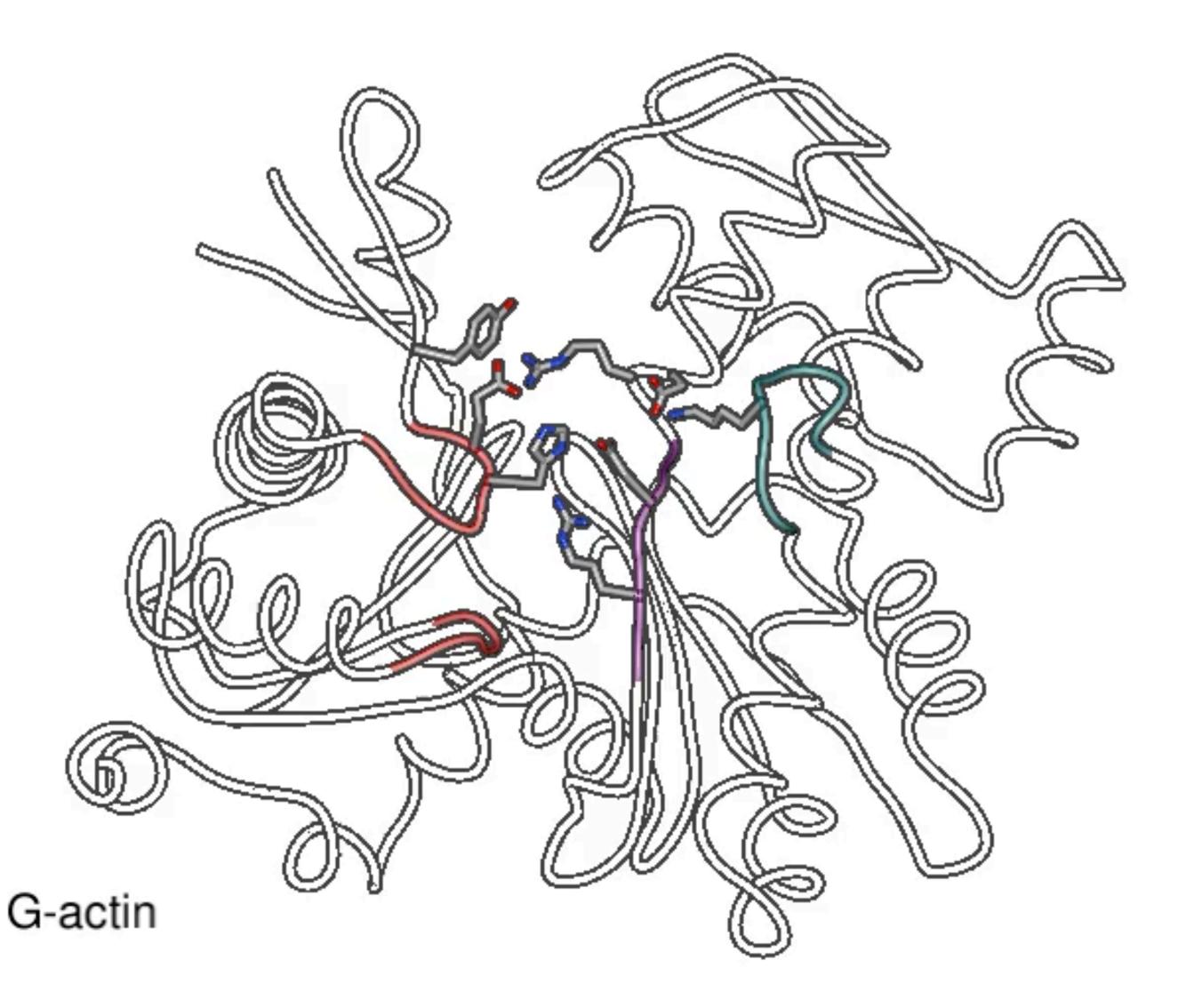
PfActl H74Q

Kumpula et al. (2019) PLOS Biol



## **OVERVIEW OF THE STRUCTURAL TRANSITIONS FROM G TO F FORM**

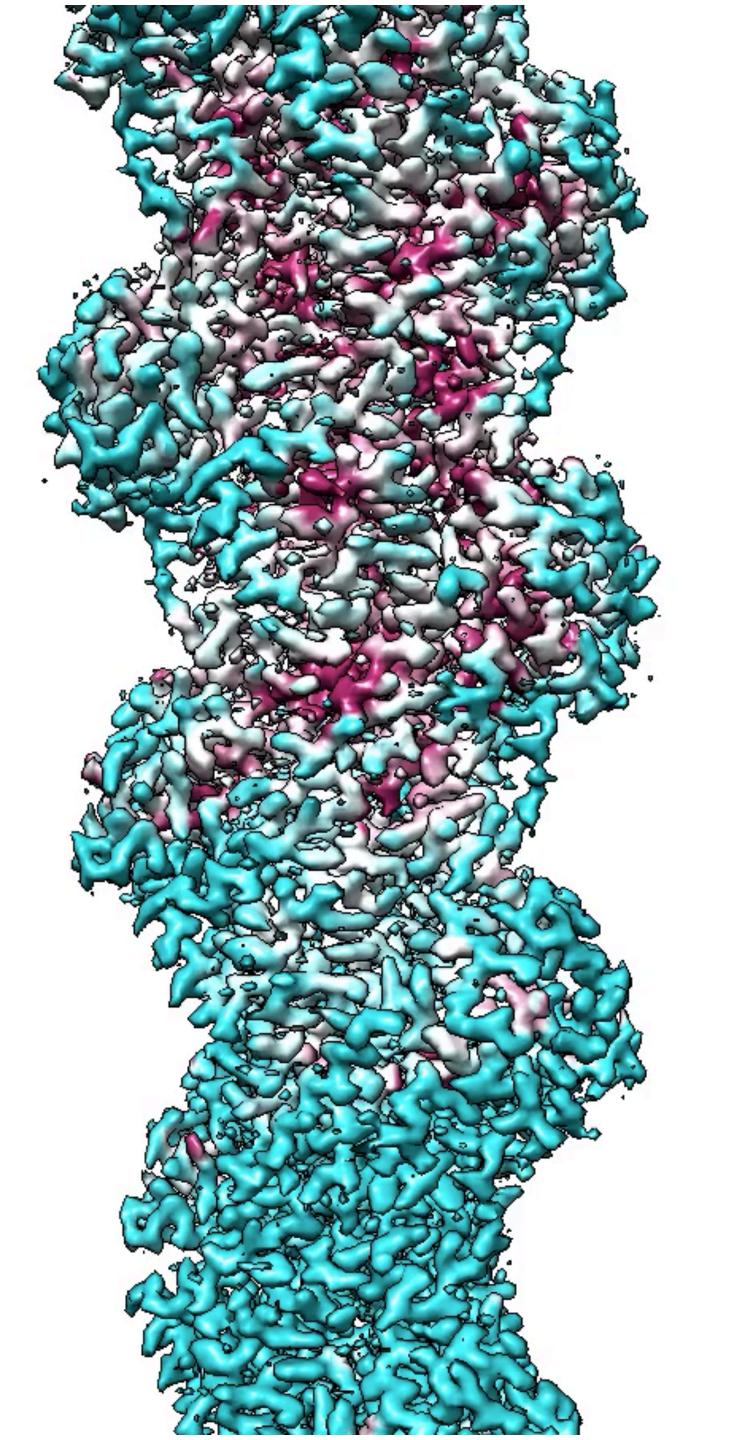
- the movement of the A-loop has not been seen in canonical actins
- it may still happen but not be as favorable due to two small amino acid differences:
  - Lys270 -> Met in canonical actins
  - His74 is methylated in muscle actins
- thus, the A-loop mobility increases the rate of phosphate release and decreases filament stability

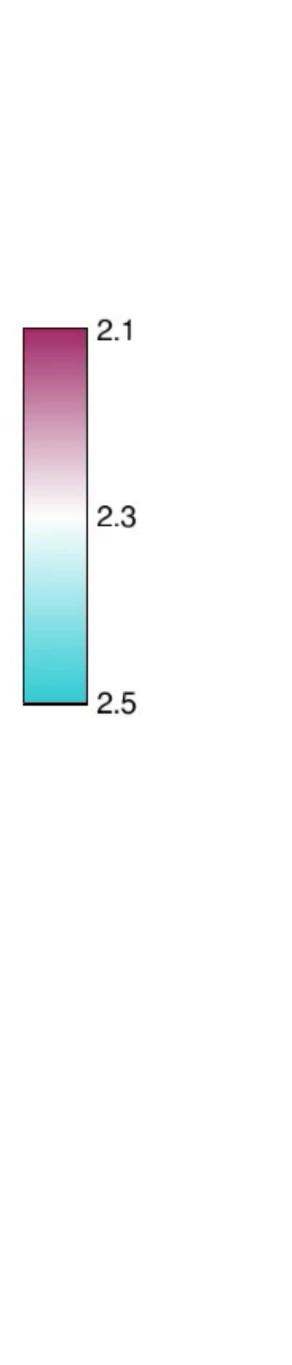




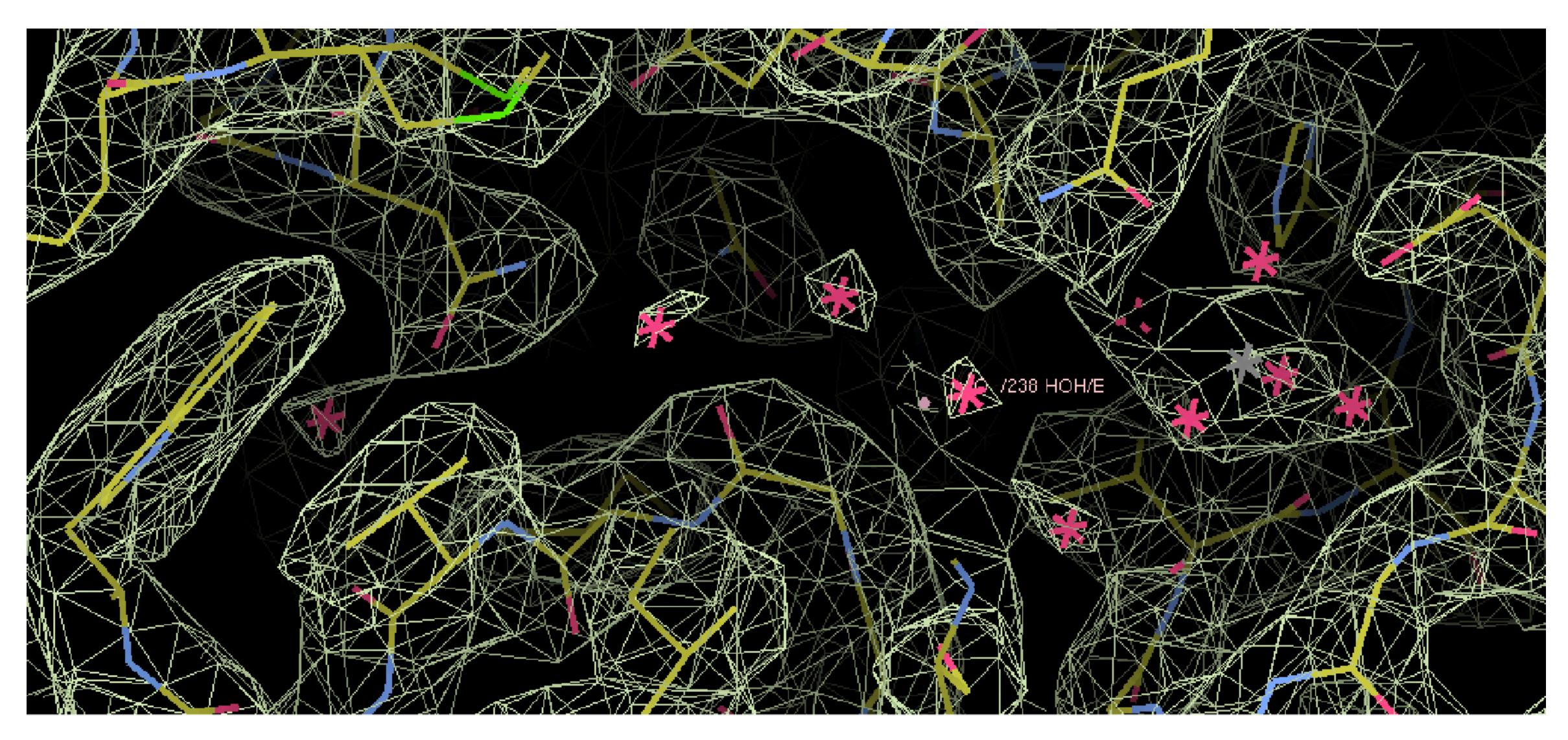
## THAT WE LEARNED FROM PHOTONS WHAT ABOUT ELECTRONS?

Vahokoski et al. (2020) bioRxiv

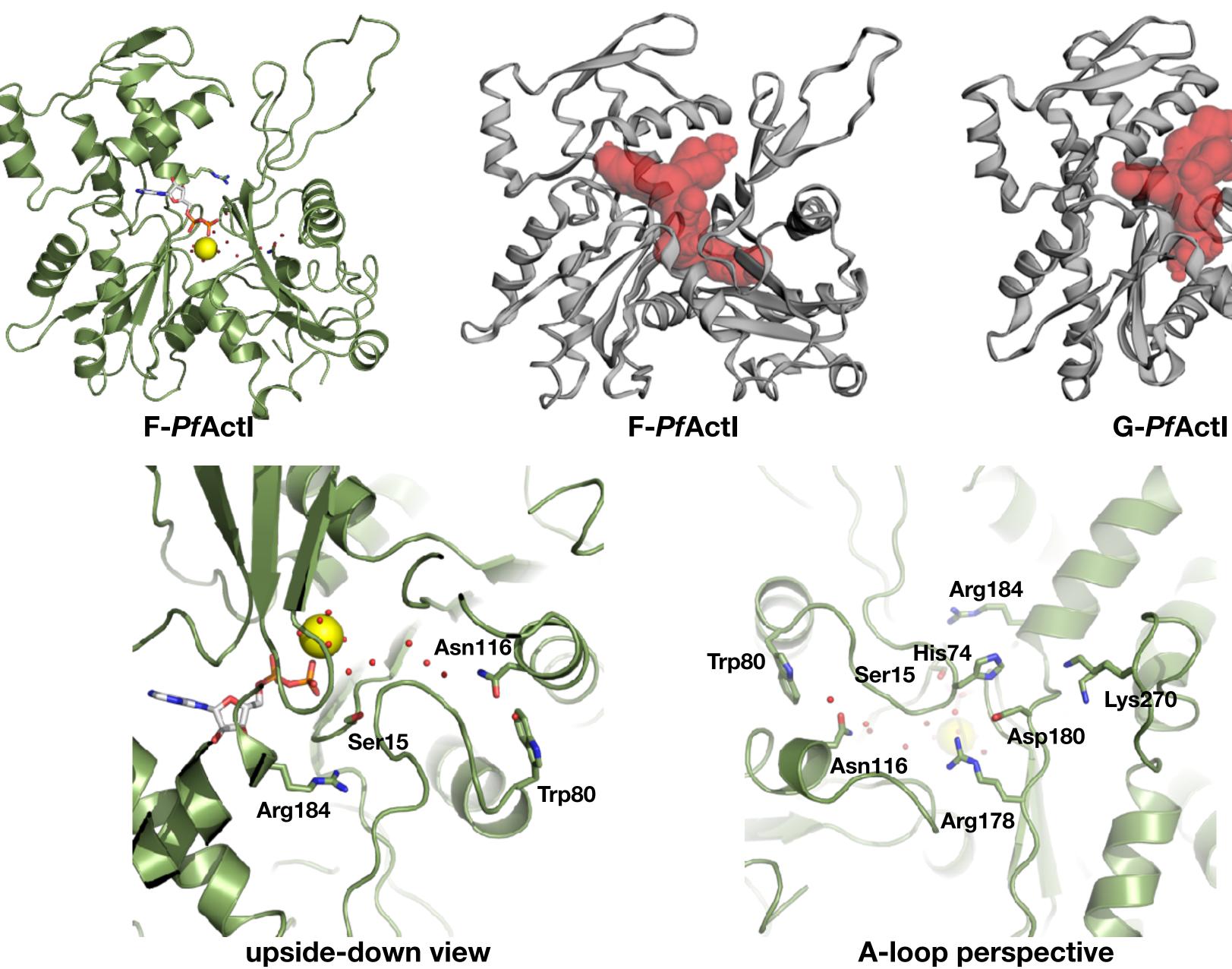


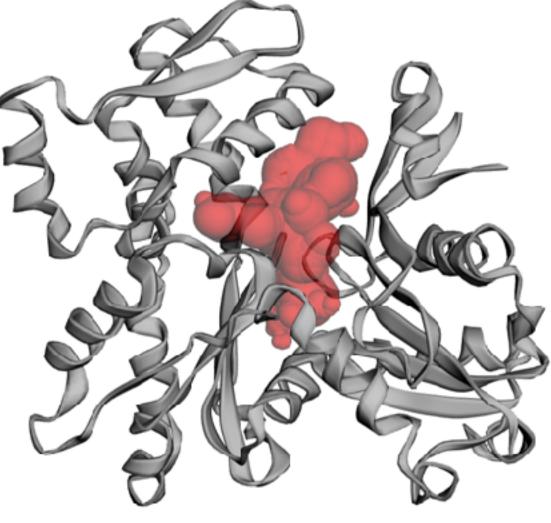


### WHAT CAN WE SEE IN ACTIN? How far can we push the resolution?



## **ALTERNATIVE PATH OF THE LEAVING PO4?**





## SUMMARY

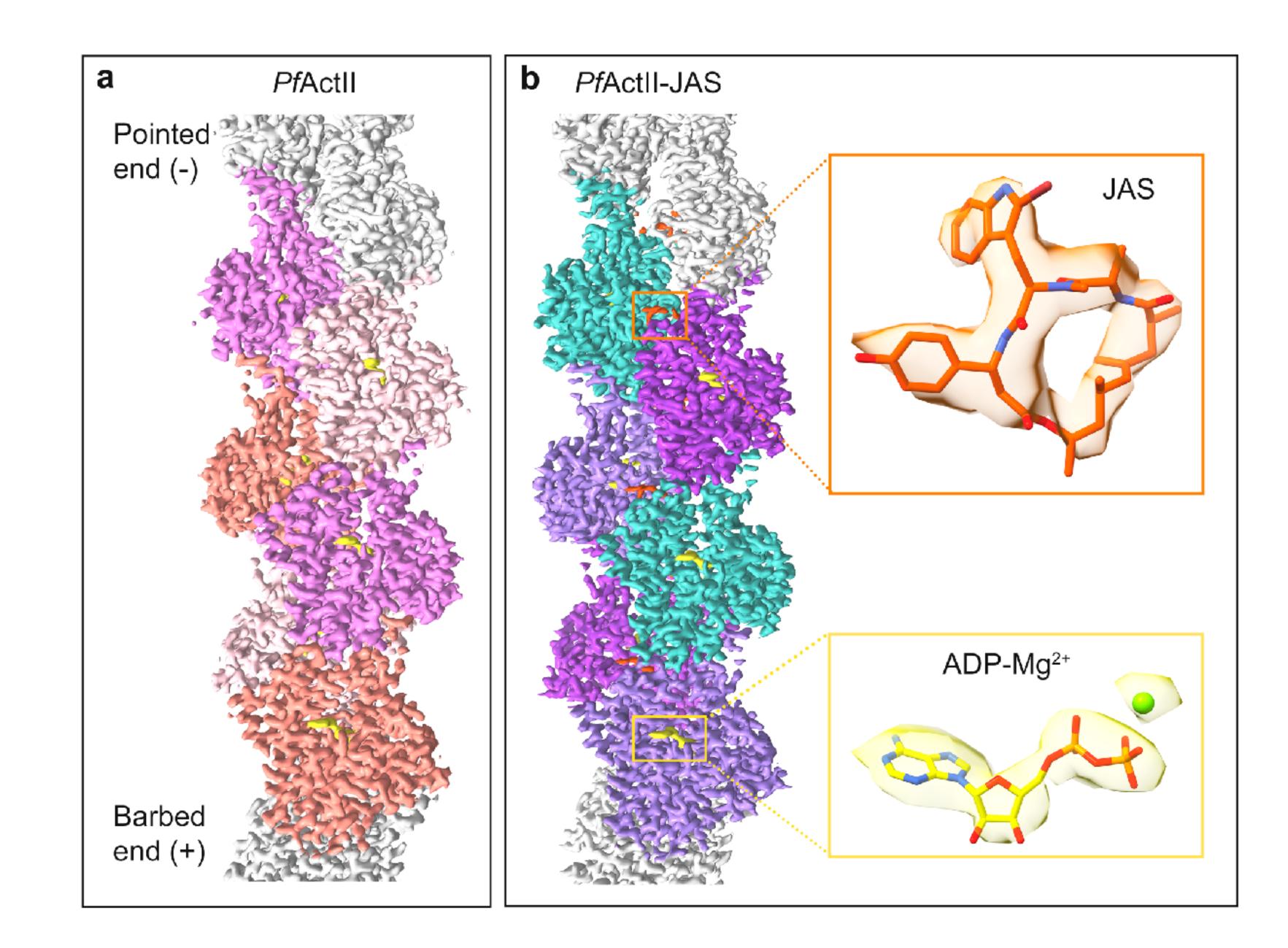
- PfActl polymerization follows the classical nucleation-elongation pathway
- a high fragmentation rate leads to short filament length
- subtle amino acid changes and the lack of methylation of His74 allow the A-loop to act as a fast switch between stable and unstable filament conditions (enforcing and weakening the ID-OD connections)
- a similar mechanism of fragmentation is possible in canonical actins, albeit at a lower frequency
- a channel gated by Asn116 and Trp80 could be the path for the leaving phosphate





#### actin II

- mutagenesis in the active site and the possible
  phosphate leaving routes
- different nucleotide states
- trap leaving phosphate?
- even higher resolution
- look into the catalytic mechanism in more detail



J P



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Ezeogo Obaji

post doc

#### Devaki Lasiwa Ph.D. student



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Henni Piirainen post doc



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Big thanks also to:

#### Esa-Pekka Kumpula



Juha Vahokoski post doc (2011-2019)



#### Andrea Lopez Ph.D. student



Bergen

#### Ju Xu lab engineer



Bergen

**Ábris Bendes** 







Isa Pires Ph.D. student

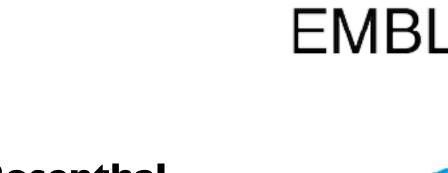


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### THANK YOU! QUESTIONS???









